

UNITED STATES DEPARTMENT OF THE INTERIOR, Fred A. Seaton, *Secretary*
FISH AND WILDLIFE SERVICE, Arnie J. Suomela, *Commissioner*

MORPHOLOGY OF THE WHITE SHRIMP

Penaeus setiferus (Linnaeus 1758)

BY JOSEPH H. YOUNG



FISHERY BULLETIN 145

From Fishery Bulletin of the Fish and Wildlife Service

VOLUME 59

UNITED STATES GOVERNMENT PRINTING OFFICE • WASHINGTON • 1959

For sale by the Superintendent of Documents, U. S. Government Printing Office
Washington 25, D. C. . Price \$1

The Library of Congress has cataloged Fishery Bulletin 145 as follows:

Young, Joseph Hardie, 1918—

Morphology of the white shrimp *Penaeus setiferus* (Linnaeus 1758) Washington, U. S. Govt. Print. Off., 1959.

iv, 168 p. illus. 30 cm. (U. S. Fish and Wildlife Service. Fishery bulletin 145)

"From Fishery bulletin of the Fish and Wildlife Service, volume 59."

Bibliography: p. 160-169.

1. Shrimps. i. Title. (Series: U. S. Fish and Wildlife Service. Fishery bulletin, v. 59, no. 145)

SH11.A25 vol. 59, no. 145 595.3843 59-60272

Library of Congress

The series, Fishery Bulletin of the Fish and Wildlife Service, is cataloged as follows:

U. S. *Fish and Wildlife Service.*

Fishery bulletin. v. 1—
Washington, U. S. Govt. Print. Off., 1881-19

v. in illus., maps (part fold.) 23-28 cm.

Some vols. issued in the congressional series as Senate or House documents.

Bulletins composing v. 47— also numbered 1—

Title varies: v. 1-49, Bulletin.

Vols. 1-49 issued by Bureau of Fisheries (called Fish Commission, v. 1-23)

1. Fisheries—U. S. 2. Fish-culture—U. S. i. Title.

SH11.A25 639.206173 9-35239 rev 2*

Library of Congress r55e¾1

CONTENTS

	Page
Introduction.....	1
<i>Penaeus</i> as comparative material.....	2
I. Skeletal and muscle systems.....	3
A. Protocephalon.....	3
1. Eyestalks.....	7
2. Antennules.....	17
3. Antennae.....	30
4. Labrum.....	43
B. Gnathothorax.....	47
1. Mandibles.....	60
2. Paragnatha.....	65
3. Maxillae.....	66
a. First maxillae.....	66
b. Second maxillae.....	69
4. Maxillipeds.....	72
a. First maxillipeds.....	72
b. Second maxillipeds.....	74
c. Third maxillipeds.....	81
5. Pereiopods.....	87
a. First pereiopods.....	87
b. Fifth pereiopods.....	94
C. Abdomen.....	103
1. Pleopods.....	115
2. Tail fan.....	125
II. Nervous system.....	132
A. Nerves of Supraesophageal ganglion and tritocerebrum.....	135
B. Ventral nerve cord.....	139
III. Circulatory system.....	140
A. Heart and pericardium.....	140
B. Blood vessels of the body.....	140
C. Appendicular blood vessels.....	148
D. Venous system.....	149
E. Respiratory system.....	149
IV. Digestive system.....	150
A. Foregut.....	150
B. Midgut.....	154
C. Hindgut.....	154
V. Excretory system.....	155
VI. Reproductive system.....	155
Literature cited.....	160
Bibliography.....	162

ABSTRACT

In this illustrated morphology of the commercially important white shrimp of the Gulf of Mexico the muscle, nervous, circulatory, excretory, reproductive, and respiratory systems are compared with those of the blue crab, *Callinectes*, a European crawfish *Astacus*, the "coon-stripe" shrimp, *Pandalus*, and other **decapod** crustaceans. The major portion of the comparative work deals with the muscles, **since** the muscle systems of a few Decapoda have been reported in much greater detail **than** other systems.

The comparative studies of muscles and nerves indicates that the Penaeidae represent a generalized anatomical condition in the Crustacea Decapoda, thus verifying the systematic research in this area. Evidently the Penaeidae are relatively close to the decapod stem in the Malacostraca. The generalized condition of Penaeidae is shown again and again by the repetition of functional muscle units. The same units have become simplified in the higher decapods, having been lost, presumably, by the fusion of separate parts. Adhering to the morphological principle that the nerves tend to retain their ancient innervations despite coalescence of parts, shifting of muscle origins, etc., and can therefore be considered as morphologically conservative, the nerves to the repeating muscle units of *Penaeus* are found to have kept their innervations to the same muscles, now fused, in higher Decapoda. The comparative morphology of decapod nerves and circulatory elements is treated only to the extent that research on these systems in other decapods has been published.

Several structures are found in *Penaeus setiferus* which have not been reported previously in the literature. A fibrous circulatory element, the capillary arbor, penetrates the distal optic ganglia. One or more hard concretions embedded in the substance of the antennal portion of the excretory gland are described. Although two pairs of muscles are associated with the labrum of Insecta, muscles have not been described in the labrum of Crustacea. The labrum of *Penaeus* has at least 12 pairs of muscles. A structure, the hindgut gland, is found in the anterior part of the sixth abdominal segment lying dorsal to the rectum. Its function is unknown. Some of the blood vessels of the heavily vascularized branchiostegal region of the carapace run parallel to the margin of the carapace, suggesting "growth rings" by their appearance.

MORPHOLOGY OF THE WHITE SHRIMP

PENAEUS SETIFERUS (LINNAEUS 1758)

By JOSEPH H. YOUNG, *Department of Zoology, Tulane University*

Penaeus setiferus, the white shrimp of the Gulf of Mexico, represents an important component of the commercial shrimp catch throughout the northern, western, and southern margins of that body of water. With the rise in importance of the commercial shrimp industry to the economy of the Gulf States in the past two decades, information about the life history, morphology, physiology, and behavior of the edible shrimps has become necessary, and even critical, to the continued well-being of the industry.

We know very little about the diet of the penaeid shrimps, how far they travel in search of food and mates, or what constitute barriers to their activities. Our knowledge of digestion, of nervous and glandular control of the processes of molting, reproduction, and coloration is limited for the most part to distant relatives of the Penaeidae. Despite numerous efforts (Müller 1863; Kishinouye 1900; Hudinaga 1935, 1942; Pearson 1939; Johnson and Fielding 1956), the stages of the life cycle of penaeids are not certainly known and consequently we have very little exact ecological information about the larvae. Opinions vary on the life span of adult penaeid shrimps. Whether the adult females reproduce once and then die at the end of a year or reproduce more than once a season, or again, live for 2 years and reproduce each season is not known. The answers to such questions will have a profound effect on the future conduct of the commercial shrimp industry.

This study sets forth in detail the anatomy of *Penaeus setiferus*. Anatomical work as such does not answer the problems suggested above, but as a map to the explorer, anatomy provides the experimental biologist with a guide to the "lay of the land," thereby facilitating his progress. And as

a map, the value of an anatomical study to its users hinges upon its accuracy and clarity. To these ends all efforts were bent, the illustrations are large and the structures shaded to give a sense of depth. The anatomical parts are labelled with full words, rather than disguised with cryptographic abbreviations listed below the illustrations or hidden in the text. The plates are defined as diagrams whereby important features may be emphasized by artwork without sacrificing accuracy.

P. setiferus is an omnivorous scavenger dwelling on or near mud-sand bottoms in the littoral zone. The animal is probably restricted to the euphotic stratum in which the light intensities are fairly high. The shrimps are gregarious and move about in large schools. The character of the bottom is probably important to the white shrimp. Evidence from various sources suggests that the animals make use of mud or mud and sand by burrowing into this soft substrate for protection during molting and possibly for other purposes. The white shrimp is a powerful swimmer and capable of migrating great distances. These and other aspects of the life habits of *P. setiferus* have been considered in terms of functional anatomy in the present study.

Several people have made significant contributions to the progress of this study. Credit for initiating the project belongs perhaps most of all to Albert Collier, United States Fish and Wildlife Service, Galveston, Tex. His unfailing cooperation, together with that of T. J. Costello of the same laboratory, contributed greatly to the completion of the anatomical work. Help and encouragement has also come from Dr. Fred R. Cagle, Dr. Royal D. Suttikus, and Dr. George H. Penn, all of the Department of Zoology, Tulane University. Percy Viosca, Louisiana Wildlife and Fisheries Commission; Charles Dawson, of

NOTE.—Approved for publication April 22, 1958. Fishery Bulletin 145. Report of research done under contract between Bureau of Commercial Fisheries and Tulane University (Saltontall-Kennedy Act).

the Bears Bluff Laboratories, Wadmalaw Island, S. C.; and Dr. Milton Fingerman, Department of Zoology, Tulane University, have all made useful comments about various aspects of the work. I am indebted to Prof. L. B. Holthuis, Rijksmuseum van Natuurlijke Historie, Leiden; Dr. Andre Mayrat, Laboratoire de Zoologie de l'Ecole Normale Supérieure, Paris; and Jerome E. Stein, Texas A. and M. College, Galveston, Tex., for helpful criticism of certain parts of the study.

Heartfelt thanks go to the project artist, Raymond Bollinger of New Orleans, La., for the high quality of the plates making up the anatomical study of the white shrimp. Of the 136 figures in this work, all but 10 were finished by Mr. Bollinger from tracings made by the investigator.

METHODS

The anatomical study of *Penaeus setiferus* was made for the most part on white shrimps purchased alive from bait shrimp fishermen. The animals were fixed in Zenker's fluid, dehydrated to 70 percent ethyl alcohol and there stored. Despite difficulties in its use in the field, Zenker's fluid was found to have several advantages over formalin. Zenker's fluid softens or removes the calcareous deposits and leaves the cuticle in a condition similar to thick cellophane. This mixture quickly penetrates to and fixes the internal organs, and in so doing prevents internal maceration caused by the post-mortem enzymatic activity of the hepatopancreas. Formalin-fixed material is useless for the study of internal organs. The fixative greatly hardens the cuticle and the external muscles and fails to penetrate to the internal organs.

Dissections were performed under a stereomicroscope. Dissecting needles which were sharpened to fine points in mixtures of strong nitric acid and ethyl alcohol were employed. Locations of muscles and attachments, and other skeletal details were studied on specimens of white shrimps cleared in strong alkali and stained with Van Gieson's Triple Stain (Curtis' Modification). The outlines of whole structures were used as templates within which muscles and other organs were sketched in layers on tracing paper as the dissections progressed. The tracings were transferred to drawing papers on a light box. The drawings were finished in ink and carbon pencil.

PENAEUS AS COMPARATIVE MATERIAL

For purposes of comparative morphology, the Penaeidae enjoy a unique position. Evolution has brought them down to us in an apparently generalized decapod condition. Naturally, we must view with caution any attempt to force a given structure or organ into the generalized category, for all extant animals must be highly specialized in specific instances, if unspecialized in others, for life today. Furthermore, the so-called generalized structure may be the superficialities of a well-hidden specialization. Bearing in mind, then, that there may be no such thing as a generalized structure in a modern species, the comparative morphologist can proceed to draw homologies between structures which look alike on phylogenetic and ontogenetic grounds. Information shedding light on the evolution of the decapod crustaceans will be advanced here wherever supporting studies are available.

Unfortunately, very few complete anatomical works exist on decapod crustaceans, and none of these complete works deal with members of the Penaeidae. The present work will have reference to the studies of many workers, but in particular to the extensive studies of Berkeley (1928) on the "coon stripe" shrimp, *Pandalus danae* Stimpson 1857, of Schmidt (1915) and Keim (1915) on *Astacus astacus* (Linn. 1758), and of Cochran (1935) on the blue crab, *Callinectes sapidus* Rathbun, 1896. Other, frequently important, studies will be alluded to in the applicable sections. The greatest handicap to the establishment of a comparative morphology of the Decapoda is our ignorance of the details of the nervous system, the innervations of muscles and other organs. Despite widespread morphological change such as shifting of muscle origins, coalescence of metameres, and other distortions to the primitive body plan, structures tend to retain their ancient innervations. A muscle may move from one segment to another in the evolution of a group. The same nerve will usually continue to innervate it. For the nervous system is morphologically conservative and thus the most informative element in the historical study of metazoan structure. Until a substantial groundwork of neurological facts exists on the decapod crustaceans, homologies between many of the less obvious structures will rest on weak grounds.

When compared externally with the crustaceans mentioned (figs. 1, 2, 3), *Penaeus setiferus* is most similar to *Pandalus danae*, a caridean prawn, and less like *Astacus astacus*. The relationship with the blue crab is obviously distant. The Penaeidae are classed in the Sub Order Natantia together with the Caridae and the Stenopidae. All of these are relatively strong swimmers with light cuticles. Although *Astacus* displays many similarities in form to *Penaeus*, the crawfish is a reptant form with a heavy cuticle. With the lobster, the crawfish has an extensively developed chela on the first walking leg, whereas the white shrimp bears a small chela similar in size to those on the other chelate legs. The antennal scale and the pleopods of the crawfish are much smaller than those of the white shrimp. The former are large in the white shrimp. In other general details, superficially, the white shrimp and the crawfish are substantially similar in structure.

In general, the present anatomical study indicates that the Penaeidae are relatively generalized decapod crustaceans. Compared with the higher decapods, the penaeids tend to have several parts to accomplish a functional end that is carried out by a single part in a higher representative. Expressing this in terms of phylogeny, the lower decapod has lost fewer structures by the fusion of parts than has the advanced form. Since the present study of *Penaeus* is largely grounded upon earlier work on higher decapods, the process of homologizing the structures tends to be reversed from the phyletic order. Homologies must therefore be drawn from the specific to the general. Among the problems thus raised is the matter of functional nomenclature, in particular, of the muscles.

The historical base for the naming of decapod muscles is, for all practical purposes, the work of Schmidt (1915) on *Astacus*. This investigator employed a system of Latinized functional names, handed down to him by earlier anatomists, for the muscles he encountered in the European crawfish. With minor exceptions Schmidt's nomenclature accurately describes the actions of the muscles of *Astacus*. However, the functional muscle names of Schmidt do not describe the actions of the same muscle having a different function in another form. The investigator is therefore faced with the decision either to transfer to a muscle in another animal the functionally inaccurate name of Schmidt, which will simplify comparisons, or to

rename the muscle in each case in accordance with its specific function, which will tend to compound the existing confusion in morphological nomenclature. With no great pride I have chosen the latter course in the present study. Until such time as a system of nomenclature having universal validity can be devised, the dilemma is inescapable.

Included in this paper is a section of bibliographical references. These items are primarily systematic, morphological, and experimental papers on Crustacea which contain valuable anatomical information used in the preparation of the present study, but not cited specifically. Since workers in many fields have to resort to anatomical studies in the course of their research, the anatomical information is necessarily disguised under titles which reflect the primary objects of their research. The student of crustacean morphology therefore, finds bibliographical compilations of works from diverse sources very helpful. The bibliography is in no sense complete.

I. SKELETAL AND MUSCLE SYSTEMS

The great mass of the shrimp body is comprised of skeleton and, in particular, muscle; accordingly, the bulk of the present anatomical study is devoted to a consideration of these elements. The description is presented in the order of the three natural body regions of the animal, the simple head, or protocephalon, the gnathothorax, and the abdomen. The skeleton falls easily into these divisions. The muscles, of course, do likewise, but not so obviously, since many of them cross skeletal subdivisions for mechanical reasons. In some anatomical works, the arthropod appendages are treated separately, as if these organs were attached to the animal in a kind of evolutionary afterthought in the arthropod line. The appendicular muscles would, indeed, so indicate, but the skeleton, the nervous system, and the innervations of the muscles tell us that the arthropod appendage is an ancient structure. The appendages, then, will be taken up with the tagmata to which they belong.

A. Protocephalon

The protocephalon is that morphologically separable pregnathal group of segments so designated by Snodgrass (1951). This simple head includes, in the order of their occurrence in the adult, the eyes, antennules, antennae, and labrum.

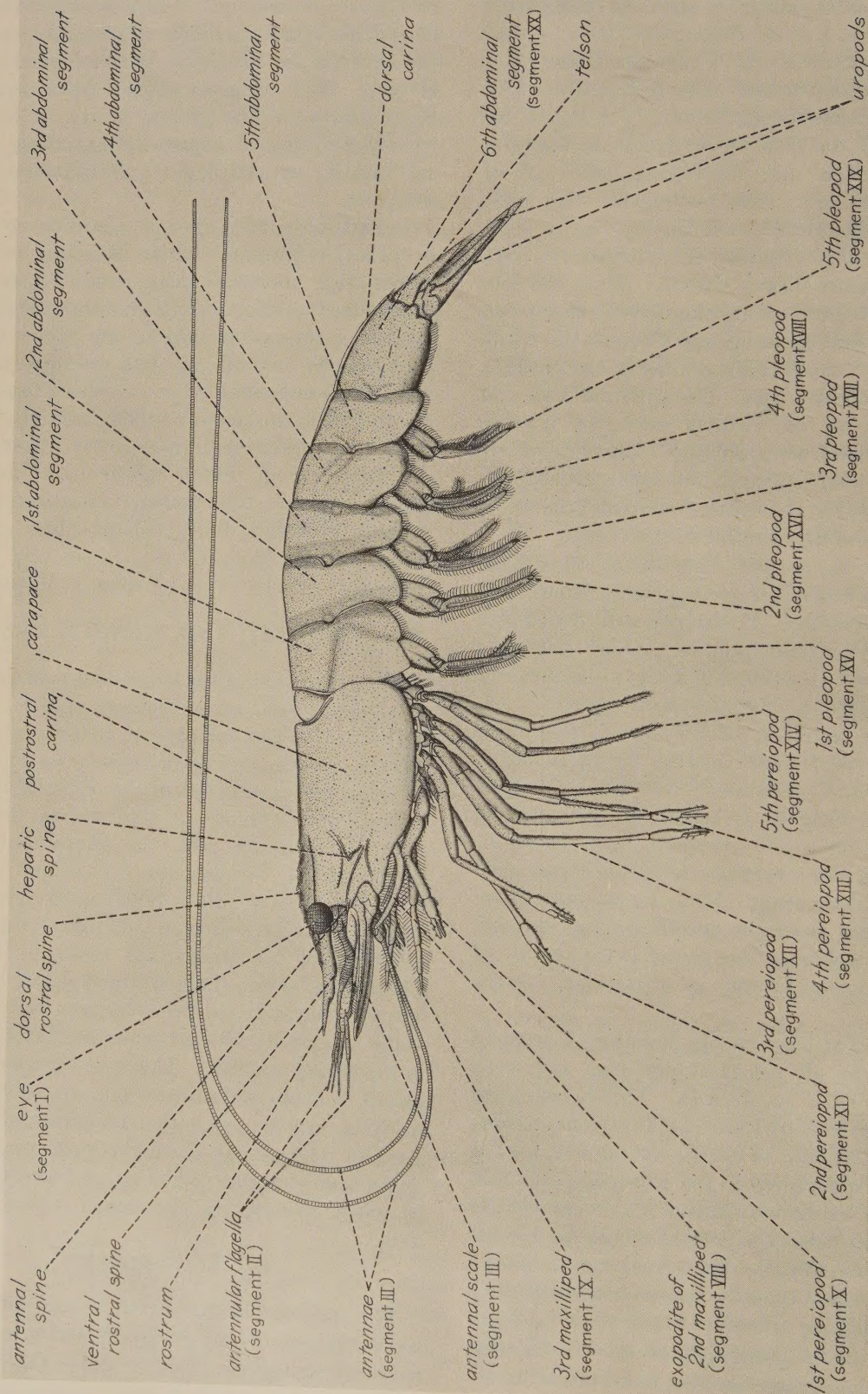


FIGURE 1.—*Penaeus setiferus*. Lateral view of adult female.

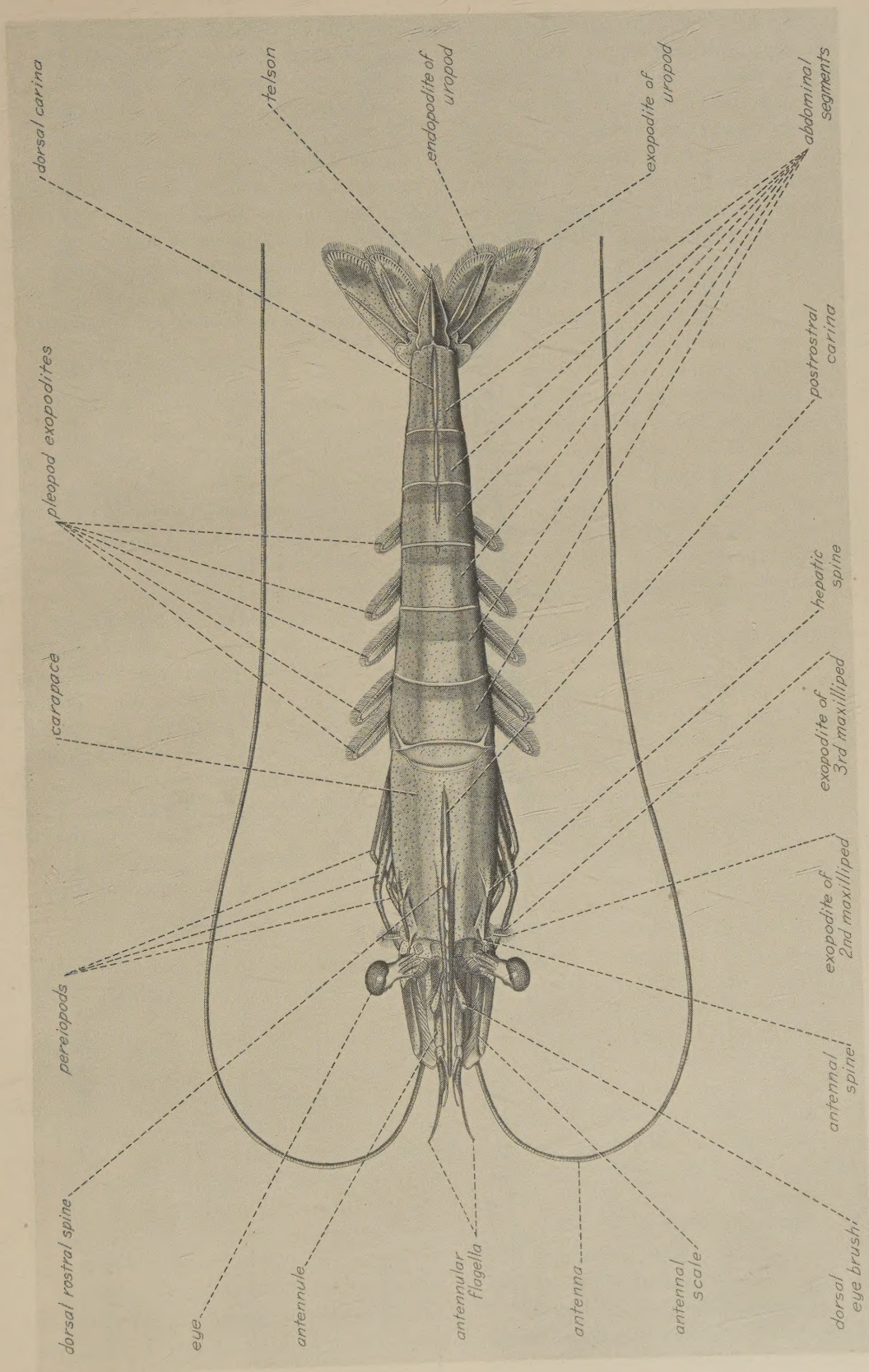


FIGURE 2.—Dorsal view of adult female.

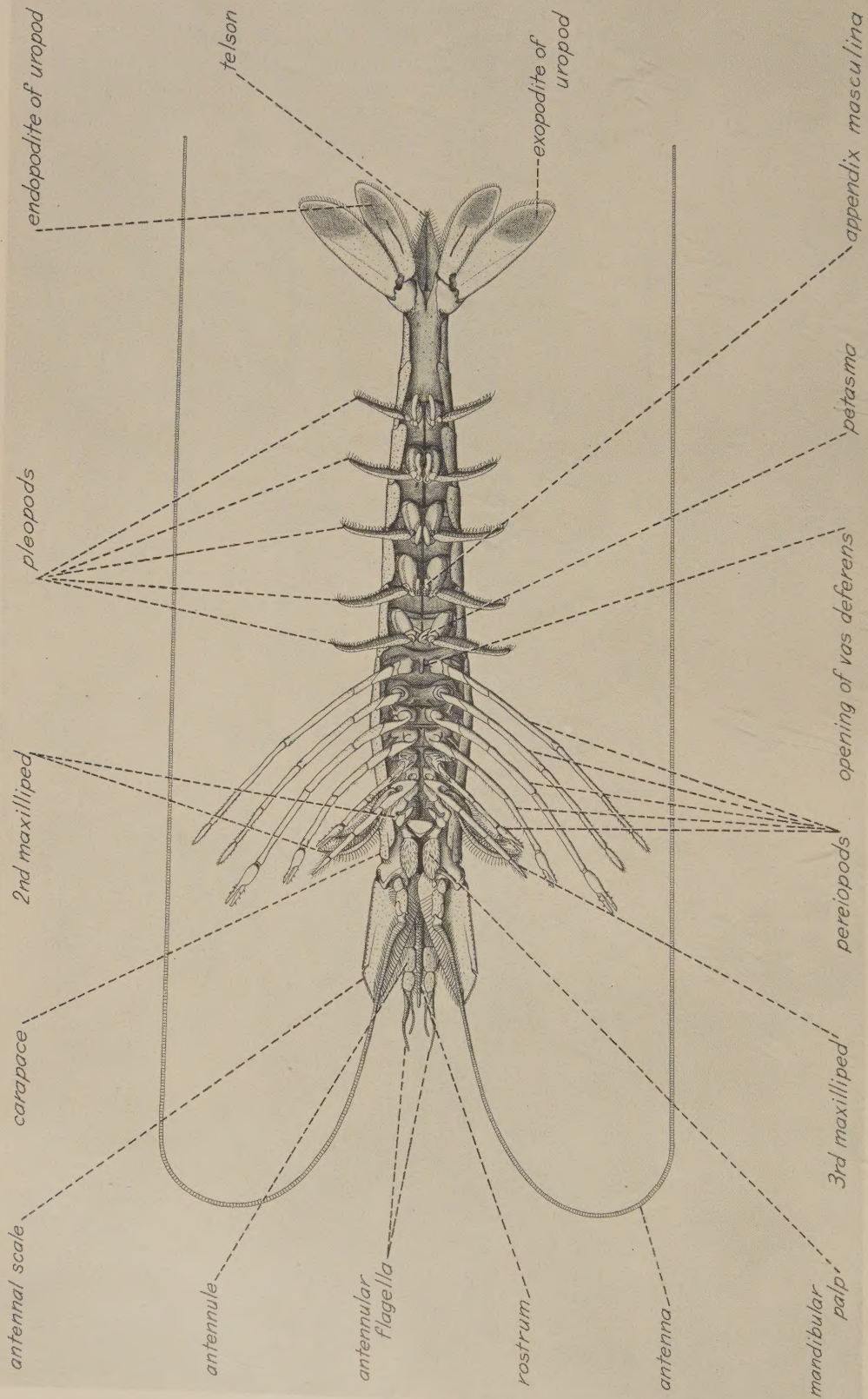


FIGURE 3.—Ventral view of young male.

The protocephalon is clearly distinct from the succeeding gnathothoracic and abdominal tagmata, and in *Penaeus setiferus*, and other species of the genus (Grobben 1917) is independently movable. Grobben, incidentally, described the area as the "sincipit," a term apparently discarded by Snodgrass in favor of the term, "protocephalon." In the section below on the eyestalks, a consideration of the muscles which move the protocephalon on the gnathothorax will be found. A discussion of the primitive order of the head segments follows in the section on the labrum.

SKELETAL ELEMENTS

The protocephalon is constructed roughly on the plan of a vertically placed hemisphere confluent posteriorly with the gnathothoracic hemocoel. The skeleton is flattened dorsally in the region of the ocular plate (fig. 4) or lobe from which the foramina of the eyestalks are perforated. The large, paired foramina of the antennules and antennae and the mesal foramen of the labrum occupy almost the entire anterior and ventral surface of the protocephalic skeleton. The latter regions are heavily sclerotized and deeply indented between the antennular and antennal foramina by the medial stem of the Y-shaped epistome (figs. 28, A; 30). The top, or posterior portion of the epistomal Y, divides across the anterior face of the labral foramen and the posterior side of the large antennal foramina. Upon the lateral epistomal bars, as shown by Snodgrass (1951), the medial mandibular condyles are located.

1. EYESTALKS

Most of the following section on the eyestalks was previously published by the writer (Young 1956).

In some of the lower and in many of the higher Crustacea, the compound eyes are set upon movable stalks or peduncles. Their presence at the ends of extensions has excited speculation for many years. Carcinologists have long discussed the reasons for the eyestalks, their similarity with other appendages (Calman 1909), and the nature of vision in a stalked-eyed animal, among other things. Yet little has been written about the mechanics of the eyestalk with respect to vision. No one has proposed any useful explanation for the fine adjustments presumably available to a compound eye which has as numerous oculomotor muscles as the crustacean stalked eye.

The presence of a set of muscles to move the corneal surface of the compound eye on the eyestalk, and of muscles to move the eyestalk about with respect to the body suggests the importance of the position of the corneal surface relative to the environment. By contrast, adjustments of corneal position in an arthropod without eyestalks suggests a function of head and "neck" muscles for activities other than feeding, if we assume any importance to the arthropod of corneal position adjustments.

Recently, the eyestalk nerves of a few crustaceans have been shown to contain neurosecretory elements which evidently proliferate hormone systems controlling such processes as molting (Passano 1953), retinal pigment migration (Welsh 1941), and chromatophore movements (Perkins 1928). In view of the concentrated attention currently being paid to matters of neurohormonal control of physiological functions in the arthropods, an understanding of the relation of vision to neurosecretion appears to be near at hand.

The white shrimp, *Penaeus setiferus*, carries its eyestalks at an angle of about 75° to the median sagittal plane and at an angle of about 10° to 15° to a frontal plane at the ocular plate (fig. 4). Only rarely are the eyes brought forward to lie in the optic depressions of the antennules, and then but for an instant for protection or possibly cleaning against the long plumules surrounding the depressions. Normally, therefore, in *P. setiferus* and many other species of shrimps, the eyes and stalks are widely spread and slightly upturned, a situation not understood by morphologists who, working with preserved materials, have described the eyestalks as projecting anteriorly. Had previous workers taken into account the lateral position of the eyestalk in the shrimps, and for that matter in the crawfishes, a certain amount of confusion in the naming of eyestalk musculature might have been avoided. For in fact, medial muscles are anterior and lateral muscles are posterior. In the interests of uniformity, however, certain of the incorrect names are here employed.

P. setiferus is a bottom feeder. According to an unpublished observation by Charles Dawson, Bears Bluff Laboratories, Wadmalaw Island, S. C., schools of penaeid shrimps are frequently found on muddy bottoms. This worker describes placing several *P. aztecus* Ives, 1891, in aquaria with an inch or two of mud on the bottom, into

which the animals immediately burrow, except for the eyes. Such behavior suggests that the long eyestalks are among the organs enabling the penaeid shrimp to make use of mud for protection, especially after molting.

In the past, observers have described square corneal facets in the eyes of several species of decapod crustaceans (Huxley 1906; Calman, 1909). A study of slides made of the corneal cuticle shows that the corneal facets in the compound eye of *Penaeus setiferus* are also square. Likewise, the underlying crystalline cone cells are square in the white shrimp and total four per ommatidium, as determined by the study of tangential sections of the eye from which the corneal cuticle had been removed. Ramadan (1952) reports a similar situation in a species of *Metapenaeus*. In longitudinal section the ommatidium of *P. setiferus* is like that of *Astacus*, as described by Bernhards (1916), with comparatively elongate crystalline cone and short rhabdom cells. If anything, the cone cell is longer in the white shrimp than in the crawfish. A light pink substance which gives the dark-adapted shrimp eye its bright color in strong light is the tapetum (Ramadan 1952). It is associated with the proximal or retinal pigment of the ommatidia.

SKELETAL ELEMENTS

The ommatidial surfaces arise from a sclerotized cup, previously named the optic calathus, or basket (Young 1956), to avoid confusion with the optic cup of the vertebrate embryo (fig. 4). The optic calathus rests upon the elongate stalk segment in a structural relation permitting universal movements, although the degree of movement varies in different planes.

Two points of articulation in the dorsoventral plane allow the optic calathus considerable horizontal movement around the distal end of its supporting stalk. These dorsoventral hinges are, however, sufficiently loose to permit vertical and rotational calathus movements, but to a lesser extent than horizontal movements. The long stalk is comprised externally of several longitudinal sclerotized bars which are separated by pliable cuticle. Two of the bars give support to the dorsoventral points of articulation (fig. 11) and others to less well-defined points of articulation between the stalk and calathus, and between the stalk and basal segment.

The stalk is movable upon the short, boxlike, basal segment in the horizontal plane. Vertical movements between the basal segment and the stalk are restricted. With respect to the structure here labelled the median tubercle (fig. 4), Anderson and Lindner (1943) and Voss (1955) state that shrimps of the subfamily Penaeinae have no distinct median tubercle on the ocular peduncle. However, many of the shrimps of this group do possess large, blunt, median tubercles, similar to those in *Penaeus setiferus*.

Set between the basal segments is the ocular plate or lobe, a broad, roughly rectangular sclerotized structure which encloses laterally the medial parts of the basal segments (fig. 4). The ocular plate is the dorsal-most region of the protocephalon. Movements between the basal segment and the ocular plate are similar in extent to those between the stalk and the basal segment. Horizontal movements are limited to an arc of about 15° or 20°.

MUSCLE ELEMENTS

PROTOCEPHALON MUSCLES OF THE OCULAR REGION

Taking origin from either the epistomal invagination or the dorsal surface of the carapace and inserting upon basal parts of the eyestalks are four pairs of muscles. The basal regions of the eyestalks will be assigned here to the dorsal part of the protocephalon.

ANTERIOR PROTOCEPHALON LEVATOR MUSCLES

FIGURES 34, 35

The tiny anterior protocephalon levator muscles are probably the muscles designated by Grobben (1917, 1919) as the protocephalon levators (*musculus levator sincipitis*) in a European penaeid and in other species of higher Crustacea. These muscles are difficult to make clear, either by dissection, or by illustration, since they take origin on the carapace, on the nearly vertical sides of the rostral base. During removal of the carapace and the underlying layers of tough, fibrous epidermis and connective tissue, these muscles are torn away. The anterior protocephalon levators insert in the heavy connective tissue associated with the posterior edge of the protocephalon. Their actual levation of the protocephalon is negligible, since they are not only minute in cross section, but

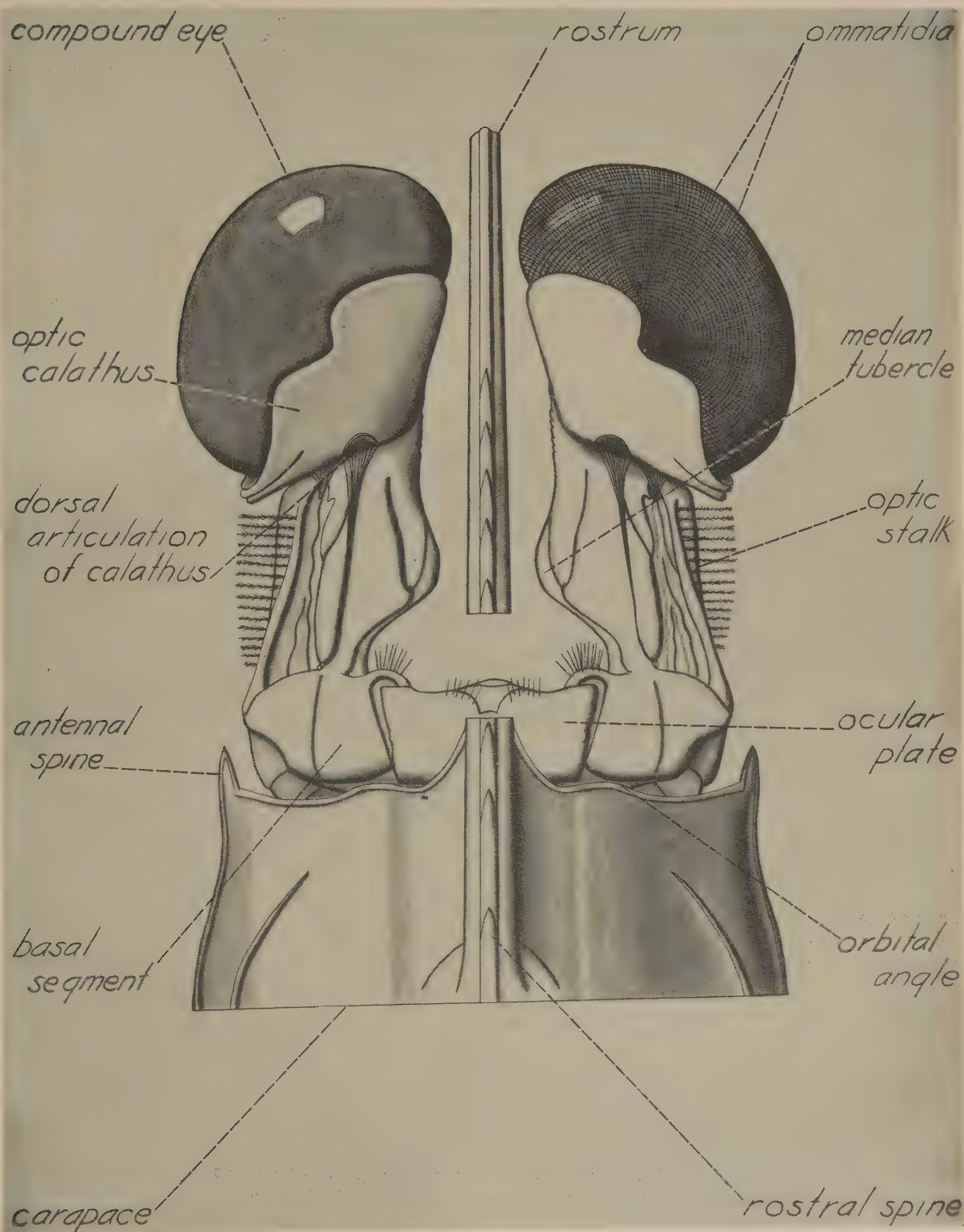


FIGURE 4.—Dorsal view of eyestalks in anterior position. Rostrum cut away to show ocular plate.

short in length. No counterpart of the anterior protocephalon levator muscles was described by Schmidt (1915) in *Astacus*, or by Berkeley (1928) in *Pandalus*. However, Grobben (1917) illustrates a protocephalon levator in *Astacus* and theoretically one would expect to find the muscle in *Pandalus*.

POSTERIOR PROTOCEPHALON LEVATOR MUSCLES

FIGURES 5, 6, 34

The function of moving the protocephalon dorsally is performed by a pair of large muscles, the posterior protocephalon levator muscles, which originate close together at the dorsal midline of the carapace somewhat posterior to the origin of the anterior protocephalon levator muscles and which run forward and downward to attach on a nearly vertical transverse plate, posterior to the postocular region of the eyestalk base (fig. 6). The muscle inserts ventrally to the insertion of the anterior levators. The contraction of the posterior protocephalon levators may also act to rotate posteriorly the eyestalk base and hence raise the extended eyestalks.

Possible homologs of the posterior protocephalon levator muscles are the median dorsal muscles designated as the musculus oculi basalis posterior. In *Astacus*, Schmidt (1915) found that these muscles arise on the median dorsal surface of the carapace and are attached by short tendons to the much longer tendons of other, more anteriorly placed muscles, the musculus oculi basalis anterior. The anterior eye base muscles, to angulize freely, are attached to the median dorsal region of the eyestalk base (Schmidt 1915). More will be said of the latter muscles below.

The posterior eye base muscles, it should be emphasized, do not attach to the eyestalk base in *Astacus*, but if the assumption is made that, due to the immovable protocephalon in *Astacus*, the attachment of the muscles to the eyestalk base has moved in that animal to the tendons of the anterior eye base muscle, then a homology with the posterior levators in the white shrimp may be proposed. However, the extensive rearrangement of muscle attachments upon which the assumption is based weakens the proposal.

Even more significant, muscles exist in *Penaeus setiferus*, as will be shown later, which are more likely to be the homologs of the anterior and posterior eye base muscles in *Astacus*, *Pandalus*,

and *Callinectes* than are the posterior protocephalon levator muscles. If the latter is true then the posterior protocephalon levators have been lost during the evolution of *Astacus*, *Pandalus*, and *Callinectes*, in which forms no trace of the muscles appears (Schmidt 1915; Berkeley 1928; Cochran 1935).

OCULAR PLATE DEPRESSOR MUSCLES

FIGURES 5, 6, 7

The ocular plate depressor muscles originate on the posterior surface of the epistomal invagination. They run anterodorsally, passing beneath the insertions of the posterior protocephalon levator muscles, and insert broadly on the posterior edge of the ocular plate (figs. 5, 6, 7). On contraction, the ocular plate depressors draw the ocular plate posteriorly and ventrad. Based on the attachment points of the muscles in *Penaeus setiferus*, they may have given rise by partial fusion to the anterior eye base muscles (musculus oculi basalis anterior) as they are found in the European crawfish, where the muscles are attached ventrally to the epistomal region by a long tendon and run dorsad to the edge of the eyestalk base. Cochran (1935) describes in *Callinectes* a pair of anterior eye base muscles which arise from a kind of epistomal invagination rather like that in the white shrimp, but instead of fusing as in the European crawfish, they diverge slightly laterad in the blue crab in probable accompaniment with the general broadening of the body in the Brachyura.

The ocular plate depressor muscles apparently are homologous with the musculus oculi basalis anterior in *Pandalus*, the name for which Berkeley (1928) has taken from Schmidt (1915). In *Pandalus*, these muscles are similar to those in *Penaeus*, except that they are slightly separated, whereas in *P. setiferus* they are close together.

PROTOCEPHALON ATTRACTOR MUSCLES

FIGURES 5, 6, 7, 8

The protocephalon attractor muscles are two very large muscles which take broad L-shaped origins on the carapace and run anteriorly to insert on two pairs of large apodemes and on other parts of the protocephalon. The largest apodeme, on which the ventral-most part of the protocephalon attractor muscles inserts, arises from the ventral surface of the antennular foramen, broadening posteriorly into a large vertical sheet

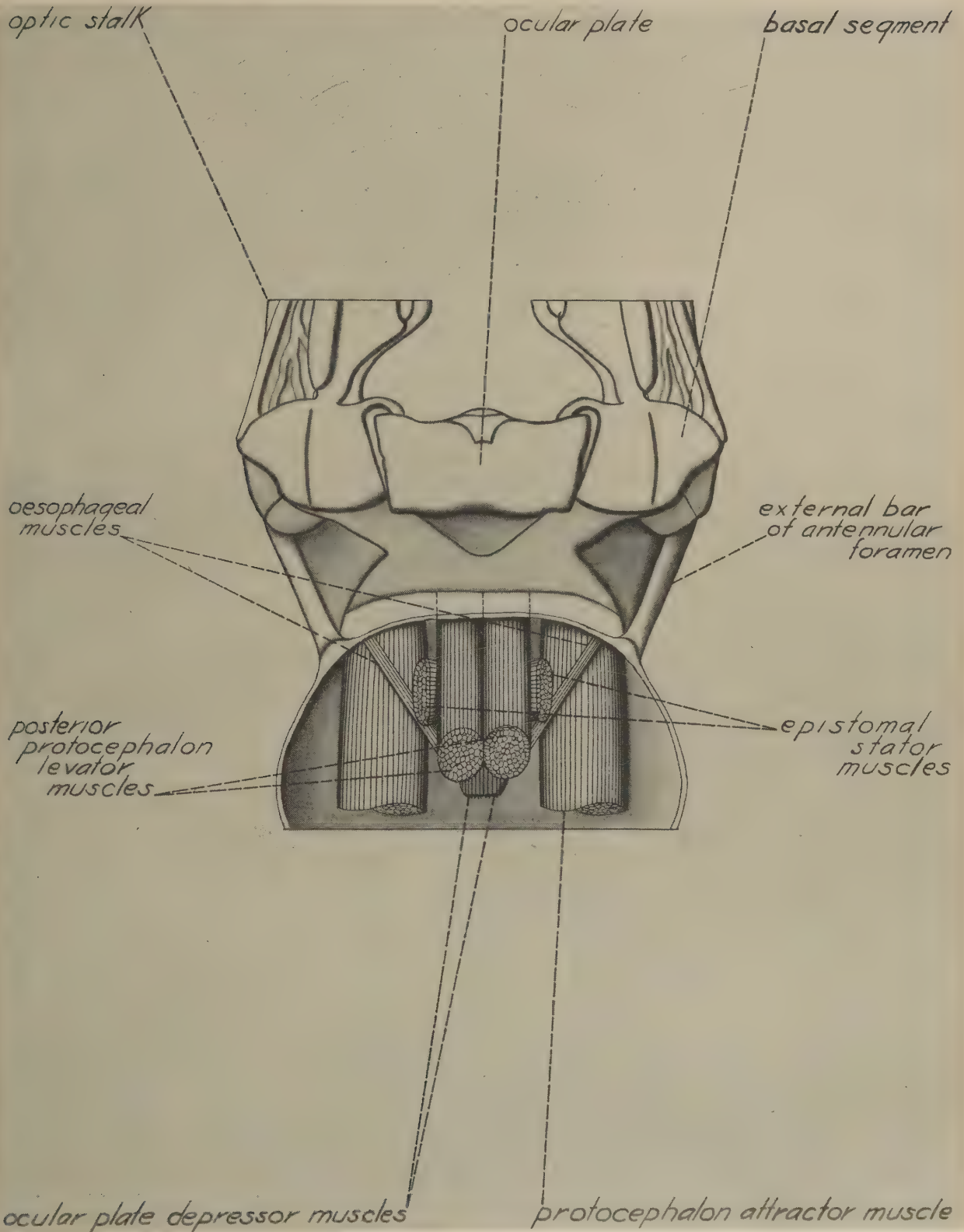


FIGURE 5.—Dorsal view of protocephalon, carapace removed, to show muscles of postocular region.

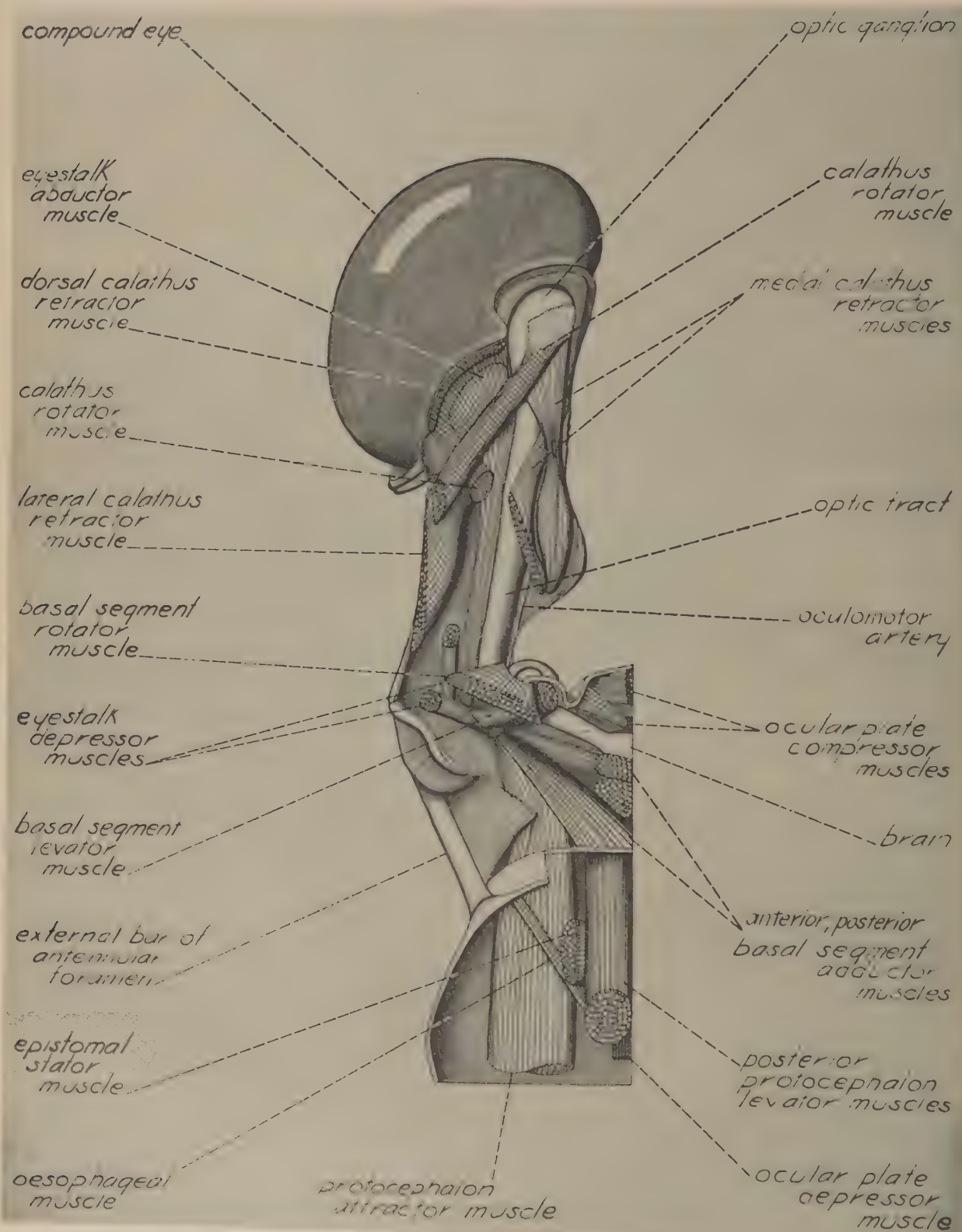


FIGURE 6.—Dorsal view of left eyestalk. Dorsal cuticle and carapace removed to show muscles.

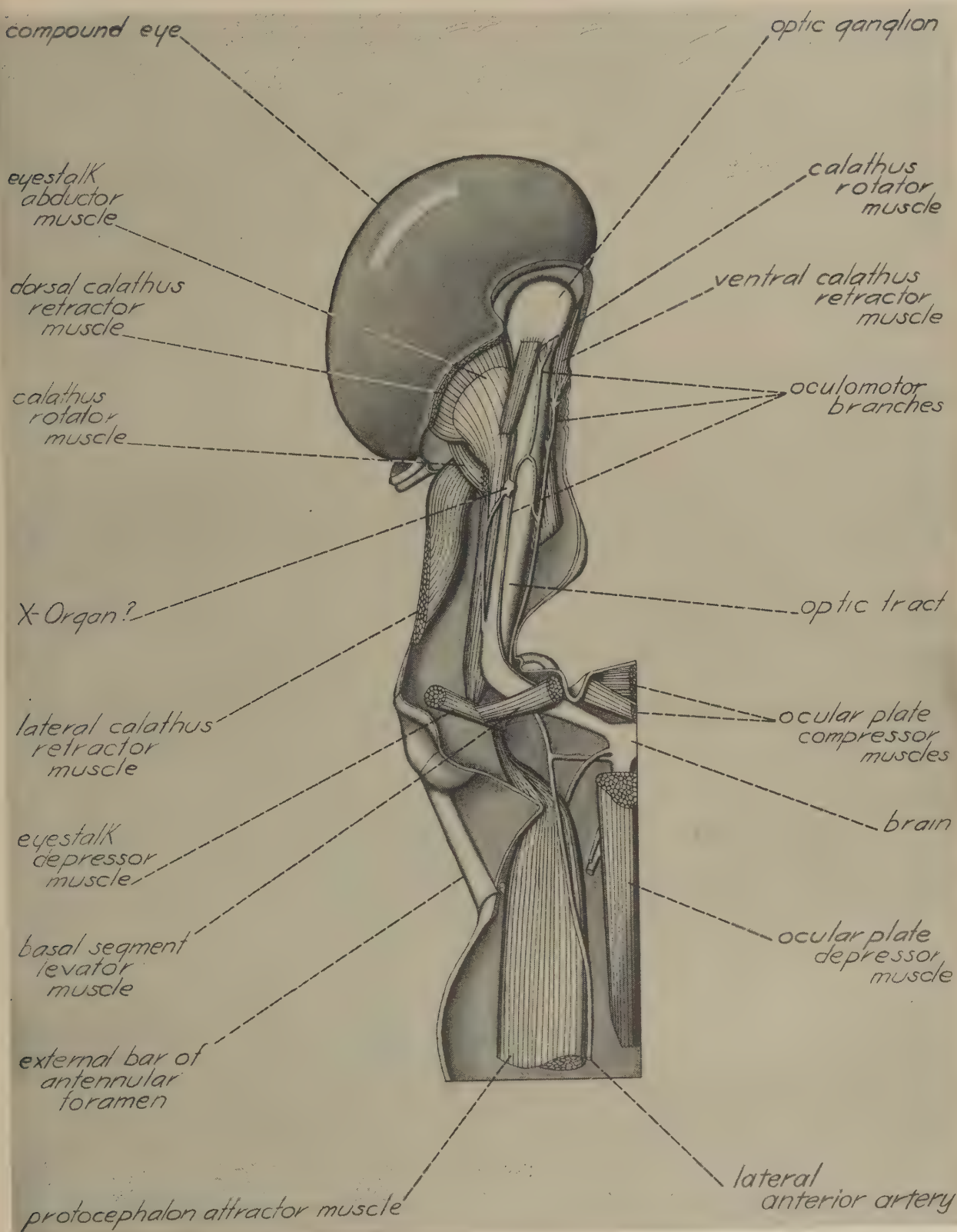


FIGURE 7.—Dorsal view of left eyestalk. Dorsal muscles removed to show branches of nerves and arteries.

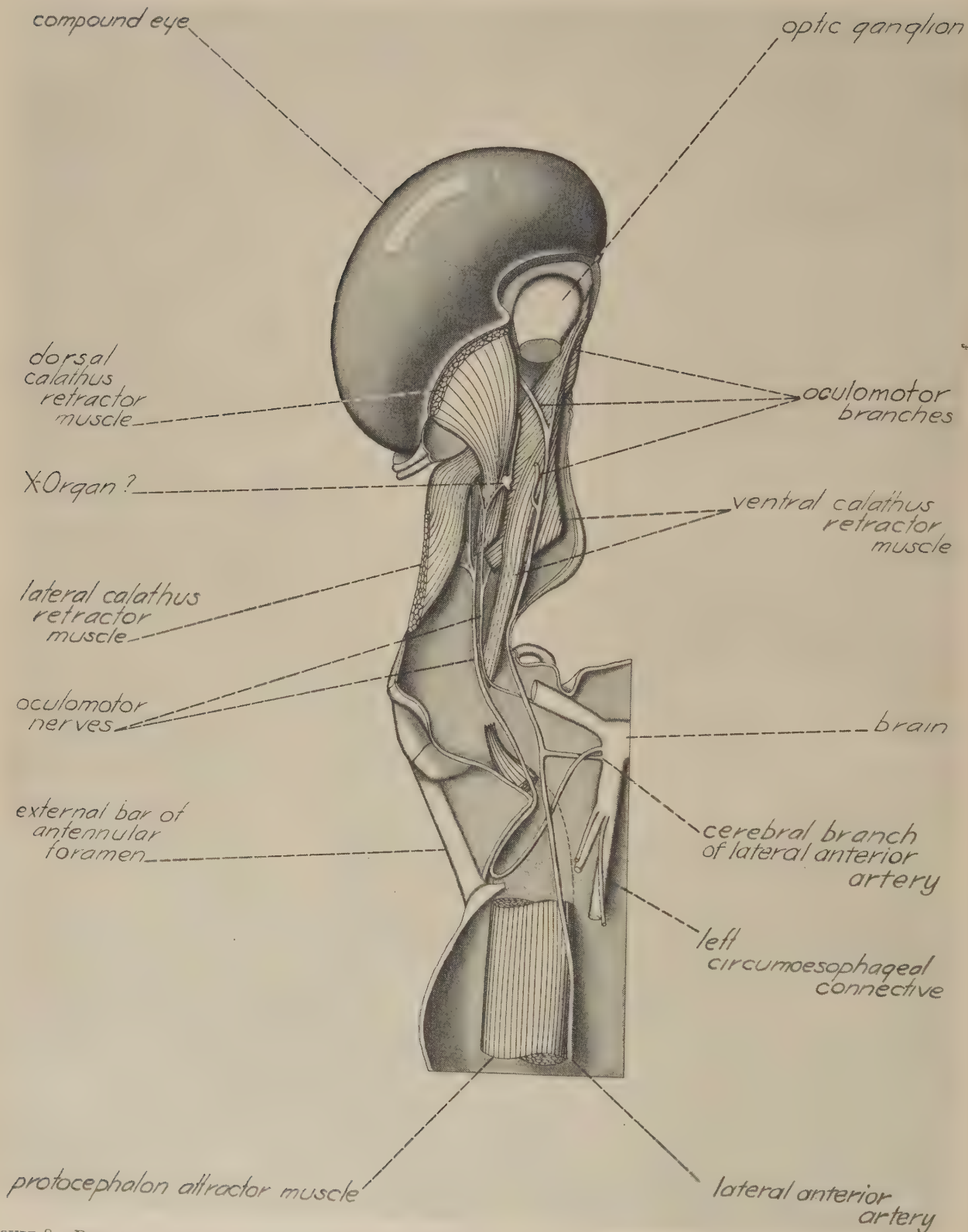


FIGURE 8.—Dorsal view of left eyestalk. Dorsal muscles and optic tract removed to show ventral muscles and branches of nerves and arteries.

of cuticular material. Slightly anterodorsal to the antennular apodeme, in the same parasagittal plane, is an apodeme which invaginates from the ventral floor of the basal segment of the eyestalk and projects through the ventrolateral side of the eyestalk foramen into the thoracic hemocoel. This apodeme, like that near the antennule, broadens vertically. By virtue of apodemal position, that part of the protocephalon attractor muscle attaching upon the eyestalk apodeme is somewhat longer than is the part inserting on the antennular apodeme.

The longest and most dorsal part of the protocephalon attractor muscle extends anteriorly beyond the antennular and eyestalk apodemes, to insert slightly laterad in connective tissue at the ventral surface of the basal segment of the eyestalk (figs. 7, 8).

To these comparatively huge protocephalon muscles we may ascribe at least two functions, namely, (1) attraction of the protocephalon, and (2) adduction of the eyes. The largest of the three inserting bodies of the muscle is the ventral-most part inserting on the antennular apodeme, described above. From a study of the points of articulation between the carapace and the protocephalon, dorsally, and the mandibular segment and the protocephalon, ventrally, the primary result of contraction of the ventral muscle body would be to draw the protocephalon directly posterior. The muscle was described as a protocephalon depressor (*musculus depressor sincipitis*) by Grobben (1917) in a number of Decapoda and in the stomatopod *Squilla*. This worker has shown that the protocephalon attractors, or depressors, are missing in those forms, like *Astacus*, in which the protocephalon is immovably fused to the thorax. The point of Grobben's discussion, that the protocephalon is movable on the gnathal tagma in the generalized crustacean, is not changed by a difference of opinion over the function of the muscle under consideration.

Furthermore, these muscles are not antennular in any way, having, as brought out by Snodgrass (1951), origins on the carapace. They are also, to say the least, not strictly antennal, since a portion of the muscle inserts in the eyestalk. Although the protocephalon attractor muscles in *Penaeus brasiliensis* Latreille, 1817, may be widely different from those in *P. setiferus*, the statement of Knowles (1953) that the two muscles lying just beneath the anterolateral side of the

carapace are antennal is probably in error. From his figures the external-most muscle is properly a remotor of the antennal scale, while the large inner muscle is the protocephalon attractor muscle.

The two anterodorsal parts of the protocephalon attractor muscles which find insertions in the eyestalks have, in addition to attraction, the function of eyestalk adduction. Upon contraction of the whole muscle, these dorsal fibers in the eyestalk draw the ocular plate and attached eyestalk segments posteriorly toward the carapace. The posterior side of the basal segments makes contact with a condylic thickening on the anterior edge of the carapace, at a point known as the orbital angle (fig. 4), thereby swinging the eyestalks forward in a horizontal plane into the ocular depressions on the antennules. As suggested in introductory comments, the movement is a quick one, much faster than the return of the eyes to the spread position. In *Penaeus setiferus* other muscles exist which function to adduct the eyes, but their effect is negligible when compared to that of the much larger protocephalon attractor muscles.

The protocephalon attractor muscles appear in *Pandalus*, designated by Berkeley (1928) as the depressor muscles *c* of the antennae, on grounds of their attachment to the basipodites of those structures. At the same time this worker ascribes to the depressor muscles *c* the function of adduction and rotation, rather than the depression of the antennae. Berkeley's name for the muscles obviously was taken from the work of Schmidt (1915) on *Astacus*, in which form the antennal depressor muscles *c*, while small, nonetheless depress the antennae. Although proof must wait upon a study of the nerves in *Penaeus* and *Pandalus*, Berkeley has homologized the so-called depressor muscle *c* of *Pandalus* and *Astacus* on the basis of their dorsolateral origins on the carapace and their insertions on the mediodorsal edge of the antennal basipodite (in *Pandalus*) and coxopodite (in *Astacus*).

That the depressor muscles *c* in *Pandalus* and the protocephalon attractors in *Penaeus* are homologous seems fairly certain, despite the apparent change of insertion in the former. A review of cleared and stained exoskeletons of *Pandalus* might show multiple insertions of the muscle as in *Penaeus*. The homology of the protocephalon attractor muscles in *Penaeus* with the depressor muscles *c* in *Astacus* is less certain. In *Callinectes*, Cochran (1935) figures a pair of

ocular attractor muscles which originate on the carapace. Their phylogenetic relation to the protocephalon attractor muscles in *Penaeus* is unlikely.

OCULAR PLATE MUSCLES

Arising in the ocular plate or postocular region dorsal to the brain are several pairs of muscles and a muscle group. Some of these muscles insert inside and some outside of the ocular plate.

OCULAR PLATE COMPRESSOR MUSCLES

FIGURES 6, 7

Attached about the shallow anterodorsal groove of the ocular plate is a group of muscles which runs to the lateral wall of the ocular lobe (figs. 6, 7), the ocular plate compressor muscles. They function to draw the sides of the head lobe and ocular plate mesad, and to depress slightly the center of the ocular plate.

ANTERIOR BASAL SEGMENT ADDUCTOR MUSCLE

FIGURE 6

The anterior basal segment adductor muscle originates on the ocular plate dorsal to the brain and attaches to connective tissue and apodemal material in the ventral part of the basal segment (fig. 6). Contractions of the muscle turn the basal segment toward the ocular plate in a horizontal plane.

POSTERIOR BASAL SEGMENT ADDUCTOR MUSCLE

FIGURE 6

The posterior basal segment adductor muscle inserts in the basal segment at the same point as the anterior basal segment adductors, but originates on the anterior side of the vertical transverse plate posterior to the postocular region (fig. 6). It, too, draws the anterior edge of the basal segment toward the ocular plate. The origins of these muscles are so widely separated that we may conclude that they have never been the same muscle. How the basal segment adductors may be homologized with muscles in *Pandalus* and *Callinectes*, in which forms no knowledge of muscle innervations exists, will be speculation. The ocular adductor muscles of *Astacus* and *Pandalus* may well be the homologs of the anterior adductor muscles of *Penaeus*, but hardly with the ocular adductors of *Callinectes*, in which animal the muscles are

located in the distal end of the long stalk segment. Phylogenetic relationships of the posterior basal segment adductor muscle are even more uncertain, although possibly it is the same muscle as the ocular attractor muscle in *Pandalus* and *Astacus*. The basal segment adductor muscles do not appear in *Callinectes*.

BASAL SEGMENT LEVATOR MUSCLE

FIGURE 6

The basal segment levator muscle originates at the anterodorsal corner of the ocular plate and runs ventrally to the connective tissue and apodemal cuticle on the ventral surface of the basal segment (fig. 6). In the normal spread condition of the eyestalk, contraction of the muscle tends to raise the basal segment and with it the extended eyestalk.

BASAL SEGMENT MUSCLES

In the functional descriptions of the muscles which follow, the eyestalks will be considered as in their lifelike, lateral positions.

BASAL SEGMENT ROTATOR MUSCLE

FIGURE 6

The basal segment rotator muscle is a short, broad structure originating on the anterodorsal edge of the basal segment and inserting on the anteroventral edge of the same segment. Upon contraction, the muscle pulls the dorsal surface of the basal segment anteriorly, thus rotating the entire eyestalk forward.

EYESTALK DEPRESSOR MUSCLES

FIGURE 6

Two very small muscles, the eyestalk depressor muscles, one slightly lateral to the other (fig. 6), function to draw the eyestalk ventrally.

EYESTALK MUSCLES

EYESTALK ABDUCTOR MUSCLE

FIGURE 6

All of the muscles of the eyestalk and optic calathus are associated with retraction and rotation of the optic calathus on the eyestalk, except the long eyestalk abductor muscle (fig. 6). The proximal end of the eyestalk abductor muscle is attached in connective tissue in the ventral region

of the basal segment. The muscle runs the length of the eyestalk to insert in connective tissue near the dorsal calathus retractor muscle. Contraction of the muscle swings the eyestalk horizontally to a lateral position. The eyestalk abductor muscle of *Penaeus* is very likely homologous with the abductor muscle described for *Astacus* and *Pandalus*, and possibly with the lateral branch of the ocular abductor muscle in *Callinectes*.

CALATHUS RETRACTOR MUSCLES

The muscles in *Penaeus* which retract the optic calathus appear to be clearly represented by the retractor muscles of the eyes of *Astacus*, *Cambarus bartoni* (Fabricius 1798), *Pandalus*, and *Callinectes*. Phylogenetically, the situation in *Penaeus* is somewhat more generalized than in the other forms which we are considering, in that several of the calathus retractor muscles in *Penaeus* have more than one part. In addition, *Penaeus* has a number of apparently independent rotator muscles, none of them previously described, which function to twist the optic calathus about a longitudinal axis through the eyestalk.

DORSAL CALATHUS RETRACTOR MUSCLE

FIGURES 6, 7, 8

The dorsal calathus retractor muscle arises in connective tissue near the ventral surface of the eyestalk and attaches to the dorsal edge of the calathus.

LATERAL CALATHUS RETRACTOR MUSCLE

FIGURES 6, 7, 8, 9

The lateral calathus retractor muscle, really the posterior retractor, originates on sclerotized material along the lateral, or actually posterior, blood sinus running the length of the eyestalk. The larger portion of this muscle attaches on the lateral edge of the calathus, the lesser part turning ventrally and running across the ventral edge of the calathus, just dorsal to the ventral retractor muscles (fig. 9). When this muscle contracts it not only retracts the calathus, but rotates the calathus about an axis longitudinal to the eyestalk.

VENTRAL CALATHUS RETRACTOR MUSCLE

FIGURE 9

The ventral calathus retractor muscle originates on several sclerotized regions on the ventral sur-

face of the eyestalk. One part of the muscle is long and slender, while the others are short and arise from broad origins (fig. 9). The muscle is inserted over a wide area on the ventral edge of the calathus.

MEDIAL CALATHUS RETRACTOR MUSCLE

FIGURE 6

The medial calathus retractor muscle originates on two points in the region of the median tubercle, and actually is comprised of two muscles (fig. 6). The larger muscle originates in the median tubercle and inserts in connective tissue dorsal to the distal optical ganglionic mass. The smaller muscle originates dorsal to the larger muscle and inserts on a ventromedial point on the calathus. The contraction of both muscles results in medial retraction of the calathus; functioning in opposition, the muscles retract the calathus in a vertical plane, reinforcing the action of the dorsal and ventral retractor muscles.

CALATHUS ROTATOR MUSCLES

FIGURES 6, 7

At least three calathus rotator muscles may be seen in the eyestalk of *Penaeus setiferus*. Rotator muscles of this type have not been described for *Pandalus*, *Astacus*, *Cambarus*, or *Callinectes*. The calathus rotators bear a certain similarity to one another, in that they are all superficial in position and insert in the heavy connective tissue underlying the thick cuticle of the calathus.

2. ANTENNULES

The antennules, or first antennae, are said to belong to the second segment of the body of Crustacea, following the eyes. In the company of the eyes, the antennae, and the labrum, the antennules are attached to the body tagma that has been termed by Snodgrass (1951a; 1952) the protocephalon. Whereas the appendicular nature of the eyestalk generally has been questioned, the status of the antennules as true crustacean appendages has rarely been attacked, despite controversy over the homologies of the component segments.

The antennule of *Penaeus setiferus* is comprised of a proximal stem divided into three basal segments, called the protopodite by Huxley (1906), and two distal flagella. Proponents of theories such as Huxley's (1906) suggesting correspondence of parts between the segments of the anten-

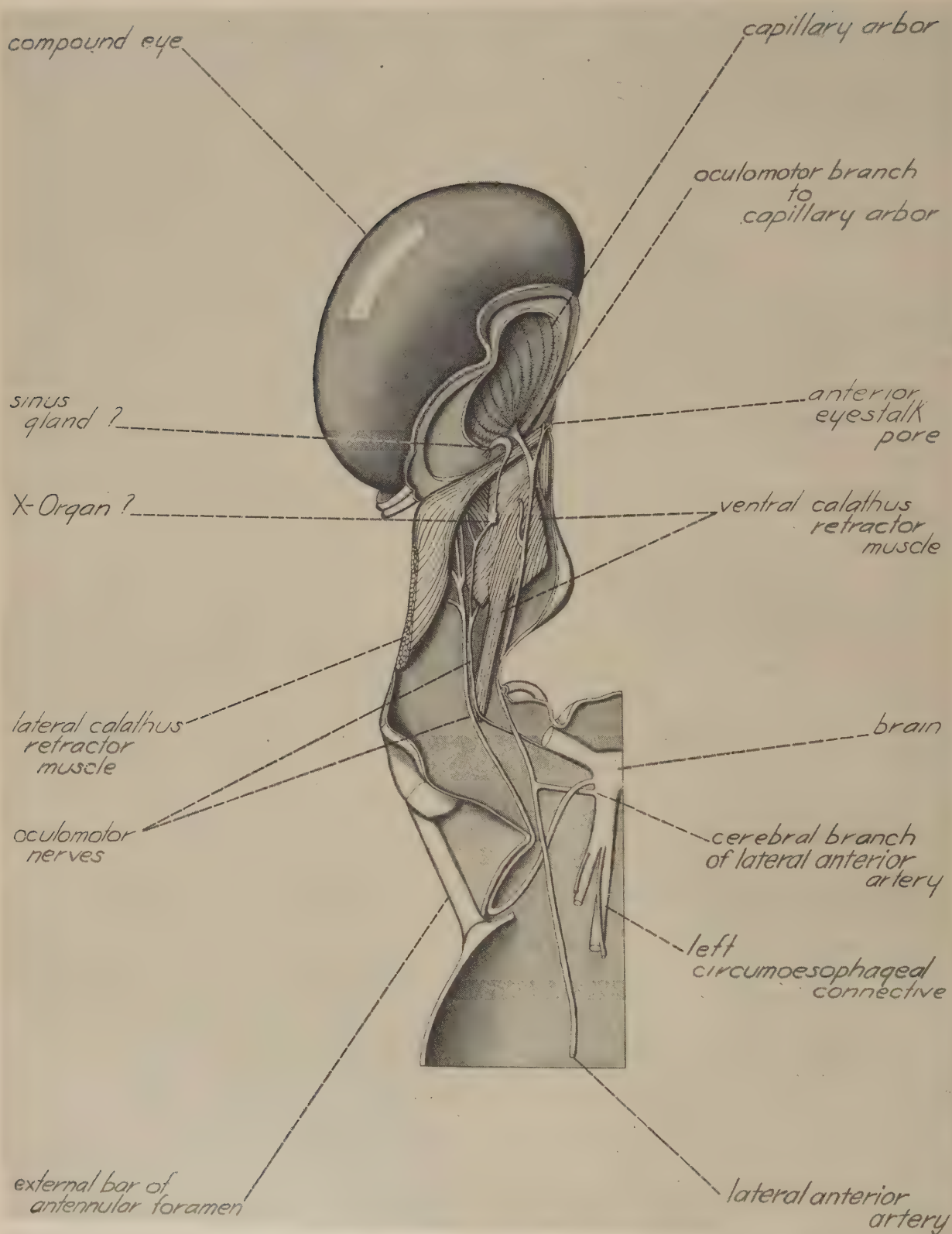


FIGURE 9.—Dorsal view of left eyestalk. Dorsal muscles and optic tract removed to show brain, branches of nerves, arterial capillary supply to distal optic ganglia, neurosecretory glands, and location of anterior eyestalk pore.

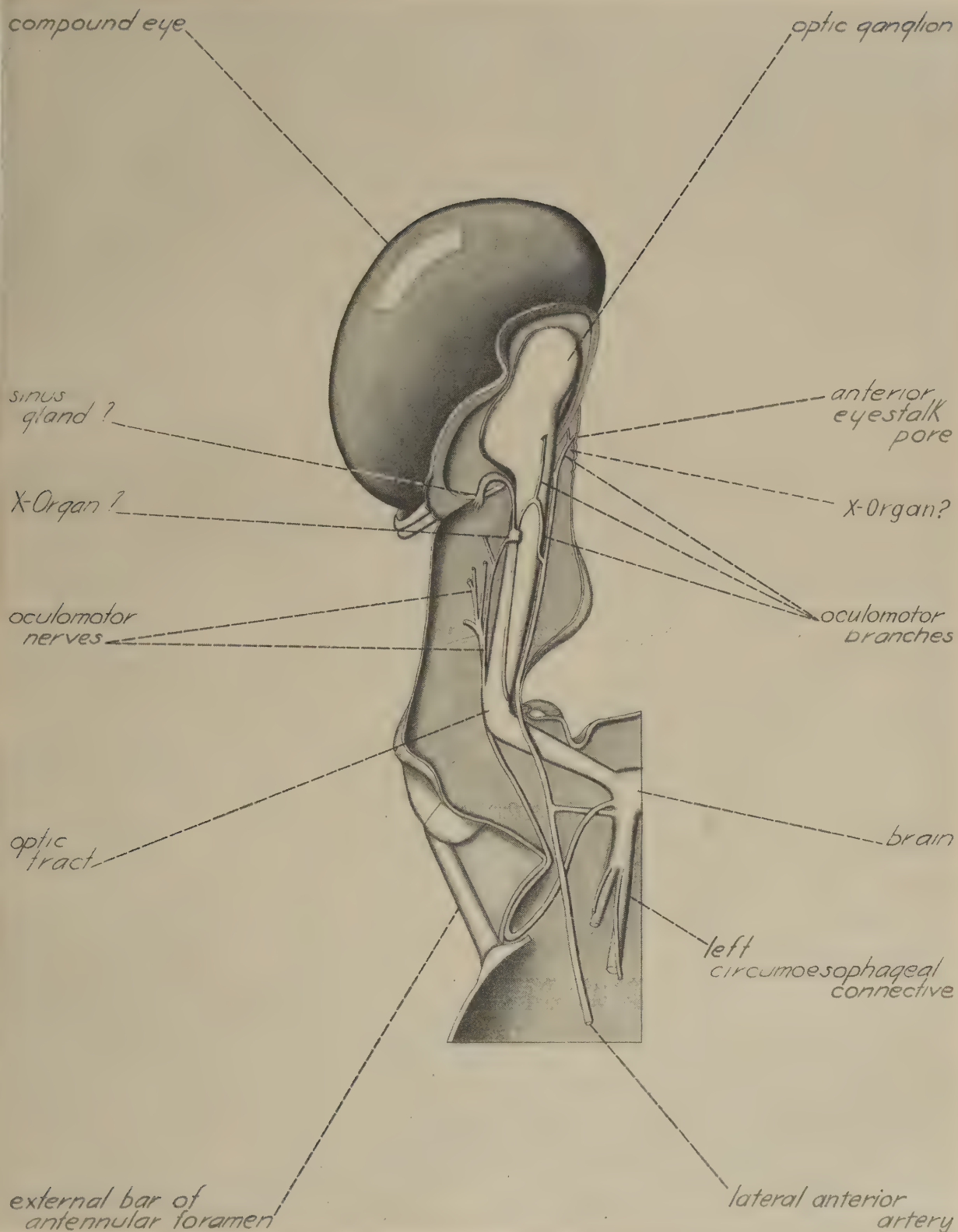


FIGURE 10.—Dorsal view of left eyestalk. Muscles removed to show optic tract, oculomotor nerves, neurosecretory glands, and branches of lateral anterior artery.

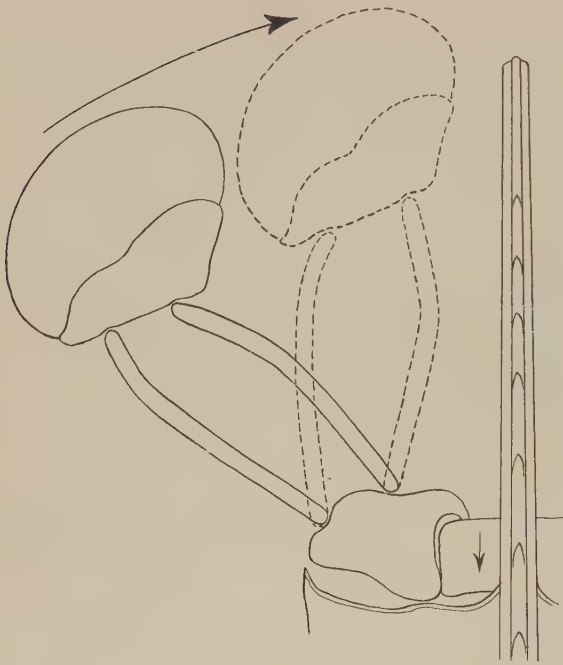


FIGURE 11.—Dorsal view of left eyestalk. Diagrammatic. The skeletal bars of the eyestalk are so arranged that the position of the ommatidial surface is maintained whatever the angle of the eyestalk with respect to the long axis of the shrimp.

nule and those of the typical crustacean appendage have not received support. Most carcinologists consider the parts of the antennule so extensively modified from the usual plan as to defy identification. At the same time, the clearly segmented structure of the antennule marks it as a true appendage, in the opinion of many workers.

The form and function of the crustacean antennule usually is said to be constant among the Decapoda, relative to the extraordinary variability in the appendage occurring in the lower Crustacea. Nonetheless, among the decapods wide differences are found. The outer or lateral flagellum is split in many of the Caridea, giving the impression that the antennule bears three flagella. Among the tribe Penaeidae, the outer flagellum is prehensile in certain genera of the Family Sergestidae. In the Solenocera each flagellum is semitubular, thus forming a siphon (Calman 1909). The antennule is small and reduced in the Brachyura.

Despite infrequent anatomical treatment of the decapod antennule, the available work suggests that a marked uniformity exists in the appendage and its parts over a broad spectrum of the Decapoda. From the standpoint of comparative mor-

phology this feature of similarity would appear to simplify the process of homologizing part to part, and does, except for the matter of muscle names, a problem discussed earlier.

SKELETAL ELEMENTS

In dorsal view the antennule is shaped in a long wedge pointing anteriorly, the broadest portion being the proximal part of the first segment at the region of attachment of the antennule with the skeleton of the body. Each successive distal segment gradually becomes narrower. The mesial edge of the basal segments is straight and flat where the appendage rests against the other antennule. The first, or proximal, antennular segment (fig. 12) is relatively larger and more complex than the segments distal to it. The first segment is attached by a large articular foramen to the protocephalon, the tagma to which the antennules belong. Here, dorsal and ventral points of articulation, or condyles, afford the first antennular segment limited horizontal movement.

Centered in the posterior region of the first antennular segment is the statocyst, the organ that is thought to mediate the special sense of equilibration (figs. 12 to 16). Earlier workers termed the structure an otocyst, having ascribed to it an auditory function (Huxley 1906). The statocyst is constructed like an incomplete sac that has dorsal and anterior openings. Hairs, presumably sensory, project from the inner surface of the cuticular sac into the lumen of the statocyst. The hairs are arranged in regular rows. The rows of hairs are confined to oddly shaped patterns located at various places on the inner surface of the statocyst. Nerves from the sensory hairs coalesce ventrad of the sac into a broad, flat nerve tract that enters the brain at a point ventral to the optic tract (fig. 14).

The dorsal opening of the statocyst is partially covered by a fleshy, heavily setose lobe, the dorsal closing lobe (figs. 12, 15). In fresh material the dorsal closing lobe may be lifted from the dorsal opening of the statocyst without great difficulty. The saccular statocyst is shut anteriorly by a thin, curled sheet of cuticle that arises vertically by evagination from the ventrolateral floor of the antennular eye depression. The sheet is designated here as the anterior closing plate (figs. 12 to 15). Neither the dorsal closing lobe nor the anterior closing plate effect complete obturation of the statocyst. As a result, water circulates through

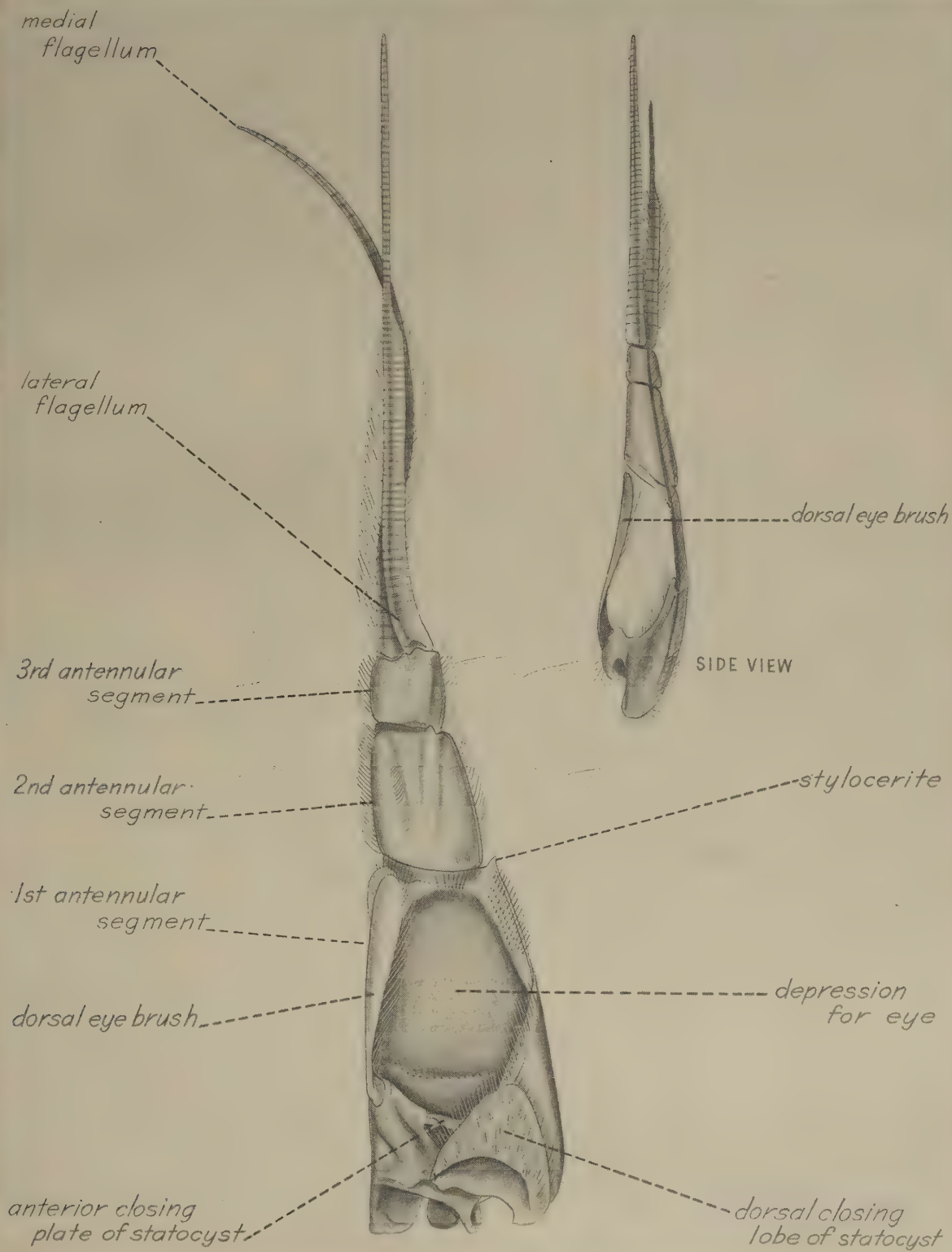


FIGURE 12.—The larger figure is a dorsal view of the right antennule, the smaller a lateral view of the right antennule.

the statocyst at all times, to the extent that the heavy investment of hairs in the area permits.

In living shrimps, as has been shown by Burkenroad (1939), the open statocyst contains sand grains. The sand probably functions as statoliths, if the statocyst is indeed equilibratory. These statoliths are obtained by the shrimp from the substrate through activities of the animal that are either directed or incidental to their collection. cursory examination of the statocyst does not reveal bodies that have obviously been secreted by the shrimp. Burkenroad (1939), however, finds that the statocyst of penaeids contains statoliths secreted by the animals. Since the statocyst is open to the water and to the substrate into which the shrimp is known to burrow, the possibility exists that the statocyst contents may undergo continual replacement during intermolt periods. However, since the shrimp loses the statocyst with its contents at each ecdysis, the statoliths are probably replaced in large measure at the time. That the animals burrow into the substrate for protection, statolith replenishment, and rest following the molt is supported by negative information provided by observing shrimps in clean aquaria at ecdysis. Upon shedding in this unnatural environment, the shrimp is unable to navigate properly and soon perishes, even if kept alone. The statocysts are found to be empty of statoliths.

The largest region of the first antennular segment is the eye depression. Beginning at the anterior closing plate and extending to the margins of the first antennular segment distally and to the sides is a broad, deep concavity into which the corneal surface of the compound eye may rest (figs. 12, 13). The eye depression is confined proximally by the anterior closing plate of the statocyst and the skeletal structures that surround the statocyst. The dorsal closing lobe projects out slightly, dorsal to the depression. A large, wedge-shaped, fleshy lobe arising in the posterolateral region of the first segment extends anteriorly along the lateral margin to a point. At the anterolateral corner of the first antennular segment is a small, sharp stylocerite (fig. 12), a structure common to the Tribe Penaeidae (Voss 1955). Along the mesial margin of the first segment lies the dorsal eye brush, or prosartema (figs. 12, 13), a long, thin lobe arising dorsally from the proximal region of the segment and extending anteriorly to the anteromesial corner of the segment.

Most of the structures lining the eye depression are heavily setose, to a degree that would make their outlines obscure if illustrated faithfully. For this reason only a fraction of the true covering in hair of the dorsal side of the antennule has been shown. The presence of this extensive investment of hair probably can be explained as a system of brushes to clean the corneal surfaces of the compound eyes. Many stalk-eyed crustaceans carry the eyes laterally and never for any length of time in the eye depressions of the antennules. However, the eyes are brought frequently into and out of the depressions, and thus through the long hairs lining the depression.

The second antennular segment (fig. 12), a far simpler structure than the first segment, is attached to the latter distally. The second segment articulates with the first segment allowing limited horizontal movements of the antennular segments distal to the first segment. The second segment is a rectangular box in shape and modified for the actions of the muscles which it contains.

The third antennular segment (fig. 12) is a small, square structure articulated with the second segment. Movement of the third segment and the flagella on the second segment is limited to an attenuated horizontal arc. No special sense organ appears in the third antennular segment.

Two flagella, the medial flagellum and the lateral flagellum (fig. 12), articulate independently with the third antennular segment. The points of articulation between the flagella and the third segment are so arranged that the flagella may move through a broad arc in the frontal plane. Each articulates with the third segment in slightly different horizontal planes. The lateral flagellum is attached dorsad of the medial flagellum. The flagella are composed of many short articles of light construction connected by rings of thin cuticle, thus permitting bending in all planes. The length of the articles differs in the two flagella, those of the lateral flagellum being shorter than the articles of the medial flagellum. Other differences between the flagella include variation in cross-sectional shape and in the types of setae and processes projecting from the articles.

Numerous studies have been made on the function of the antennular flagella and the variety of hairs and processes that they bear. These investi-

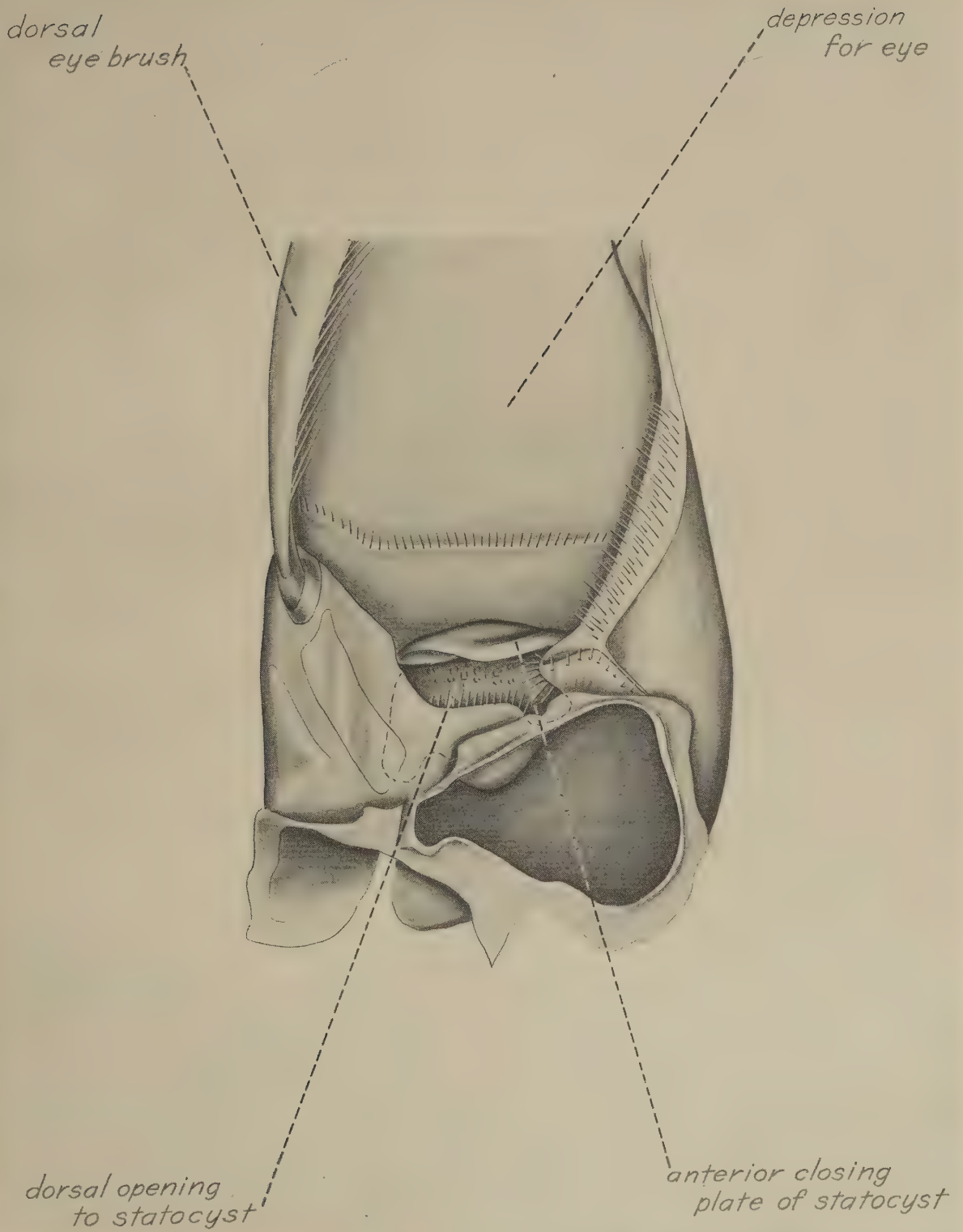


FIGURE 13.—Dorsal view of proximal region of first antennular segment, right antennule. Cleared specimen showing entrance to statocyst. Dorsal closing lobe removed.

gations show that the flagella are involved in chemoreception (Doflein 1911; Balss 1913; Bell 1906; Lissman and Wolsky 1935; Spiegel 1927). The olfactory nature of the antennular flagella is related to an interesting structural modification. This modification is the apparent sexual dimorphism of the medial flagellum in the adult male of *Penaeus setiferus* (fig. 17). The change to the medial flagellum, which is easily visible to the naked eye, results in a pronounced dorsoventral flattening, together with the appearance of numerous, stout processes of two sizes on the dorsal margin. That these processes may be predominantly proteinaceous is suggested by their loss following treatment of the preserved material in strong alkali, for as is well known strong alkali degrades the glucosamine of chitin but does not immediately remove it, whereas the alkali-soluble proteins of the internal organs and cuticle are rendered soluble and washed out (Richards 1951).

Other changes to the condition of adult maleness probably occur during the same molt that modifies the medial flagellum. The most obvious secondary sexual character is the mesial joining of the previously free wings of the modified pleopod endopodites of the first abdominal segment to form the definitive and functional petasma, or sperm-transfer organ.

Since the antennular flagella are olfactory, the sexually dimorphic medial flagellum of the male *Penaeus setiferus* probably functions to enable the male to find the sexually mature female during the time of mating. The occurrence of secondary sexual modifications of the antennular flagella are widespread among the Crustacea Decapoda. Meredith (1954) describes a modified outer antennular flagellum in the adult male of *Crangon vulgaris* and suggests that the structure may function in mating. This worker finds that the character is of use in identifying adult males in the field. The most extensive treatment of the subject of sexual dimorphism in Crustacea is that of Rioja (1939a, 1939b, 1940a, 1940b, 1940c, 1941a, 1941b, 1942a, 1942b), who has described in detail the sexually modified medial flagellum of *Penaeus setiferus* and several other species of penaeid shrimps. This worker considers the character to be constant in occurrence and sensory in function, and closely related to the sexual activities of the shrimps.

In the opinion of the present writer, the modified medial antennular flagellum is a constant

character in the sexually mature male of *Penaeus setiferus*. However, the subject needs further study. A large, statistically significant number of male and female *Penaeus setiferus* shown to be sexually mature by the histological methods of King (1948) should be examined to prove the point. In the present study 10 adult males and 8 adult females were considered in connection with the character.

MUSCLE ELEMENTS

The antennule of *Penaeus setiferus* contains 13 muscles, as contrasted with 12 for the caridean *Pandalus*, and 9 for the antennule of *Astacus*. Cochran (1935) lists 9 for *Callinectes*, including a double antennular promotor muscle and a double antennular remotor muscle. If these divided muscle bodies are actually 2 promotor and 2 remotor muscles, then the antennule of *Callinectes* can be said to have 11 muscles.

ANTENNULAR ABDUCTOR MUSCLE

FIGURE 14

The antennular abductor muscle of the first antennular segment is attached posteriorly to an apodeme that arises from the ventrolateral margin of the articular foramen by which the antennule is connected to the protocephalon. The plane of orientation of this apodeme is vertical. The antennular abductor runs anteriorly a brief distance and divides into two large branches that attach to sclerotized bars supporting the posterior region of the first antennular segment in the neighborhood of the statocyst. Contraction of the first segment abductor muscle swings the first antennular segment, and with it the distal antennular segments, outward from the mid-sagittal plane of the body in the limited horizontal motion possible to the first segment. The antennular abductor of *Penaeus* is homologous with the musculus promotor I antennae of *Astacus*, *Pandalus*, and possibly to the first segment promotor muscles of *Callinectes*, judging from the origins, insertions, and arrangements of the muscles.

However, the functions of the homologous muscles are different, as often happens. The difference lies primarily in the restriction of antennular movement created by the presence of the antennal scales ventral to the antennules in *Penaeus*. In the white shrimp, the scales are relatively large and, when lying in their usual longitudinal posi-

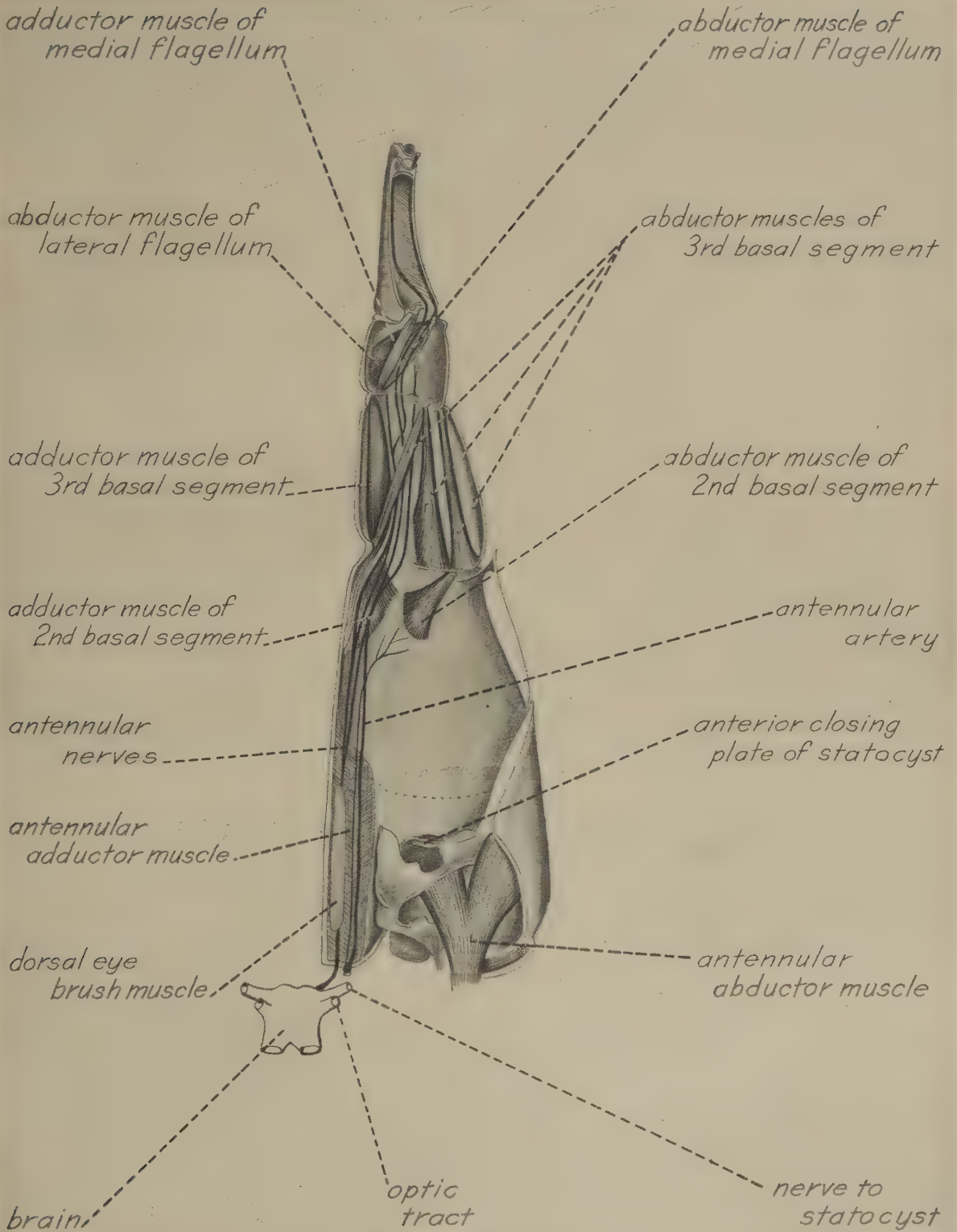


FIGURE 14.—Dorsal view of right antennule showing dorsal muscles. Dorsal cuticle removed.

tion, their mesial margins overlap beneath the antennules. Movements of the antennules ventrad are thereby prevented. In *Pandalus* and *Astacus* the first antennular segments may be raised and lowered about a transverse axis through the articular foramina of the antennules, in the sagittal plane. That Schmidt (1915) in his study of *Astacus* named the muscle a "promotor" is open to question. The muscle might better be described as a levator of the first antennular segment. Berkeley (1928) and Cochran (1935) adopted the terminology of Schmidt for this muscle in *Pandalus* and *Callinectes*, although, in *Callinectes* the musculus promotor I antennae apparently moves the antennule toward the midline of the animal.

ANTENNULAR ADDUCTOR MUSCLE

FIGURE 14

The antennular adductor muscle of the first antennular segment originates on a vertically-oriented apodeme arising from the ventromesial region of the articular foramen between the antennule and the protocephalon. The muscle courses anteriorly along the mesial margin of the first antennular segment, inserting in the cuticle at many points along the mesial edge of the first segment (fig. 14). The antennular adductor muscle functions to turn the first antennular segment toward the mid-sagittal line of the shrimp.

The musculus remotor I antennae described by Berkeley (1928) in *Pandalus*, by Schmidt (1915) in *Astacus*, and by Cochran (1935) in *Callinectes* are in all probability the homolog of the first segment adductor muscle in *Penaeus*. In the case of the former three animals, two muscoli remotor I antennae have been found in the antennule of each, a remarkable uniformity in animals as distantly related as these. *Penaeus*, too, has these muscles, but the dorsal-most, discussed in the following section, appears to have a function different from that of remotion or adduction.

DORSAL EYE BRUSH MUSCLE

FIGURE 14

The dorsal eye brush muscle, or prosartema muscle, lies upon the adductor muscle of the first antennular segment, and is almost certainly the homolog of the musculus remotor *b* I antennae found in the antennules of *Pandalus*, *Astacus*, and

possibly of *Callinectes*. However, instead of taking part in the adduction of the first antennular segment, the eye brush muscle in *Penaeus* serves to stiffen the dorsal eye brush (figs. 12, 13). Fibers of the muscle enter the eye brush at its point of attachment to the first antennular segment, and their contractions presumably enhance the function of the brush as an eye cleaner.

ABDUCTOR MUSCLE OF SECOND BASAL SEGMENT

FIGURE 14

The second antennular segment is turned away from the midline on its points of articulation with the first antennular segment by contractions of the abductor muscle of the second basal segment (fig. 14). This muscle originates on the anteroventral part of the first antennular segment at about the midpoint between the anterior margins of the segment. The muscle runs a short distance antero-laterally to insert on a posterolateral apodeme of the first antennular segment.

The second basal segment abductor muscle of *Penaeus* has a homologue in the musculus productor₂ I antennae of *Pandalus*, *Astacus*, and *Callinectes*, although the use of the term "productor" for the action of the muscle in *Callinectes* is questionable. In the blue crab musculus productor₂ I antennae is said by Cochran (1935) to pull the second antennular segment downward, indicating that the muscle functions as a depressor.

ADDUCTOR MUSCLE OF SECOND BASAL SEGMENT

FIGURES 14, 15

The adductor muscle of the second basal segment originates ventrally in the anteromesial corner of the first antennular segment and inserts on a small apodeme at the posteromesial corner of the second antennular segment (figs. 14, 15). Contraction of the adductor muscle turns the second antennular segment, together with the distal elements of the antennule, toward the mid-sagittal plane of the animal. The second basal segment adductor muscle of *Penaeus* has a counterpart in the musculus reductor₂ I antennae of *Pandalus*, *Astacus*, and *Callinectes*. The muscle functions in *Pandalus* and *Astacus* to depress the distal antennular segments and flagella, while in *Callinectes* the distal elements are raised toward the midline by the action of the musculus reductor₂. The muscle might better have been called a second segment levator in the blue crab.

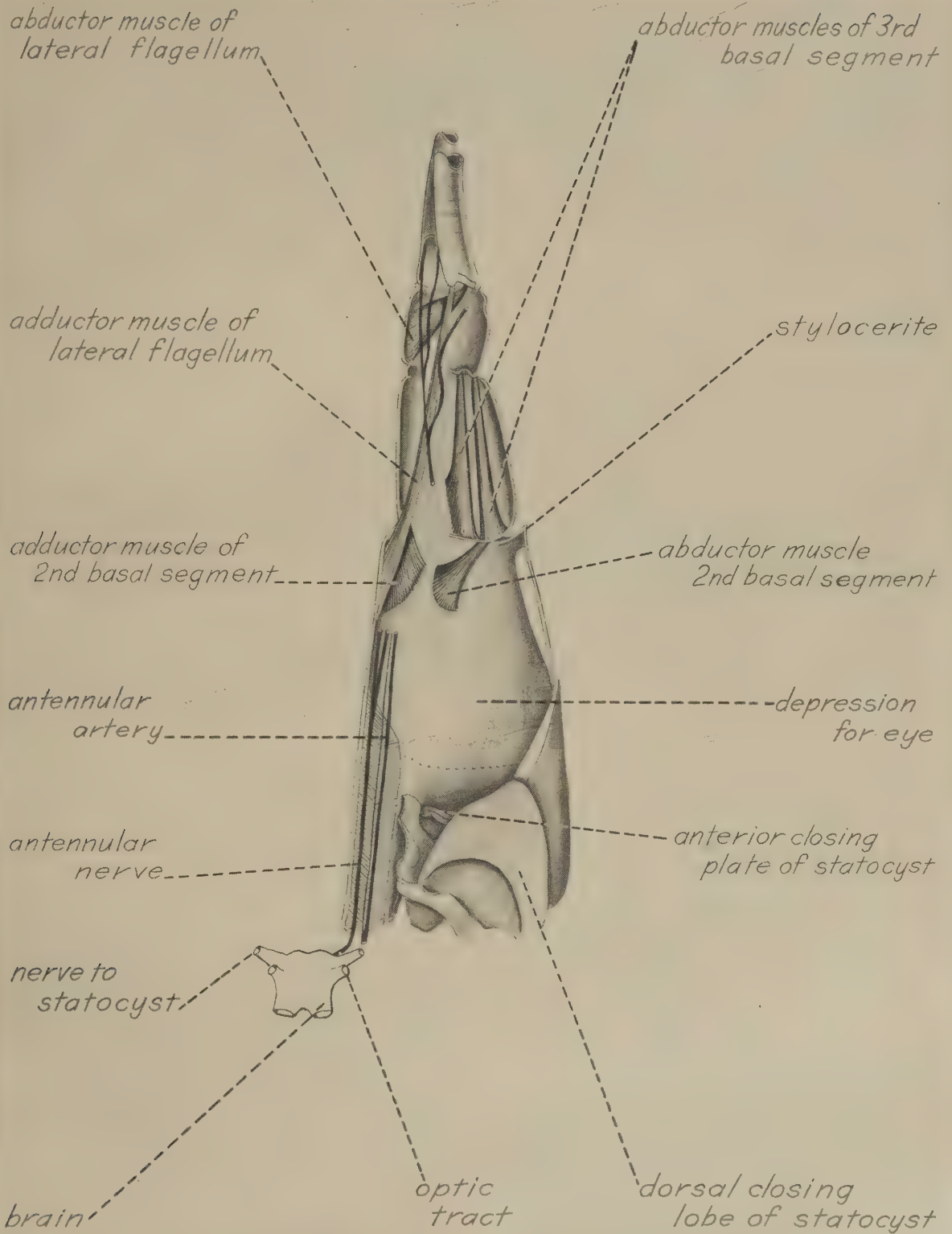


FIGURE 15.—Dorsal view of right antennule showing ventral muscles.

ADDUCTOR MUSCLE OF LATERAL FLAGELLUM

FIGURE 15

Arising from the anteromesial margin of the first antennular segment, just dorsal to the origin of the second basal segment adductor muscle, is the long, slender adductor muscle of the lateral flagellum. The muscle courses anteriorly through both the second and third antennular segments to insert on an apodeme arising from the mesial edge of the lateral flagellum base. Upon contraction of the lateral flagellum adductor the lateral flagellum is turned on its points of articulation toward the mid-sagittal plane. The muscle has a probable homolog in the musculus adductor I antennae of *Pandalus*, although in the latter the muscle inserts in the ventral part of the third antennular segment rather than on a flagellum. A lateral flagellum adductor muscle does not occur in the antennule of *Astacus* or *Callinectes*. Cochran (1935) describes a musculus adductor₂ I antennae for *Callinectes* that functions somewhat like the musculus adductor I antennae found by Berkeley (1928) in *Pandalus*, but which is not homologous with the latter muscle or with the lateral flagellum adductor muscle in *Penaeus*. A study of the nerves might show that the musculus adductor₂ I antennae in *Callinectes* is part of the musculus reductor₂ I antennae (second segment adductor muscle of *Penaeus*).

ADDUCTOR MUSCLE OF THIRD BASAL SEGMENT

FIGURE 14

The relatively large adductor muscle of the 3d basal segment originates in the posteromesial corner of the second antennular segment and runs directly anterior to insert on an apodeme in the posteromesial corner of the first antennular segment. Contractions of this muscle turn the third antennular segment and the flagella mesad, reinforcing the action of the proximal antennular adductors. The third basal segment adductor of *Penaeus* has homologs in the musculus reductor₃ I antennae of *Pandalus*, *Astacus*, and *Callinectes*. Cochran's (1935) use of the name "reductor" for the muscle in *Callinectes* is unfortunate. The reductor muscle of the third antennular segment in the blue crab appears to function virtually opposite to the muscle of the same name in *Pandalus* and *Astacus*, a situation that underlines the impropriety of transferring functional muscle names from one animal to another.

ABDUCTOR MUSCLES OF THIRD BASAL SEGMENT

FIGURES 14, 15

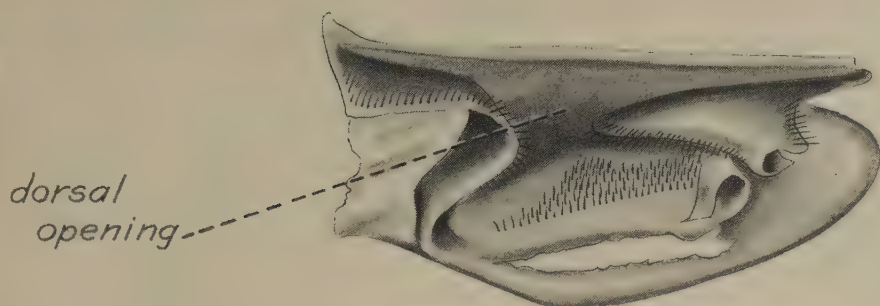
The second antennular segment of *Penaeus* contains three abductor muscles of the 3d basal segment. The lateral abductor muscle originates in the posterolateral corner of the second antennular segment and runs directly anterior to its point of insertion on a small apodeme of the posterolateral corner of the third antennular segment. Just mesad of the lateral, or first, abductor, a second abductor muscle originates broadly on the ventral surface of the posterior region of the second segment. This second abductor muscle courses anteriorly, parallel to the lateral-most abductor muscle, and inserts on a short apodeme slightly mesad of the insertion of the lateral-most muscle (figs. 14, 15). The mesial third basal segment abductor muscle originates in the posteromesial corner of the second antennular segment dorsal to the origin of the third basal segment adductor muscle. The mesial abductor runs diagonally to the anterior to insert on the apodeme of one of the lateral abductor muscles (fig. 14). Contractions of the third segment abductor muscles turn the third segment and the flagella away from the mid-sagittal plane. Their action reinforces that of the first and second segment abductor muscles.

The lateral-most abductor muscle of the third antennular segment of *Penaeus* is homologous with the musculus productor₃ I antennae of *Pandalus*, *Astacus*, and possibly *Callinectes*. In *Pandalus* and *Astacus* the musculus productor₃ swings the third segment dorsally in the sagittal plane, while in *Callinectes* the muscle named by Cochran (1935) a "productor" appears to flex the third segment. The second lateral third-segment abductor muscle in *Penaeus* has homologs in the musculus abductor₃ I antennae of *Pandalus* and *Astacus*. The muscle has been lost in *Callinectes*. The mesial third-segment abductor muscle of *Penaeus* does not appear in the antennule of any of the crustaceans to which reference has been made here.

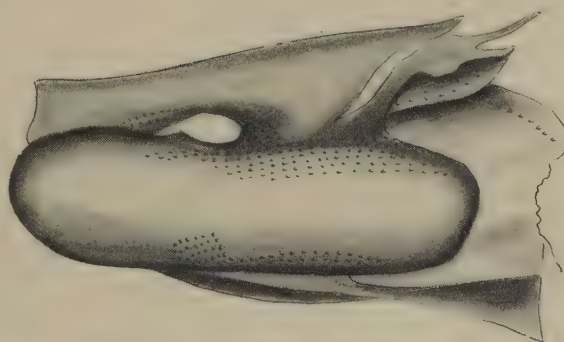
ABDUCTOR MUSCLE OF LATERAL FLAGELLUM

FIGURES 14, 15

The abductor muscle of the lateral flagellum has its origins along most of the mesial side of the third antennular segment. The muscle nar-



A. Anterior View



B. Posterior View

FIGURE 16.—Cleared statocyst. A. Anterior view. B. Posterior view.

rows as it runs anterolaterally, ventral to the lateral flagellum adductor muscle, to insert on an apodeme on the lateral edge of the base of the lateral flagellum (fig. 15). The lateral flagellum abductor muscle turns the lateral flagellum away from the midline in the horizontal plane. The muscle in *Penaeus* is very likely homologous with the musculus reductor₄ I antennae of *Pandalus*, *Astacus*, and *Callinectes*, although evolutionary rearrangements have given rise to several changes. Berkeley (1928) describes the reductor₄ muscle in *Pandalus* as consisting of two parts, inserting on two opposite margins of the base of the lateral flagellum, and having at times an antagonistic action to one another. Such action suggests very strongly the existence of two muscles, rather than one with two functions.

ABDUCTOR MUSCLE OF MEDIAL FLAGELLUM

FIGURE 14

Originating in the posteromesial corner of the third antennular segment is the small adductor

muscle of the medial flagellum. The adductor muscle courses anteriorly and inserts on an apodeme on the mesial edge of the flagellum base. Contractions of the muscle turn the medial flagellum toward the midline in the horizontal plane. No counterpart of this muscle in *Penaeus* is described for the antennule of *Pandalus*, *Astacus*, or *Callinectes*.

ABDUCTOR MUSCLE OF MEDIAL FLAGELLUM

FIGURE 14

The abductor muscle of the medial flagellum originates ventral to the origin of the medial flagellum adductor muscle and inserts on the medial flagellum by a short apodeme located at the lateral margin of the flagellum base. The muscle turns the medial flagellum outward from the midline in a horizontal plane. The medial flagellum abductor muscle of *Penaeus* has been lost in the antennule of the other crustaceans referred to in this study.

3. THE ANTENNAE

The antennae, or as they are frequently termed, the second antennae, display wide variations among the Crustacea. In certain of the Copepoda and other groups in the lower Crustacea they are large swimming organs. They are modified for clinging in other copepods. Frequently the antenna of the male crustacean is modified as a sexual clasper. Sexual dimorphism of the antennae is widespread in the lower Crustacea, the appendage of the adult male being more highly developed than that of the female. In several groups the structures may be extremely reduced or even lost in the adult animals.

The antenna of the higher Crustacea is fairly uniform, although exceptions do occur. The malacostracan antenna is said to consist of a 2- or 3-segmented protopodite, an endopodite of 2 or 3 segments, the distal-most bearing a flagellum, and an exopodite reflected into a flat scale. The protopodite usually is comprised of the coxopodite and basipodite, but some forms may have a proximal pre- or sub-coxa in addition (Calman 1909). Carcinologists use the expression "peduncle" to refer to the total number of endopodite and protopodite segments, usually 5 or 6, in the Malacostraca. Representatives of the lower Decapoda, like *Penaeus setiferus*, have a 5-segmented antennal peduncle, made up of the coxopodite and basipodite in the protopodite, and 3 proximal endopodite segments. The exopodite is scalelike. The scale is often missing or reduced to a spine in the higher decapods and the endopodite and flagellum may be relatively small.

Functionally, the antennae have always been thought to be sensory. More specifically, the antennal flagella are said to be centers of tactile sensation, whether pressure or simple touch is not known. No experimental data to support these contentions exist. The functions of the antennal scale would appear to be varied, in accordance with its great variation in size. The scale is missing in the Brachyura and small in the crawfishes and other Astacura, making its function somewhat difficult to determine by observational means. The organ is very large in the swimming decapods of the Natantia. The suggestion has frequently been made that the scale is an anterior swimming plane in the latter group.

Some attempt has been made to ascertain the function of the antennal scale in the swimming

of the white shrimp. Normally the scale is carried in a directly anterior position, rotated slightly on its longitudinal axis. The rotation is such that the thin mesial margins of the scale lie somewhat ventrad of the heavy lateral margins. The effect produced is that of a ship's bow. Shrimps in an aquarium were lightly secured by a loop of string to the end of a rod. Lifted from the bottom in this position, the animals are stimulated to swim forward. Jets of water from a pump were directed at the motionless shrimps from various directions to see whether the antennal scale was used as an anterior steering device. No compensatory movements of the scale were observed. The organs appeared simply to cleave the water ahead. During normal movements about an aquarium, the scales are occasionally spread widely, but are never kept in the spread condition for more than a moment.

The antenna in *Penaeus setiferus* is typical of the natant Decapoda. The antennal scale is broad and strong and extends as far as the anterior tip of the rostrum. The protopodite is comprised of two segments, a short, incomplete coxopodite and a very large basipodite. To the large basipodite is articulated the exopodite, or scale, and the basal segments of the antenna. The size of the basipodite reflects its support of the heavy scale rather than that of the smaller antennal segments. The long antennal flagellum is carried laterally, from which position the movements of the shrimps through the water cause the flagellum to drag alongside and some distance behind the animal. The flagellum is approximately twice the body length in the white shrimp.

SKELETAL ELEMENTS

The first antennal segment, or coxopodite (fig. 18), is an incomplete ring by which the antenna is attached to the protocephalon. The foramen of the antenna at which the first antennal segment articulates with the protocephalon is by far the largest of the head foramina. At its broadest portions, muscles insert upon the first antennal segment.

The basipodite, or second antennal segment (figs. 18, 23), is a large, heavily sclerotized article connected firmly to the first antennal segment proximally and articulating distally with the third antennal segment, or ischiopodite, of the endopodite, and with the antennal scale, or exopodite. At

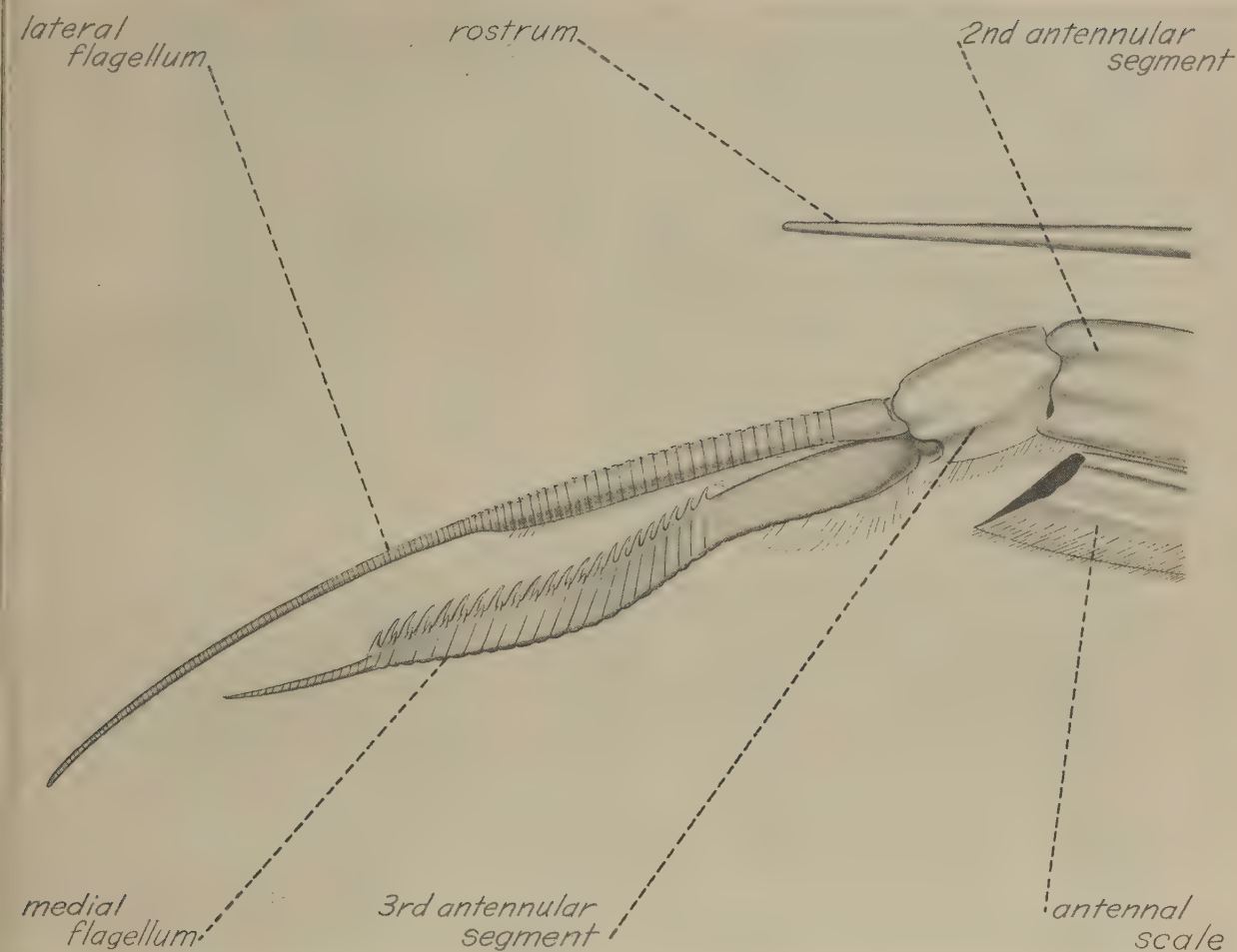


FIGURE 17.—Enlarged view of left antennular flagella showing sexual modification to medial flagellum in adult male.

the point of articulation with the antennal scale, the second antennal segment is deeply notched anteriorly providing an articular foramen for the large antennal scale. Strong dorsal and ventral points of articulation, or condyles (figs. 18, 22, 23) on the rim of the scale foramen permit the scale considerable horizontal movement. As brought out earlier, the great weight and strength of the second antennal segment exists as support for the antennal scale, rather than for the long, slender antennal flagellum. The second antennal segment articulates with the third antennal segment, or ischiopodite, by a small foramen located ventro-mesially (figs. 18, 23). The extent of movement between the second and third antennal segments is very limited.

The third antennal segment (figs. 18, 23) is a small, heart-shaped structure whose apex is directed posteriorly. Its small vertical movement,

at right angles to its long axis, against the second antennal segment permits some rotation of the distal antennal segments. Movements of the little shield-shaped fourth antennal segment (figs. 18, 23) against the third antennal segment are, on the other hand, extensive. The fourth antennal segment attaches laterally to the third segment at about a 45° angle. By means of a dicondylic articulation, the fourth segment rotates through an arc of nearly 90° , and with it the distal antennal parts.

According to Schmidt (1915), the third antennal segment represents the fusion of the ischiopodite and meropodite and he evidently has morphological support for this view in *Astacus*. Curiously, no trace of a division is apparent in the third antennal segment of *Cambarus*, nor does Berkeley (1928) find one in the caridean *Pandalus*. No evidence for the fusion of the ischiopodite and meropodite can be seen in *Penaeus setiferus*.

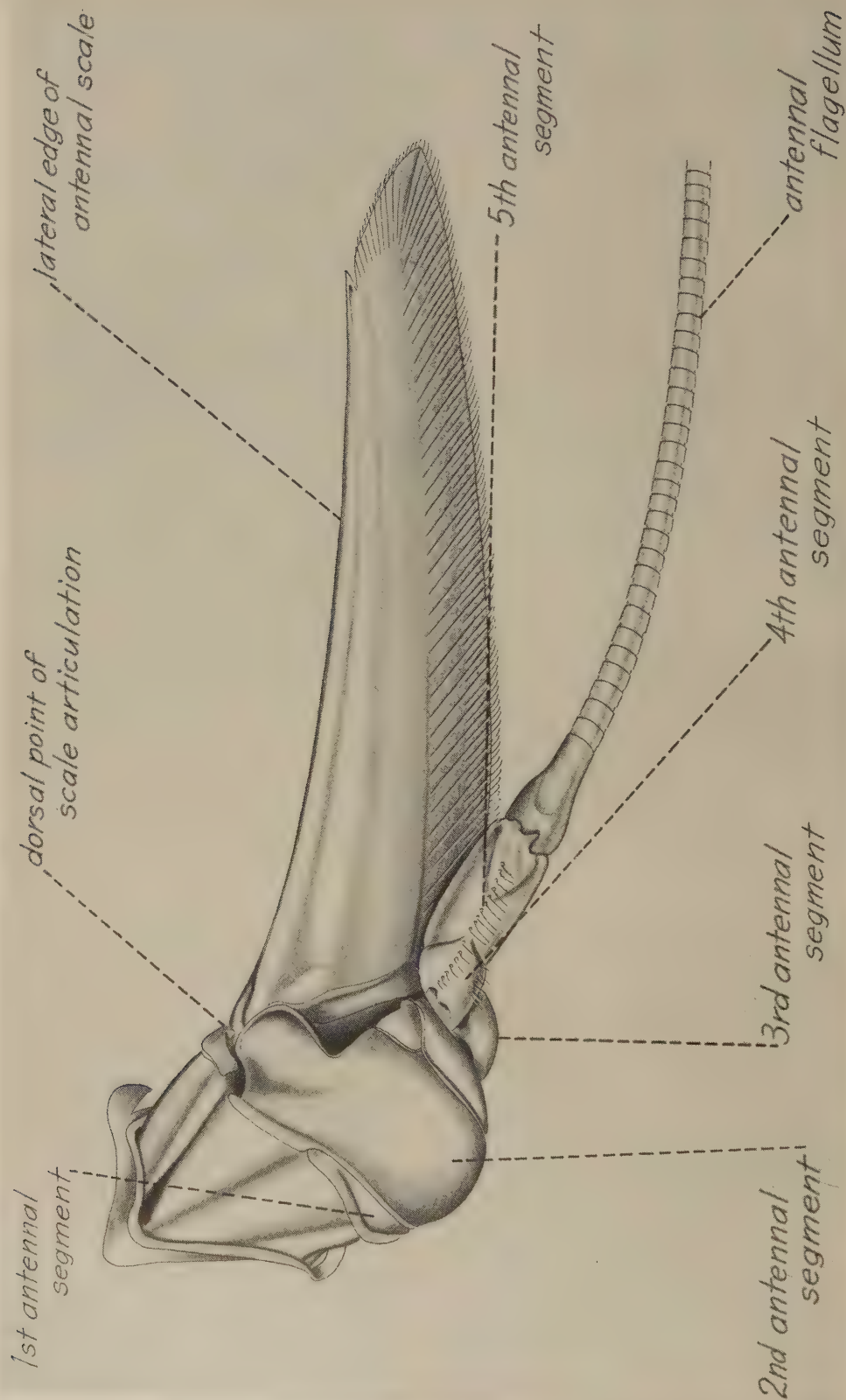


FIGURE 18.—Lateral view of right antenna.

The fifth antennal segment (figs. 18, 23) lies on the axial line distal to the fourth antennal segment and is connected to the pointed fourth segment by a broad V-shaped surface in the proximal end of the fifth segment. Two condyles in the vertical plane located at the apex of the V-shaped surface permit limited horizontal movements of the fifth segment on the fourth antennal segment. The relatively large fifth antennal segment bears the base of the long flagellum on its distal end. Strong dorsoventral condyles allow the flagellum to turn through an arc of more than 90° and the size of the fifth segment is probably an evolutionary response to the long muscles needed to operate the flagellum. In living shrimps, the distal part of the flagellum is carried at right angles to the fifth antennal segment, the rest of the long flexible flagellum floating behind the animals. The antennal flagellum (figs. 23, 25) owes its flexibility to its annular construction. Each small annulation is capable of a little movement with respect to its neighbors.

An enlargement of the flagellar rings (fig. 25) shows probable sensory structures. On the dorsal surface of each ring (fig. 25, A) may be seen a pair of dorsal setae. On the ventral surface (fig. 25, B) a pair of plumose ventral setae project anteriorly from the distal portion of each annulation. Between the bases of the ventral setae is a ventral pit. The interior of the flagellum consists of blood vessels and nerves.

MUSCLE ELEMENTS

The skeletal parts of the antenna of *Penaeus setiferus* are operated by 12 types of muscles, including at least 26 individual muscles. Berkeley (1928) describes 15 types of muscles in *Pandalus*. Schmidt (1915) lists 18 muscle types in the European crawfish, the appendage containing 21 separate muscles to carry out its complex movements. The reduced antenna of *Callinectes* has only 8 muscles (Cochran 1935). The evolutionary trend of reduction of the number of muscles and muscle types has apparently been reversed in the case of the crawfish antenna, in which form much more complicated antennal activities are displayed by the living animals than in *Penaeus* or *Pandalus*.

FIRST ANTENNAL SEGMENT ADDUCTOR MUSCLE

FIGS. 21, 24

Attaching on the median rim of the coxopodite foramen, the first antennal segment adductor mus-

cle inserts in the first antennal segment near the external opening of the excretory apparatus (fig. 21). Contractions of this short, powerful muscle turn the coxopodite and hence the antenna mesad. Homologs of this muscle in *Astacus* and *Pandalus* are difficult to determine without full information on the nerves. Allowing for functional differences in the antennae of the various crustaceans here considered, the most likely homolog of the first segment adductors in *Astacus* is the musculus depressor *a* II antennae. Berkeley (1928) designates two medial antennal base muscles as the musculus depressor *a* and *b* II antennae, after Schmidt (1915). From her illustrations of the antenna in *Pandalus*, the antennal depressor muscle *a* appears to be the same muscle as the first antennal segment adductor in *Penaeus*. The muscles in the latter two forms are similar, strongly suggesting an homology. The antennal depressor muscle *a* in *Astacus* is much less suggestive of phylogenetic similarity.

SECOND ANTENNAL SEGMENT PROMOTOR MUSCLES

FIGS. 19, 22

Far and away the heaviest musculature of the anterodorsal region of the white shrimp is that concerned with the antennae. The dorsal-most of these is a large, flat muscle originating in connective tissue slightly laterad of the postrostrum (fig. 19). This muscle, a second antennal segment promotor muscle, runs anteriorly and laterally to insert in what appears to be a free apodeme just beneath the dorsal rim of the coxopodite. This apodeme is not connected to the coxopodite, but instead (fig. 19) consists of a transverse fascia in which the distal second antennal segment promotor muscle originates. The presence of the free apodeme of the proximal second segment promotor muscle may indicate that the muscle is in reality a first antennal segment (coxopodite) promotor. The free apodeme, however, produces a functional second antennal segment (basipodite) promotor and the muscle is therefore so described. The proximal second segment promotor is evidently the homolog of the musculus promotor II antennae in *Astacus*, *Pandalus*, and *Callinectes*, although in these forms the muscle clearly attaches to the dorsal margin of the coxopodite.

The distal second antennal segment promotor muscle extends the functional connection of the muscle group to the dorsal edge of the second an-

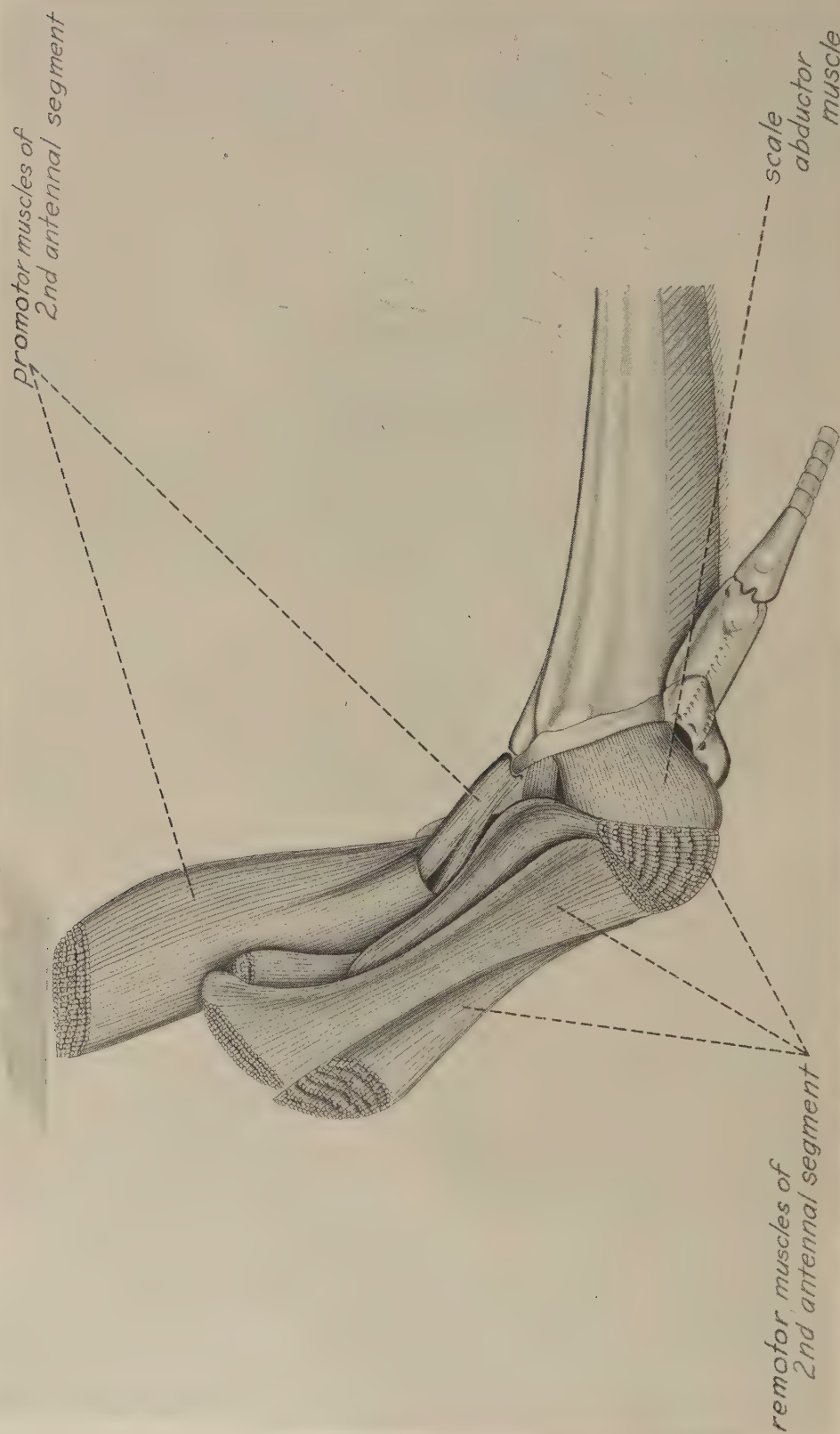


FIGURE 19.—Lateral view of right antenna, carapace and cuticle removed to show muscles.

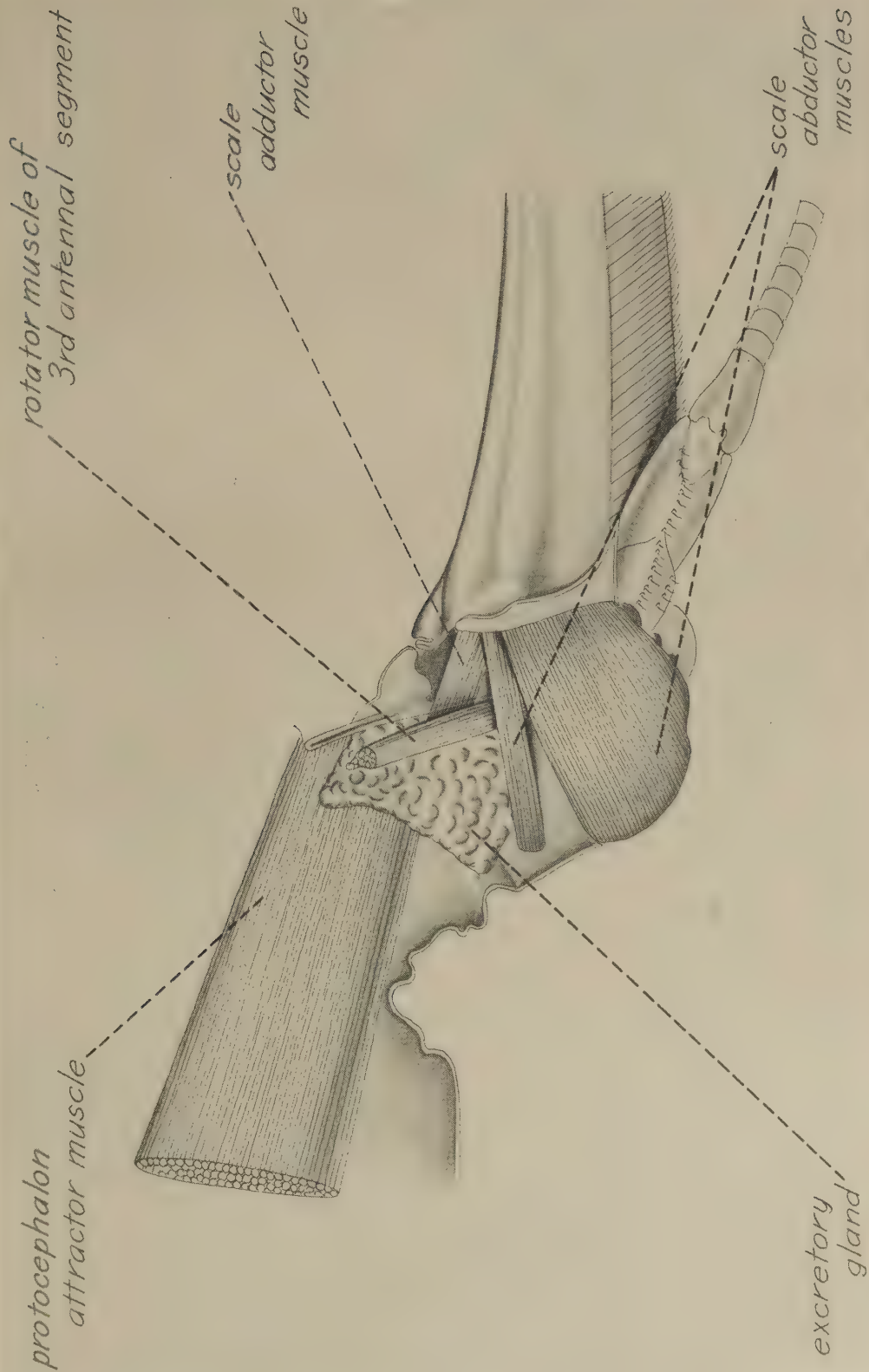


FIGURE 20.—Lateral view of right antenna. Lateral muscles removed to show excretory gland.

tenal segment, or basipodite. Upon contraction the promotor muscles raise the antennal scale and endopodite segments dorsally. The muscles actually function as levators. Without information about the innervation, no homolog of the distal promotor muscle is here suggested.

SECOND ANTENNAL SEGMENT REMOTOR MUSCLES

FIGS. 19, 24

At least three large antennal muscles arise on the dorsolateral carapace just anterior to the hepatic spine. These muscles, here considered the second antennal segment remotor muscles (figs. 19, 24), run anterolaterally to extensive insertion areas in the lateral and posterior regions of the second antennal segment (basipodite). The lateral- and posterior-most of these muscles might be thought to insert on the ventral margin of the first antennal segment, in which case the two muscles would be first segment, or coxopodite, remotor muscles. Repeated dissections in the area indicate otherwise, however, and for this reason the muscles are assigned to the basipodite. The posterior-most remotor displays a definite torsion as it passes from its lateral point of origin to a posterior and even slightly medial insertion area. While the function of all three remotor muscles is actually the depression of the large scale, that of the posterior remotor muscle may also include adduction of the scale, together with depression. Since the antennal scale of *Penaeus* bears a major portion of the weight of water striking the anterior end of the animal, large remotor (depressor) muscles are needed to maintain the scale in position.

The homologies of the second antenna segment remotors is made confusing by the functional muscle nomenclature. A comparison of the function of the antennae in the crawfish and *Penaeus* shows wide differences. The crawfish antennal segments are constructed to permit extension movements of the comparatively short, stiff flagellum. One kind of extreme of this modification has been achieved in the antenna of *Palinurus*, in which form movability is combined with great size and power for the protection of the animal. As has been suggested, the antennal movements of *Penaeus* are comparatively limited by virtue of segmental architecture, particularly in the segments of the protopodite. The heavy musculature of carapace origin is in reality associated

with the simple movements of the scale; the muscles of the distal antennal segments of the endopodite are comparatively small.

As a consequence of the many antennal functions in different crustaceans, homologous muscles have different functions and nomenclature. At least part of the second antennal segment remotor muscle mass in *Penaeus* is undoubtedly homologous with the musculus remotor II antennae in *Astacus*, *Pandalus*, and *Callinectes*. Part also may be homologous with the musculus depressor *c* II antennae in *Astacus* and perhaps even with a part of the large musculus depressor *c* II antennae in *Pandalus*, although certainly the major part of the antennal depressor *c* in *Pandalus* is the protocephalon attractor muscle. The other antennal depressor muscles, *a*, *b*, and *d* (*d* is not found in *Pandalus*) in *Astacus* and *Pandalus* are not evident in *Penaeus*.

SCALE ABDUCTOR MUSCLES

FIGURES 19, 20, 24

Taking origin from large areas to the posterior, ventral, and medial region of the second antennal segment (basipodite), the proximal scale abductor muscles (figs. 19, 20, 24) insert on the lateral margin of the antennal scale (exopodite), lateral to the dorsoventral scale condyles. In addition to the huge ventral scale abductor, at least two and probably three small-scale abductors (figs. 20, 24) are found in the second antennal segment of *Penaeus*. A long, distal scale abductor muscle (fig. 24, *B*), originating in the distal part of the scale, runs proximally along the lateral margin of the scale to insert on the lateral margin of the basipodite foramen, lateral to the axis of the scale condyles. When the scale abductor muscles contract, the large scale is swung laterally some distance. The functional reason for this movement is not clear. Shrimps in an aquarium occasionally spread the scales, at times in association with cleaning activities of the head appendages and a sudden flushing out of the gill chamber.

The proximal scale abductor muscles of *Penaeus* are probably homologous with the second antennal exopodite abductor muscles *a*, *b*, and *c* in *Astacus* and with the single exopodite abductor in *Pandalus*. The scale is reduced in *Callinectes*. Schmidt (1915) does not show a distal scale abductor muscle in *Astacus* such as exists in *Penaeus*, and Berkeley (1928) makes no mention of the muscle

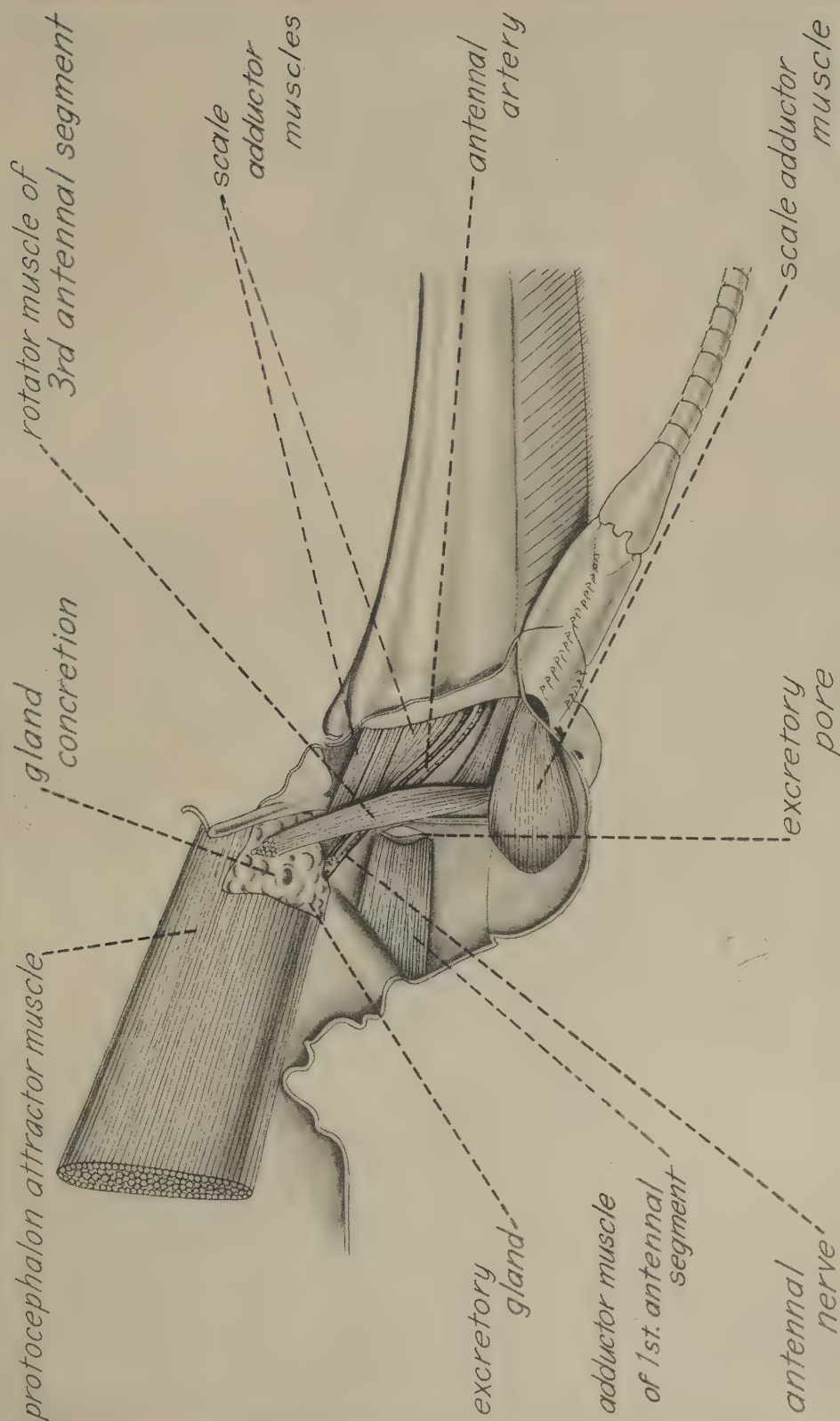


FIGURE 21.—Lateral view of right antenna. Lateral muscles removed to show medial muscles, nerves, and arteries.
Part of excretory gland removed to show gland concretion.

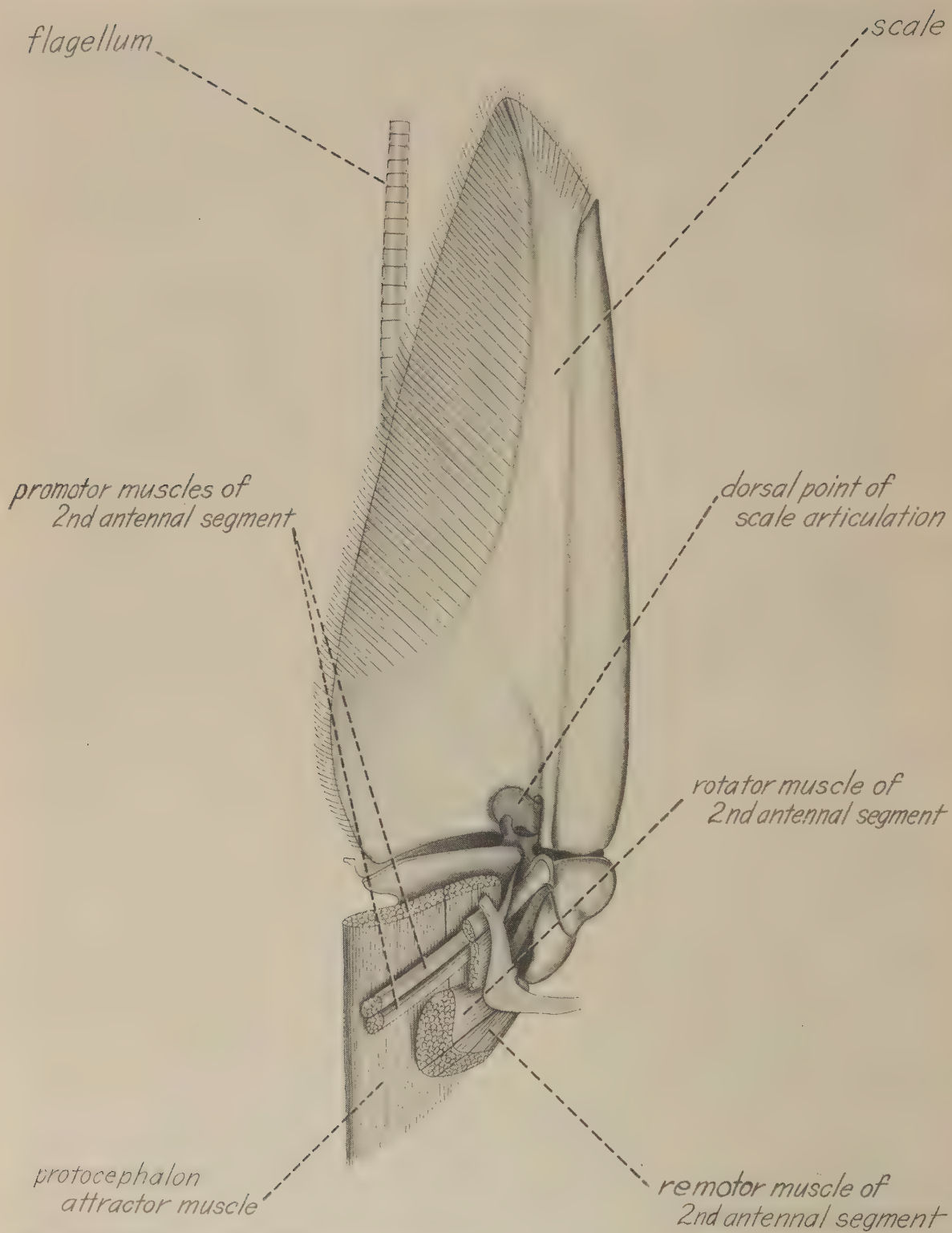


FIGURE 22.—Dorsal view of right antenna showing antennal scale and protopodite muscles. Carapace removed.

in *Pandalus*. However, Berkeley illustrates a distal scale muscle which she designates as the musculus adductor exopoditis *b* II antennae and which from the standpoint of its arrangement appears to be the distal scale abductor muscle in *Penaeus*. A careful review of the insertion of Berkeley's exopodite adductor muscle might show that it is in reality an abductor muscle.

SCALE ADDUCTOR MUSCLES

FIGURES 20, 21, 24

Two types of scale adductor muscles (figs. 20, 21, 24) are found in the antenna of *Penaeus*. At least two scale adductors originate on the medial wall of the second antennal segment ventral to the excretory pore (fig. 24, *B*), and run diagonally to insertion points on the ventral and medial margins of the scale foramen. Their insertions are mesad of the axis of the scale condyles. The distal scale adductor muscle (fig. 24, *B*), like the distal scale abductor, is located in the body of the scale. It originates in the distal region of the exopodite and runs caudad parallel to the distal scale abductor to insert on the margin of the basipodite foramen mesad of the scale articles. Upon contraction, the scale adductor muscles move the antennal scale inward toward the median line of the shrimp, in opposition to the action of the scale abductors.

THIRD ANTENNAL SEGMENT ROTATOR MUSCLE

FIGURES 20, 21, 24

Arising on the dorsomedial rim of the first antennal segment (coxopodite) foramen and running ventro-medially, the third antennal segment rotator muscle (figs. 20, 21, 24) inserts on an apodeme located on the lateral margin of the third antennal segment (figs. 23, 24). Contractions of the muscle accomplish the movement described in the section on the skeletal elements, namely, lateral rotation of the third antennal segment and the antennal parts distal to the third segment.

The homologs of this muscle in the other crustaceans referred to are not clear. The best possibility in *Astacus* is one or both of the meropodite muscles *a* and *b*. On the basis of area of insertion, the most likely homologs is the musculus meropoditis *a*, although the third segment rotator in *Penaeus* and the meropodite muscle *a* in *Astacus* have different origins, the latter being on the basipodite of the crawfish antenna. The

meropodite muscle in *Pandalus* is more nearly similar to the situation in *Penaeus* than in *Astacus*. In both *Astacus* and *Pandalus*, Schmidt (1915) and Berkeley (1928) illustrate a muscle, the musculus reductor ischiopoditis II antennae, said to oppose the action of the meropodite muscles *a* and *b*. No similar muscle has been found in *Penaeus*, although the shrimp may have a functional analog in the fourth antennal segment adductors.

FOURTH ANTENNAL SEGMENT ADDUCTOR MUSCLE

FIGURE 24

The fourth antennal segment adductor muscle (fig. 24) originates in a broad fan on the ventral surface of the basipodite. The muscle runs anterodorsally, narrowing to its apex at its point of insertion on a small, movable article presumed to be a part of the fourth antennal segment. Certainly a muscle originating on the basipodite and inserting on the fourth antennal segment is unusual. Controversy could be avoided by assigning the small, movable article to which this muscle inserts to the third antennal segment; however, the movable article appears to be widely separated from the third segment and instead is lateral to the fourth segment and clearly connected to it. When the fourth segment adductor muscle contracts, the movable article is drawn posteroventrally with the result that the fourth antennal segment is rotated upon the third segment and turned a short distance mesad. As such, the fourth segment adductor represents in part an opposing muscle to the third segment rotator muscle. The homology of this muscle is uncertain.

FOURTH ANTENNAL SEGMENT ABDUCTOR MUSCLE

FIGURE 24

Originating from a broad area slightly posterior to the origin of the fourth segment adductor muscle, the fourth antennal segment abductor muscle (fig. 24) runs to an insertion on the same small movable article of the fourth antennal segment to which the fourth segment adductor attaches. The fourth segment abductor is much larger than the fourth segment adductor. Upon contraction the fourth antennal segment abductor reinforces the action of the fourth segment ad-

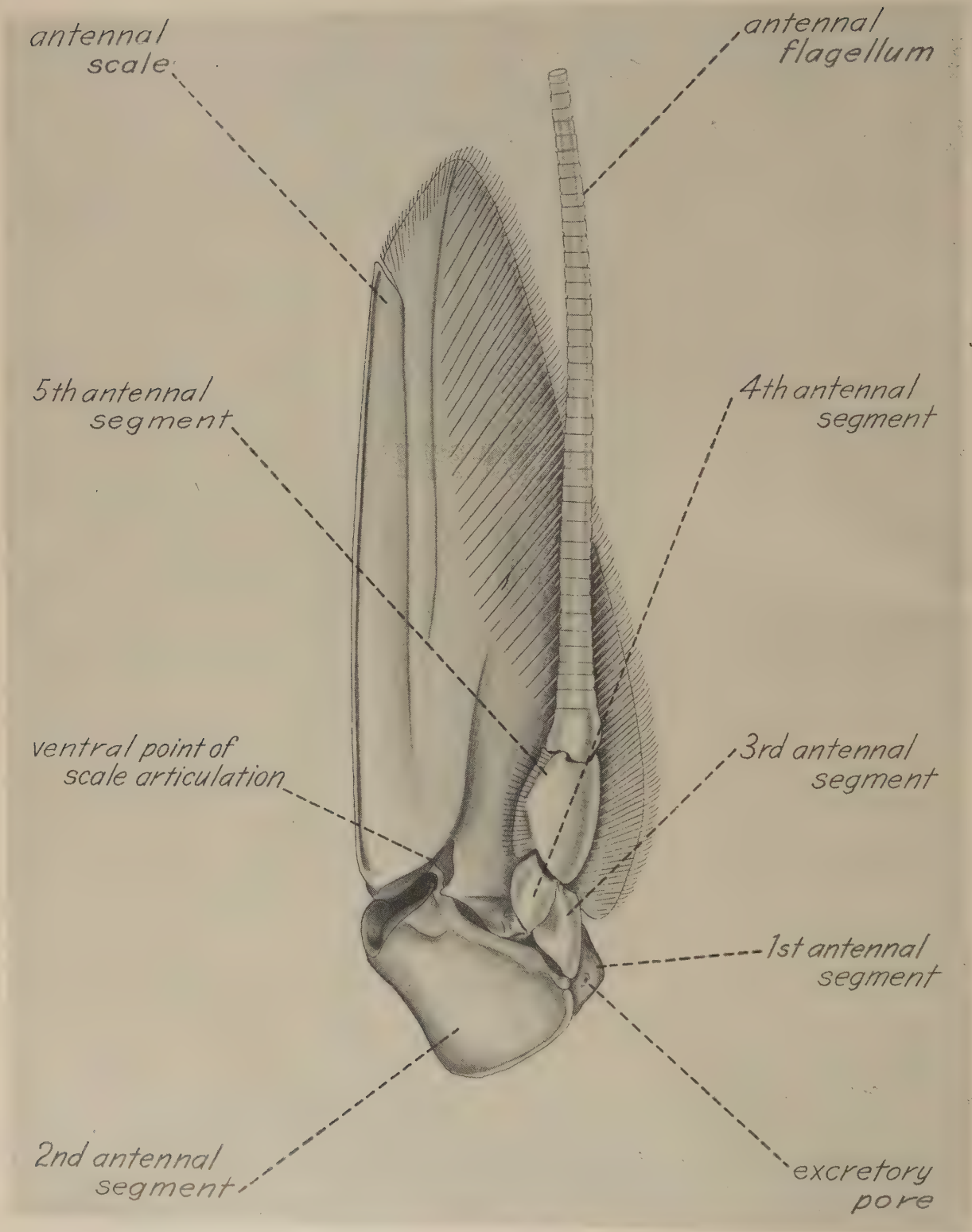


FIGURE 23.—Ventral view of right antenna showing skeletal elements.

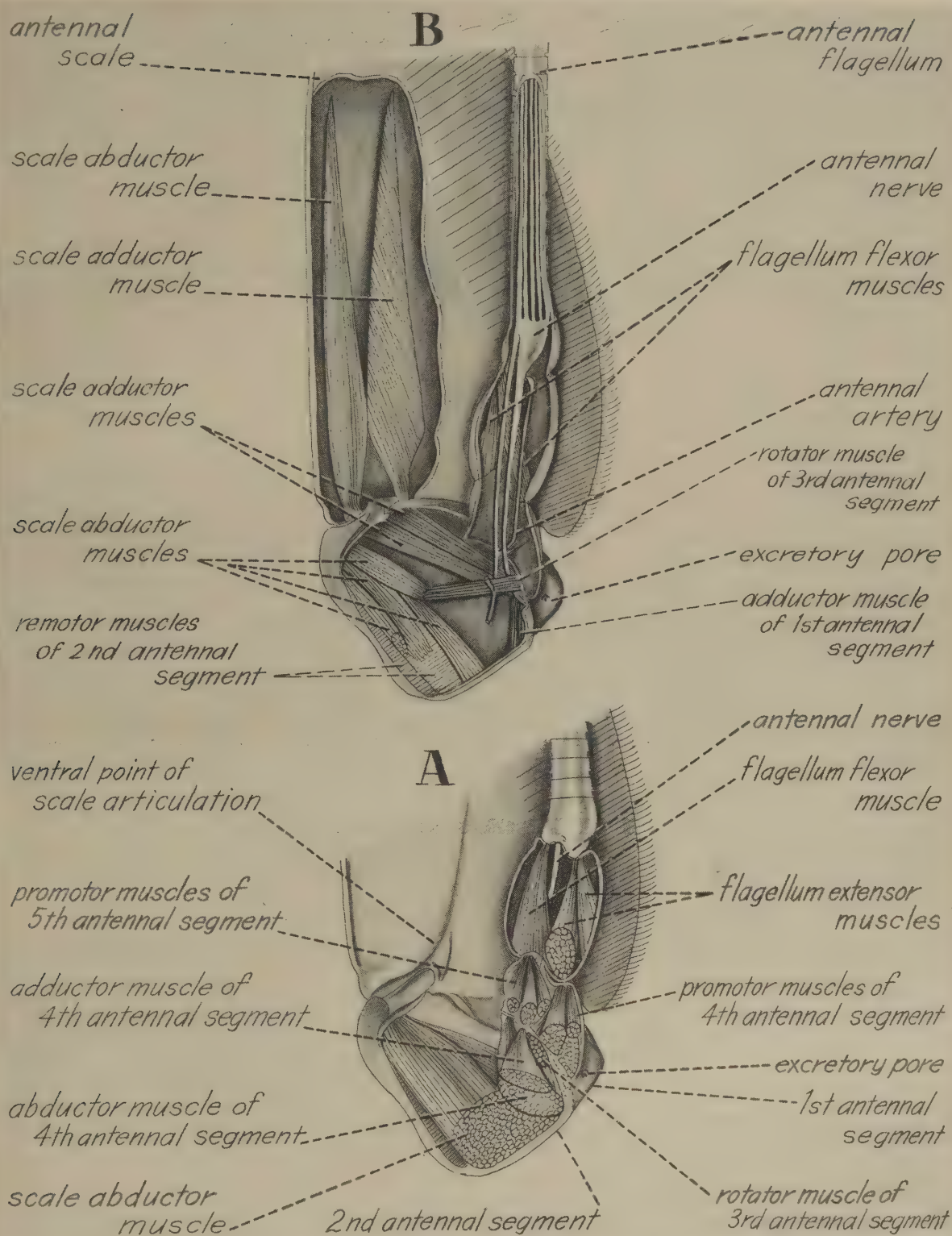


FIGURE 24.—Ventral view of right antenna showing ventral muscles, nerves, and blood vessels. Ventral cuticle removed. A. Superficial ventral elements. B. Dorsal elements.

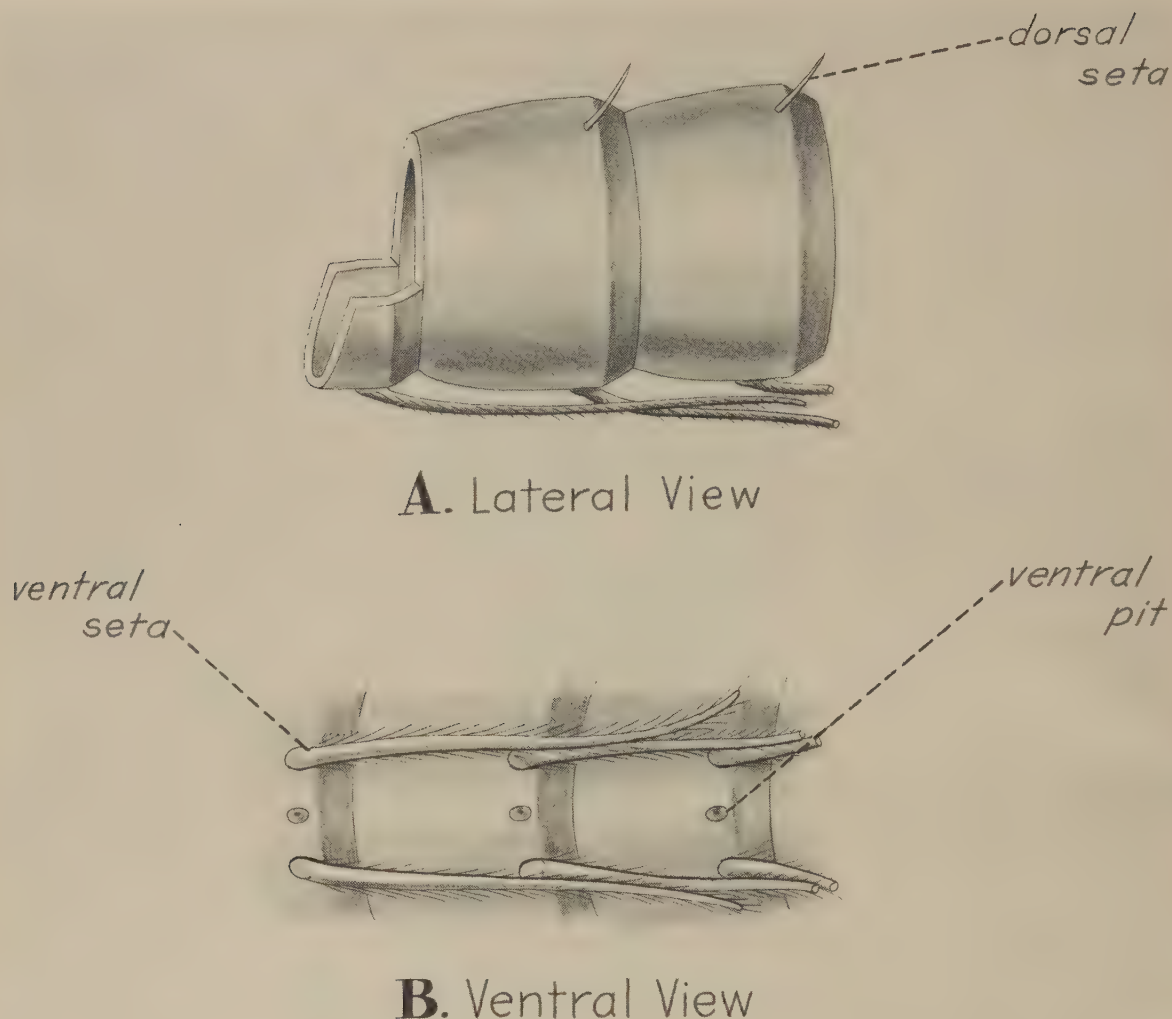


FIGURE 25.—Enlargement of proximal segments of antennal flagellum showing sensory structures.

ductor muscle in rotating the fourth segment on the third segment. In opposition to the fourth segment adductor, the fourth segment abductor turns the fourth antennal segment laterad a short distance. No homolog of this muscle is here advanced.

FOURTH ANTENNAL SEGMENT PROMOTOR MUSCLES

FIGURE 24

The fourth antennal segment promotor muscles (fig. 24) are situated within the body of the third antennal segment. At least three of these muscles occur in *Penaeus*. The short, thick promotors originate throughout the ventral surface of the third antennal segment and insert at their

narrow apical tips on an apodeme of the fourth antennal segment. Contractions of the fourth antennal segment promotors move the fourth segment and the distal antennal parts anteriorly. How these muscles are represented in the other crustaceans referred to in this paper is uncertain.

FIFTH ANTENNAL SEGMENT PROMOTOR MUSCLES

FIGURE 24

The fifth antennal segment promotor muscles (fig. 24) are comprised of a tuft of at least four small muscles restricted to the fourth antennal segment. The muscles originate on the lateral and posterior margin of the fourth article and attach to an apodeme arising from the posterior

groove of the fifth antennal segment. Contractions of the fifth segment promotors move the fifth antennal segment anteriorly in a limited way. The homolog of these muscles in *Astacus* is very like the musculus extensor propoditis II antennae. Berkeley (1928) does not find the muscle in *Pandalus*.

FLAGELLUM EXTENSION MUSCLES

FIGURE 24

The fifth antennal segment of *Penaeus* contains two flagellum extensor muscles (fig. 24), occupying the medial half of the segment. One of the flagellum extensors originates on the medial side of the fifth segment and runs distally to insert on the large extensor apodeme on the medial side of the flagellum base. The second extensor takes origin on the posteroventral region of the fifth segment, runs distally, and inserts on the large extensor apodeme on the flagellum. These muscles bring the base of the flagellum directly anterior to the proximal antennal segments. One or both of these muscles is undoubtedly homologous with the antennal dactylopodite extensor muscle as shown by Schmidt (1915) in *Astacus* and by Berkeley (1928) in *Pandalus*. The related muscle, if any, in *Callinectes* is uncertain without adequate information about the nerves.

FLAGELLUM FLEXOR MUSCLES

FIGURE 24

Three flagellum flexor muscles in the fifth antennal segment of *Penaeus* turn the antennal flagellum to its normal position at right angles to the proximal antennal segments. The largest and ventral-most of these muscles originates broadly along the proximal groove of the fifth segment and inserts on the flexor apodeme on the flagellum base. Dorsal to the large muscle, two flagellum flexor muscles insert on the same flexor apodeme on the flagellum. The dorsolateral flagellum flexor originates in the lateral corner of the fifth segment, the dorsomedial flexor originating in the medial corner of the fifth antennal segment. At least one of these muscles in *Penaeus* is the homolog of the musculus flexor dactylopoditis II antennae in *Astacus* and *Pandalus*. Whether homologs exist in *Callinectes* is not known.

4. LABRUM

The labrum is the final component of the procephalon to be considered. In all arthropods the

labrum is a lobe or sac suspended over the mouth from a sclerotized region of the head known as the epistome (the hexapod clypeus). Crustacean morphologists ordinarily do not consider the labrum an appendage. Most workers consider the labrum an unpaired structure. Some students of Crustacea place the epistome as the ventral element of a preoral, premandibular segment, behind the eyes, antennules, and antennae, in the order of their occurrence in many adult crustaceans. Others have even assigned the epistome to the sternum of the antennal or mandibular segments, giving the labrum an utterly indefensible postoral position.

The position of the epistome and labrum in adult arthropods is variable. In some crustaceans, like the adult isopods and amphipods, and in most insects, the epistome is anterior or facial (Snodgrass 1951). The labrum thus is ventral or anterior to it. However, in most crustaceans, some chilopods, and a few insects the head segments have thrust forward, overgrowing the epistome anteriorly. The result is a secondary ventral position of the epistome and labrum in some arthropods. In point of fact, the labrum is the anterior end of the arthropod. I am in full agreement with Snodgrass (1951) in this view. Furthermore, the labrum is here considered not only the most anterior part of the arthropod, but also the anterodorsal "upper lip" of these forms, and as such the dorsal part of the first segment. Considerable support for this interpretation has been adduced by Ferris (1947) and Henry (1948a), based on the study of the labral nerves.

If we accept the view of various workers, including Henry (1948a), that the tritocerebrum of arthropods is in reality the first ganglion of the ventral nerve cord, whatever its fate in the adult, then we are bound to regard structures innervated by tritocerebral nerves as primitively anterior. Since the labrum of *Penaeus* is clearly innervated by a pair of nerves from the tritocerebral ganglia (figs. 27, 76), similar to the situation in other Crustacea and Insecta (Henry 1948a, 1948b), then the labrum is segment 1 in *Penaeus*.

No evidence is here advanced to suggest that the labrum is a reduced appendage. Indeed, this ancient "upper lip" was probably never an arthropod appendage in the true sense at any time in its history. The labrum is, however, very likely paired. In support of this is the morphology of the nerves and muscles. The labral nerves are paired and arise from the clearly bilateral trito-

cerebral ganglia. Each nerve enters the sac of the labrum and innervates only those sensoria and muscles in half the labrum from the medium sagittal plane laterad. In *Penaeus* no nerve can be seen crossing the median sagittal plane. Furthermore, the embryonic labrum of many higher arthropods (Johannsen and Butt, 1941; Young 1953) develops as a paired structure, with paired coelomic sacs.

SKELETAL ELEMENTS

FIGURES 26, 27

In *Penaeus setiferus* the labrum is a soft, lightly sclerotized sac attached between the widely spread posterolateral bars of the epistomal Y (fig. 28, A). It may be noted in passing that these lateral epistomal bars are morphologically anterior, although due to rearrangements of the head segments, discussed above, the position of the epistome is reversed in the head of many crustaceans. The anterior or medial bar of the epistome, dividing the antennal foramina, curves anterodorsally to form a deep ventral pit, best seen in lateral view (figs. 28, A; 30) of a cleared anterior skeleton cut along the median sagittal plane. To the epistomal invagination, or apodeme, is attached a pair of muscles to be discussed below.

The labrum is shaped to fit between the antennal bases anteriorly and the incisor and molar surfaces of the mandibles posteriorly (figs. 26, A, B). Various auricles and lobes project from the labral surface to enhance its function as an aid in feeding. A toothed structure, the posterior feeding process (figs. 26, 27), projects directly into the mouth aperture.

MUSCLE ELEMENTS

EPISTOMAL STATOR MUSCLES

FIGURES 5, 6, 30, 34

Originating on the dorsal surface of the carapace, lateral to the posterior protocephalon levator muscles, and converging on the anterior side of the epistomal invagination is a pair of small muscles which are named in the present work the epistomal stator muscles (figs. 5, 6). The name derives from the fact that contractions of the muscles would appear to hold the epistomal invagination in position during the contraction of other muscles in the area. The epistomal stator muscles are homolo-

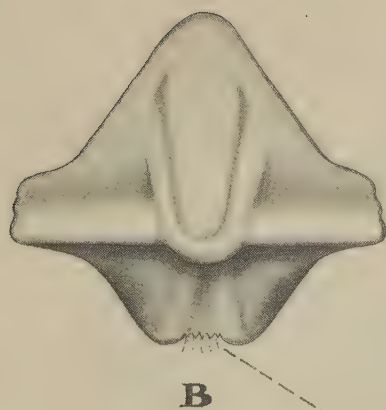
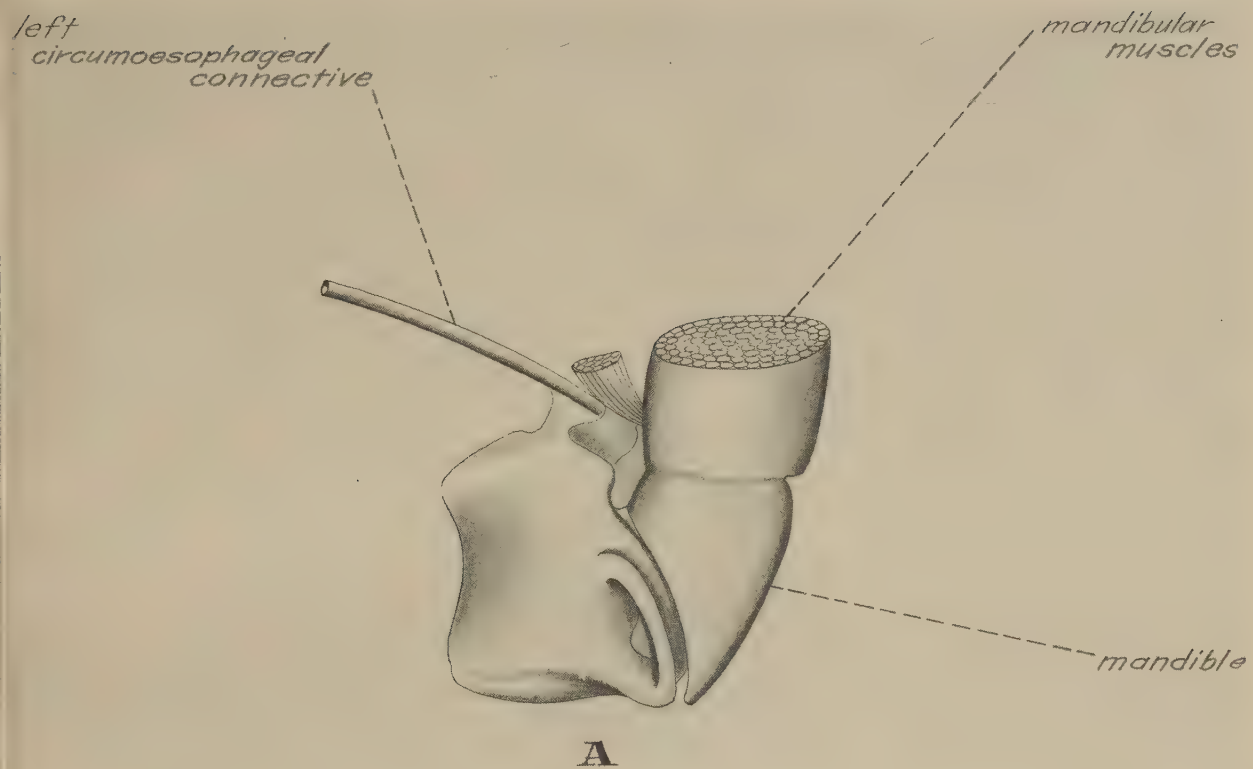
gous with the musculus oculi basalis posterior in *Pandalus*, *Astacus*, and probably in *Callinectes*.

Giving the name, epistomal stators, to these muscles may be adding confusion to the morphological scene, since these muscles are undoubtedly the musculus attractor sincipitis described by Grobben (1919) in the stomatopod *Squilla mantis* and the muscle attracteur du synciput illustrated by Mayrat (1955, 1956a, 1956b) in *Praunus flexuosus* O. F. Müller. The "sincipit" (Mayrat spells it "synciput") of Grobben (1917) is of course the protocephalon of Snodgrass (1951) as applied to the Crustacea. No great objection is offered here to designating the muscles in *Squilla* and *Praunus* as the synciput attractors. The muscles are indeed synciput or protocephalon muscles apparently functioning in certain forms to draw the protocephalon posteriorly. However, the same muscles in *Penaeus* do not attract the protocephalon. Furthermore they insert on a specific region of the protocephalon, the epistome, and so deserve as special a name as possible. The problem can be resolved by a study of the nerves, for if the epistomal stator muscles belong to the epistome, they should be innervated by epistomal or clypeal nerves.

LABRAL MUSCLES

FIGURE 27

One of the most astonishing features of the anatomy of *Penaeus setiferus* is the musculature of the labrum. In the generalized insect labrum, the structure is moved by two pairs of extrinsic muscles, the anterior and posterior labral muscles arranged for production and reduction. To the intrinsic labral compressor muscles of insects may be assigned various functions. In contrast, a review of general and special accounts of the anatomy of Crustacea has shown no reference to labral muscles in this class. Yet the labrum of *Penaeus* (fig. 27) is operated by at least 12 pairs of intrinsic muscles, bilaterally situated, and at least 1 intrinsic muscle running across the entire lobe. From their arrangement, the labral muscles obviously distort the labrum in all sorts of ways in the function of the organ as a tongue. In addition, at least 2 pairs of extrinsic muscles insert on the edge of the labral foramen to move the entire organ. No attempt has been made here to assign functional muscle names to the individual labral muscles.



posterior feeding process

FIGURE 26.—Labrum. A. Lateral view of labrum from the left side showing relation of labrum to mandible. B. Ventral view of labrum showing posterior feeding process.

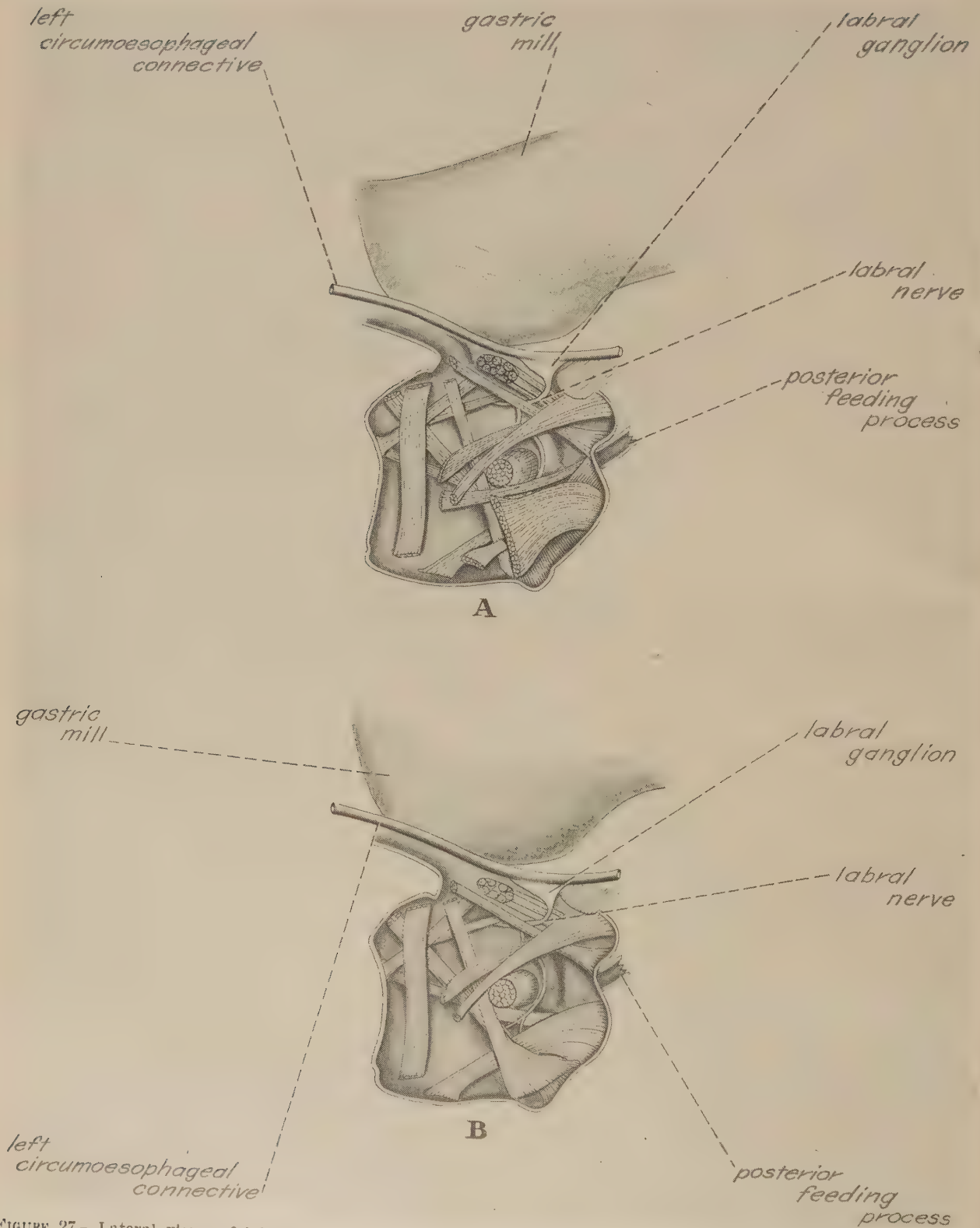


FIGURE 27.—Lateral views of labrum from the left side showing muscles and nerves. A. Cuticle removed to show superficial lateral muscles of left side. B. Interior muscles of labrum.

Although not shown in the illustrations of the labrum, material of a glandular nature is found in median ventral regions of the structure. Gland cells in the crustacean labrum have been described in the past.

B. Gnathothorax

The gnathothorax will be considered here as those segments following the protocephalon involved primarily with feeding and walking. The latter are not truly separable as body tagmata in the Crustacea, since after the mandibles and maxillae, varying numbers of walking legs may be adapted for feeding (Snodgrass 1951). Walking legs adapted for feeding are referred to as maxillipeds. In the Crustacea Decapoda, the gnathothorax is comprised of the mandibles, paragnatha, 2 pairs of maxillae, 3 pairs of maxillipeds, and 5 pairs of walking legs. In the present study, the gnathal segments will include the maxillipeds, and to the thorax will be assigned the five walking legs, without implying any morphological rigidity to the division. Above and to the sides, the gnathothorax is protected by the large dorsal shield, or carapace. Nearly all trace of segmentation has disappeared from the dorsal regions of the gnathothorax and the carapace. The few remaining sutures and markings of the carapace are not well understood in the Crustacea; consequently systematic nomenclature which has grown up around these devices is highly artificial.

The gnathothorax is constructed in the form of a rather special box arranged to provide both rigidity and movability. The immobile carapace is heavily sclerotized for protection of the internal organs and support of muscle origins, and extends ventrad in a deep fold of the tergum to cover completely the laterally placed gills (figs. 30, 31). The lateral carapace is called the branchiostegite, since the structure forms a chamber for the gills. The deep fold of the rigid carapace forming the branchiostegite permits movement between the carapace and the architecture of the ventral gnathothorax. The whole is reminiscent of a modern sedan in which a rigid body above is attached to a chassis able to respond to imperfections in the road surface. Upon the dorsal carapace originate numerous important muscles, including some of those of the protocephalon appendages, the mandibles, maxillae, gastric mill, and dorsal and ventral abdominal muscles.

Compared to the ventral skeleton of Astacura and Brachyura, that of *Penaeus* is very lightly sclerotized (Snodgrass 1952; Huxley 1906). In the crawfishes, the median sternal elements are rigidly fused together, except for the sternum of the last thoracic segment, and thus provide a rigid keel from which the pleurosternal arms arise. Pleural (laterotergal) and sternal apodemes arise from the invaginations between the fused arms of two adjacent segmental units. Similar apodemes occur in the same locations in the ventral skeleton of *Penaeus*, but the ventral sternal element of each segmental unit is separated from its neighbors by a transverse slit of thinly developed cuticle (fig. 28). The slits permit movements between segments along the anteroposterior axis of the thorax, even though the lateral pleurosternal arms are fused.

The ventral skeleton of the gnathothorax in *Penaeus* (figs. 28, 30) consists of a series of sclerotized units which are slightly movable with respect to one another, but not articulated (fig. 28). Each unit bears the paired foramina and muscle apodemes of the jointed appendages attached thereto. The typical segmental unit contains contributions from two sources: The tergum, in the form of the dorsal tergum, and the vertical, laterotergal pleural plates (Snodgrass 1952), dorsal to the leg bases, and the sternum which comprises the ventral region between the leg bases. The dorsal condyles of the coxopodites are situated on the laterotergal plates while the ventral coxopodite condyles are sternal. In the anterior region of the gnathothorax of *Penaeus*, the pleural plates lie horizontally to become the roof of the gill chamber, similar to the arrangement in *Cambarus longulus* Girard as shown by Snodgrass (1952).

The delicate nature of the ventral skeleton in *Penaeus* is even more noticeable in lateral view (fig. 28, A). From here the pleurosternal arms may be seen to bifurcate in the pleural region and those of each segmental unit tend to unite dorsally in the form of a reverse-curve, or ogee, arch of the architect. Again, compared to the composite pleura of the crawfishes (Snodgrass 1952), those of *Penaeus* appear to have been retained in a somewhat more generalized condition, with 7 or 8 pleura clearly distinguishable from one another (figs. 28, 30).

Huxley (1906), Calman (1909), and other students of Crustacea often refer to the system

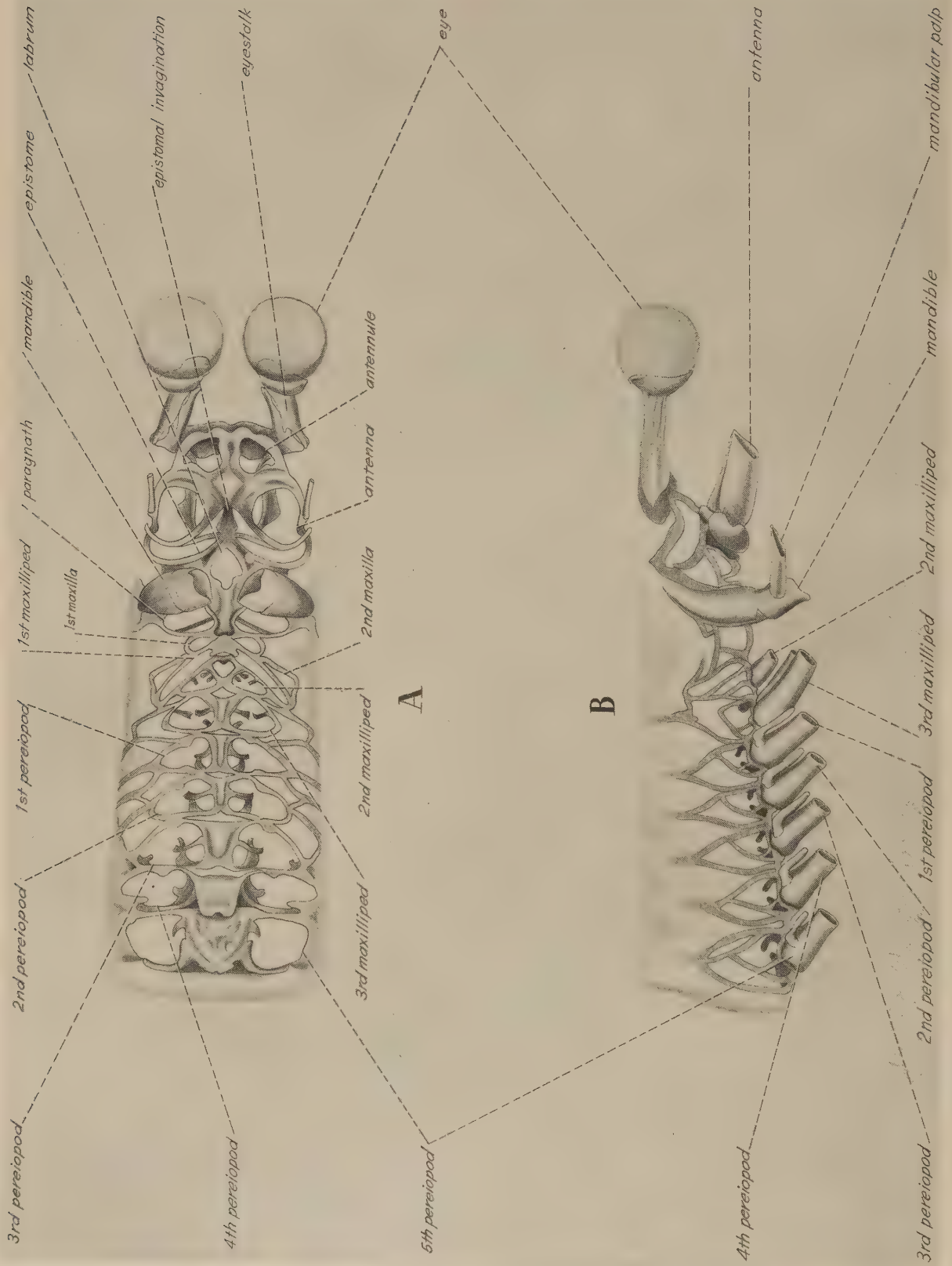


FIGURE 28.—Ventral skeleton of protocephalon and gnathothorax. A. Ventral view. B. Lateral view of right side.

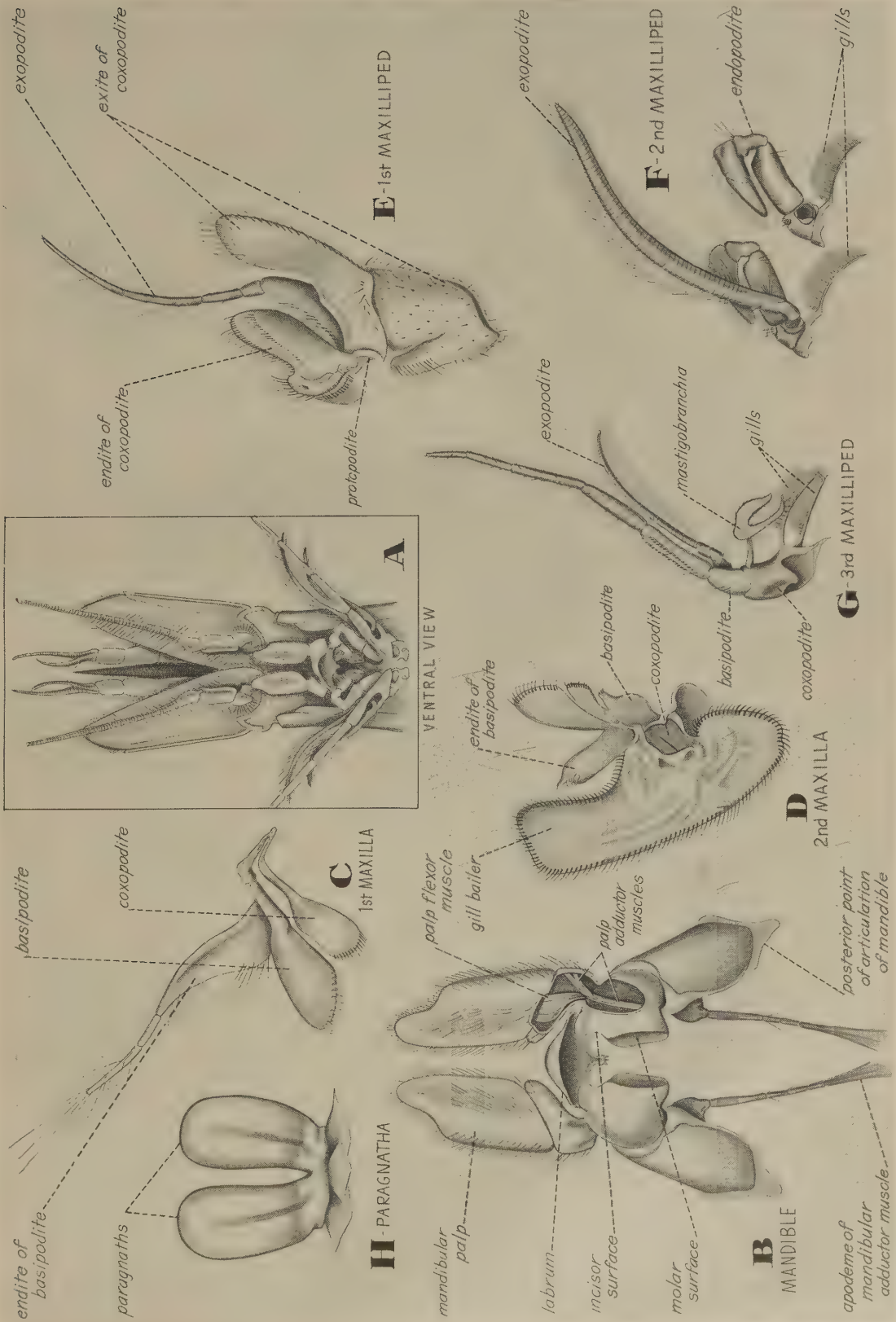


FIGURE 29.—Mouthparts.

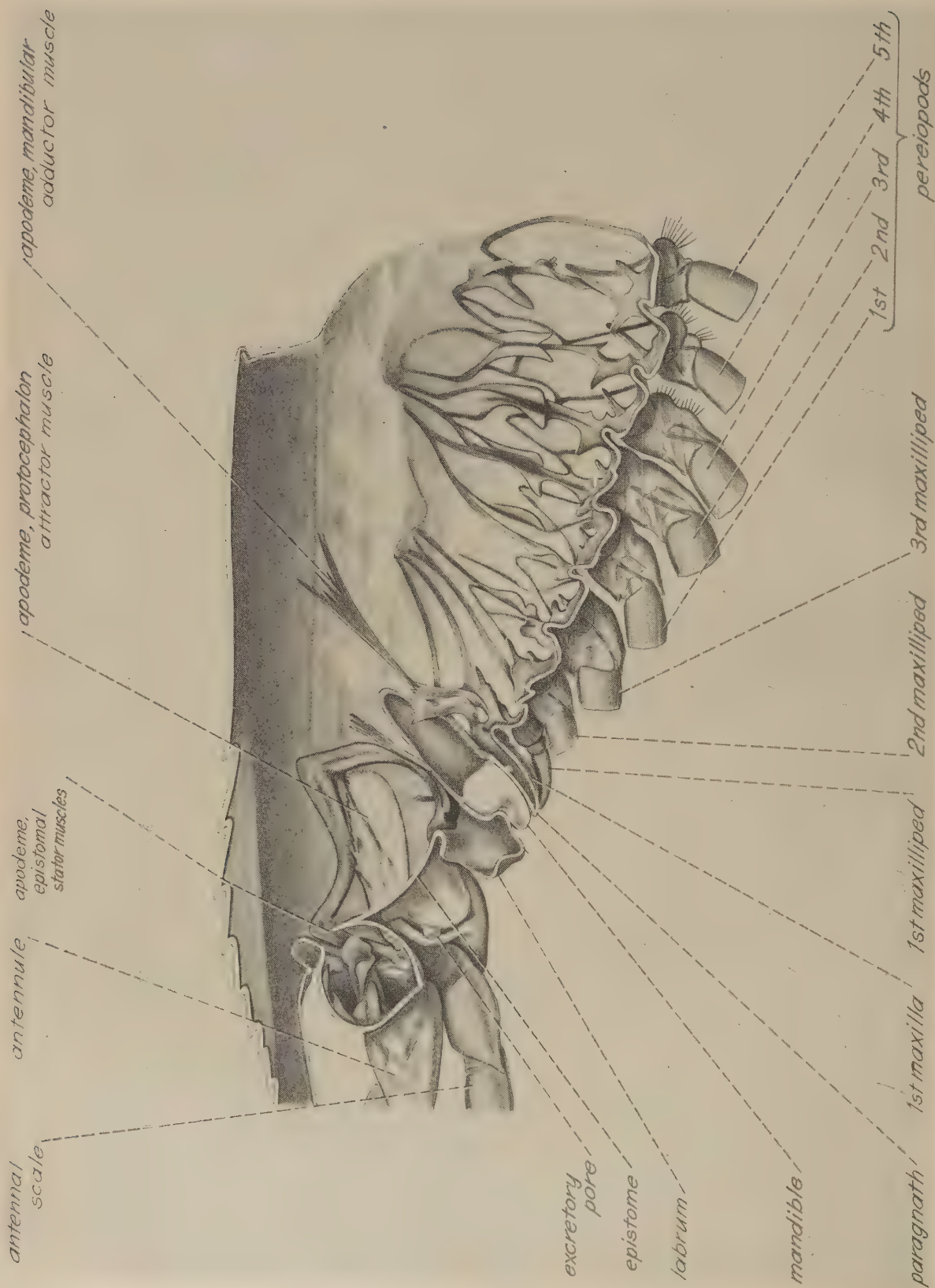


FIGURE 30.—Internal view of right half of protocephalon and gnathothorax.

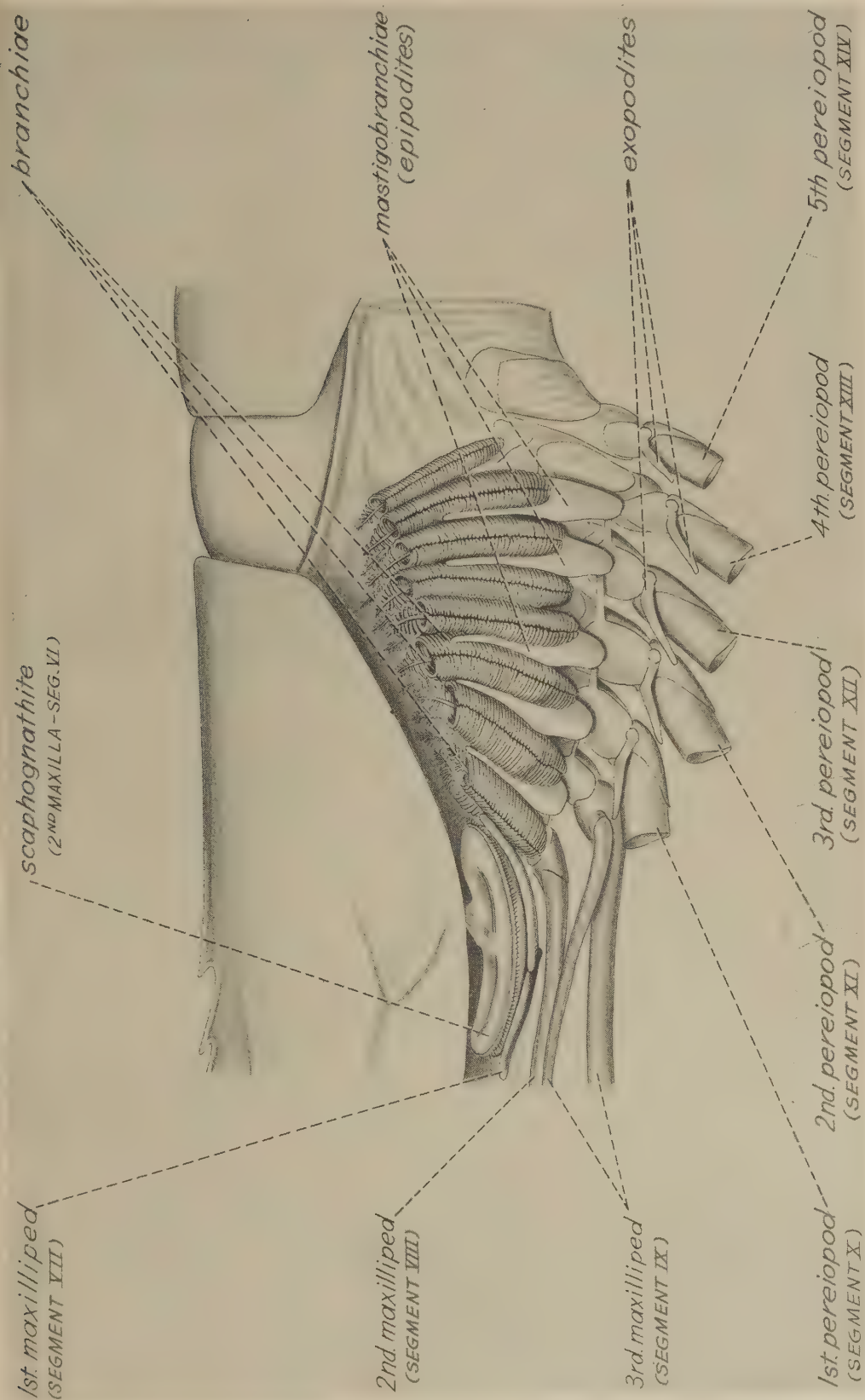


FIGURE 31.—Lateral view of left side of thorax, carapace cut away to show gill chamber, gills, and mastigobranchiae.

of pleurosternal invaginations or apodemes in decapods as the endophragmal system. In *Astacura* and *Brachyura* these apodemes fuse internally to form complex endoskeletons consisting of septate structures in the thoracic segments above the ventral nerve cord. The sternal apodemes fuse in the midline to form the sternal furca in the Insecta. In the gnathothorax of *Penaeus* the laterotergal and sternal apodemes are light and do not fuse, consequently no endophragmal system is found, unless we consider the transverse mandibular apodeme (see endosternite, fig. 38) an endophragm of some sort. This structure will be considered more fully in the treatment of the mandibles.

Although the ventral sternal elements of the penaeid gnathothorax are not coalesced into a rigid keel as in *Astacura*, the sternal plates do broaden from the anterior to the posterior ends of the gnathothorax, and abruptly so in the last three thoracic segments. The sternal plates of the last two thoracic segments are particularly modified in the female to receive the spermatophore from the male (figs. 28, *B*; 89). These structures will be discussed in detail in the section on the reproductive organs, page 155.

Gills

The gills may be exposed by cutting away the branchiostegal region of the carapace along the dorsal-most reaches of the inner lining of the branchiostegite, where the lining joins the laterotergal plates. The gills are thereby found to occupy a chamber (fig. 31), open to the outside ventrally and posteriorly by a narrow slot between the leg bases and thoracic wall on the inside and the extreme margin of the branchiostegite on the outside. The chamber is closed dorsally by the branchiostegal fold. The chamber is rather shallow transversely in its broad, posterior region, but becomes narrow anteriorly and is made much deeper in the region of the second maxilla by the lateral and horizontal reflection of the laterotergal, pleural plates. This narrow, deep, anterior chamber is thus a funnel, closed dorsally by the pleural bridges, medially by the vertical pleural wall, laterally by the branchiostegite, and ventrally by the large, flat coxopodite exites (figs. 29, *E*; 42) of the first maxilliped. Inside the funnel resides a pump, the scaphognathite of the second maxilla (figs. 29, *D*; 31; 41). Details of

its mechanical action will be considered when the scaphognathite muscles are described, page 69.

The tightly packed gills or branchiae rise roughly dorsad from their points of origin on the leg bases and pleura (fig. 31). Each gill consists of an axial circulatory rachis from which the individual gill filaments branch. Details of gill structure will be given below in the section on respiration. Interspersed among the gills are six flat, setose, bilobed structures, the mastigobranchiae or epipodites. The lateral margins of the mastigobranchiae may be seen upon removal of the branchiostegite (fig. 31), but they are best seen if the gills are removed (fig. 32). If a shrimp whose branchiostegite has been removed is cooled so that the body processes are reduced but not stopped, the long, fine setae of the mastigobranchiae may be seen to beat in phase with the movements of the scaphognathite, or gill bailer, thus suggesting that the epipodites play a part in the water flow and cleaning of the gills.

The older literature of Crustacea abounds in so-called branchial formulae, the formal study of gill origins. In the decapods the generalized situation is 4 gills for each side of the segment (fig. 32); 1 arises from the laterotergal, pleural plates (pleurobranchia), 2 from an articular element dorsal to the coxopodite (arthrobranchia), and 1 from the coxopodite (podobranchia) (Calman 1909; Snodgrass 1952). The epipodite (mastigobranchia) arises from the coxopodite. As shown by Calman (1909), the use of the gill origins as evolutionary landmarks is limited by the practical difficulty of distinguishing between the pleurobranchiae and arthrobranchiae in different species, since ontogenetic changes of gill origin are frequently seen. In fact, evidence exists suggesting that all the gills develop from the appendages, rather than from the limb bases or body wall. Apparently in *Penaeus* the podobranchiae develop embryologically from the mastigobranchiae.

As may be seen in figure 31, the branchial formula for *Penaeus setiferus* consists of 1 tiny arthrobranchia on the first maxilliped (fig. 42); 1 podobranchia, 1 mastigobranchia, and 2 arthrobranchiae on the second maxilliped; 1 mastigobranchia, 2 arthrobranchs, and 1 pleurobranch on the third maxilliped, and the first, second, and third pereopods; 1 arthrobranchia and 1 pleurobranchia on the fourth pereopod, and 1 pleurobranch on the fifth walking leg. In a European

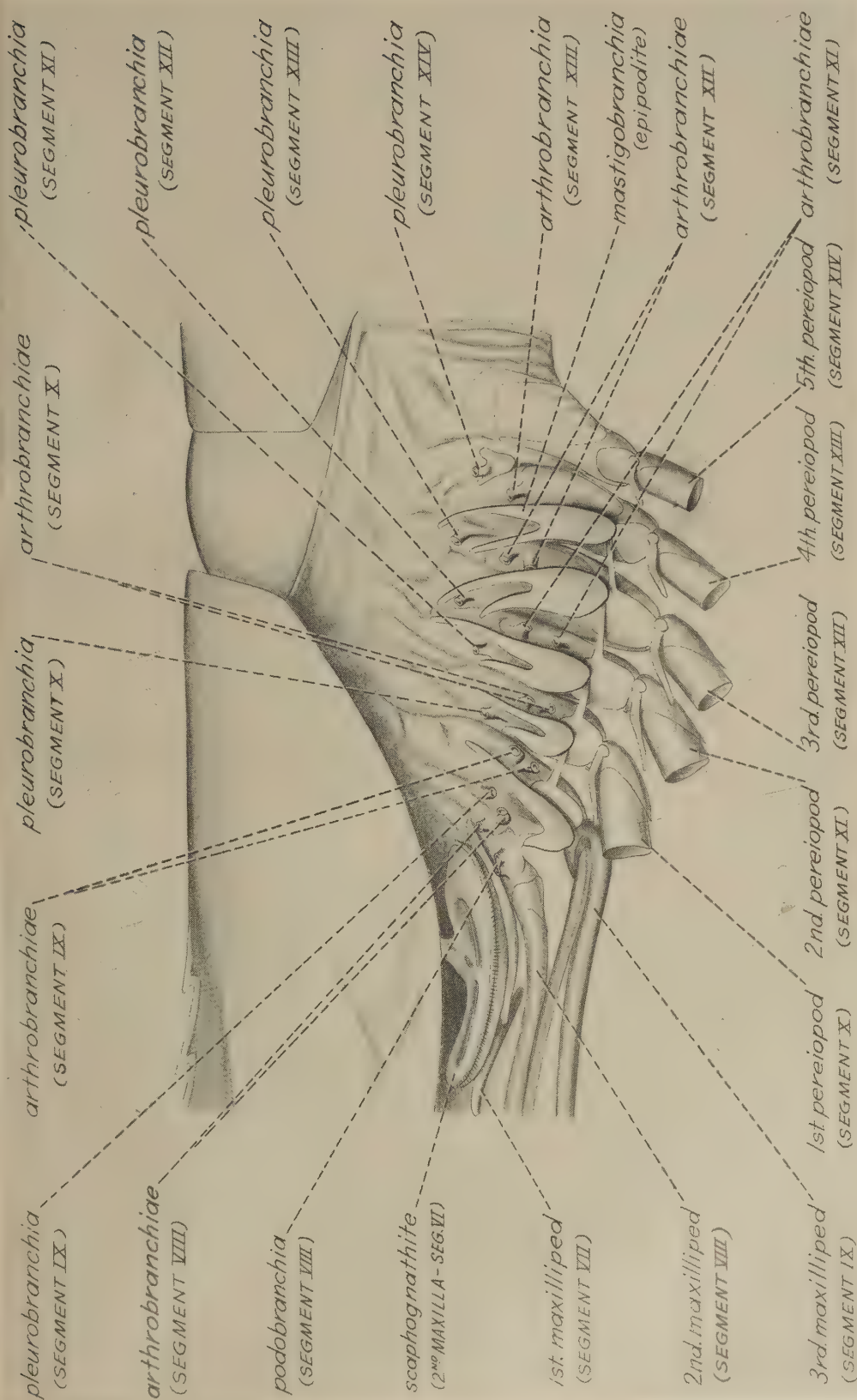


FIGURE 32.—Lateral view of left side of thorax, carapace, and gills removed to show mastigobranchiae and gill attachments.

species of *Penaeus*, Calman (1909) describes a mastigobranchia on the first maxilliped and 1 arthrobranch and 1 pleurobranch on the second maxilliped. Otherwise the branchial formulae of the two penaeids are the same.

MUSCLE ELEMENTS

Of the numerous muscles attaching throughout the gnathothoracic region, many belong to the mouthpart and thoracic appendages and will be taken up when the latter are discussed. Others are protocephalon muscles and have already been considered. Still others are associated with the alimentary canal and heart and will be dealt with in the sections concerned with these organ systems. The many remaining muscles are either small superficial lateral and ventral muscles of the thorax, or large dorsal and ventral muscles. Some of these are morphologically thoracic muscles, and some are morphologically abdominal muscles. In the functional sense, the foregoing muscles may be classified as abdominal musculature, for all of the large muscles taking origin on extensive areas of the dorsal and ventral thoracic skeleton represent the major muscular attachments of the abdomen to the thorax. Substantial movements, especially in the dorsoventral plane, are possible between the thorax and abdomen and it is these muscles which mediate the movements.

SUPERFICIAL LATERAL THORACIC MUSCLES

FIGURE 33

Stretching between the dorsal reaches of the pleural brachia are at least five superficial lateral thoracic muscles. These muscles are extremely thin and weak. Apparently they give rigidity to the thin cuticle of the area during lateral movements of the shrimp along the anteroposterior axis. Schmidt (1915) and Berkeley (1928) do not describe these muscles in *Astacus* and *Pandalus*. The possibility that the epimeral attractor muscles in *Astacus* and *Pandalus* are the superficial lateral thoracic muscles of *Penaeus* is slight.

VENTRAL MUSCLES

SUPERFICIAL VENTRAL THORACIC MUSCLES

FIGURE 36

The superficial ventral thoracic muscles (fig. 36) are situated slightly laterad of the ventral nerve cord. Their median parts are also ventrad

of the nerve cord and the ventral (subneural) artery. These muscles are thin and fan shaped, broad anteriorly and narrowing to posterior attachments to thin connective tissue fasciae. In *Astacus* the lateral and ventral pleurosternal apodemes fuse above the ventral nerve cord to produce the mesophragm of the endoskeletal system. The superficial ventral thoracic muscles of the crawfish attach to these mesophragms. Berkeley (1928) finds that the ventral thoracic muscles in *Pandalus* attach to the endophragmal paraphragms, the lateral fusion product of the pleural and sternal apodemes. As stated above, *Penaeus* has neither paraphragm or mesophragm; however, one would expect the superficial ventral muscles to attach to the little pleurosternal apodemes near the limb foramina. Contrary to such expectations, the superficial ventral thoracic muscles in the white shrimp attach to small apodemes on the pleurosternal brachia. The result is to place these muscles ventrad of the nerve cord. Their function of drawing the ventral thoracic segments together is probably the same as in *Pandalus* and *Astacus*.

In *Penaeus* 7 pairs of superficial ventral thoracic muscles are evident compared to 6 for *Astacus* and *Pandalus*. Schmidt (1915), however, finds two superficial ventral thoracoabdominal muscles in *Astacus* compared to one in *Penaeus* and *Pandalus*.

ANTERIOR THORACIC MUSCLES

FIGURES 33 TO 36

By far the largest ventral muscles of the thorax are the lateral and median anterior thoracic muscles (figs. 33 to 36). The anterior-most muscle of these is the lateral anterior thoracic muscle 1 (figs. 33 to 36). This muscle originates by a large, lateral oval in the region just posterior and slightly dorsad of the hepatic spine. In dorsal view (fig. 35) the muscle may be seen to run posteroventrally to join the other anterior thoracic muscles on the ventral surface of the thorax. In this area all the anterior thoracic muscles are interconnected to segmentally arranged fasciae.

The other lateral anterior thoracic muscles (figs. 34, 35), Nos. 2, 3, 4, and 5, may best be seen in ventral view (fig. 36). These muscles take origin from the ventrolateral fascia of each thoracic segment and run posteriorly into the abdominal musculature with the other anterior thoracic muscles.

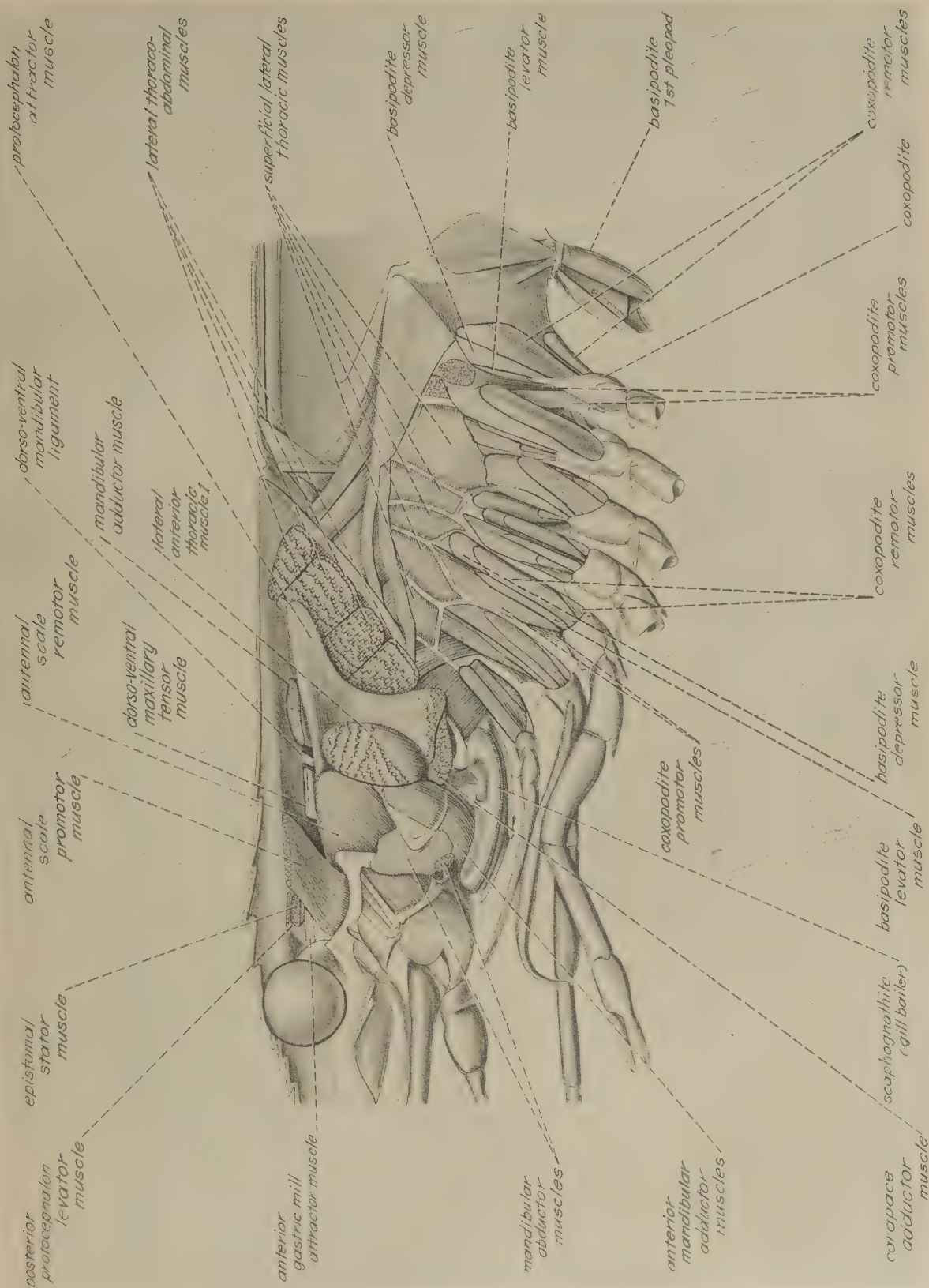


FIGURE 33.—Lateral view of left side of protocephalon and gnathothorax showing muscle origins. Carapace and left body wall removed.

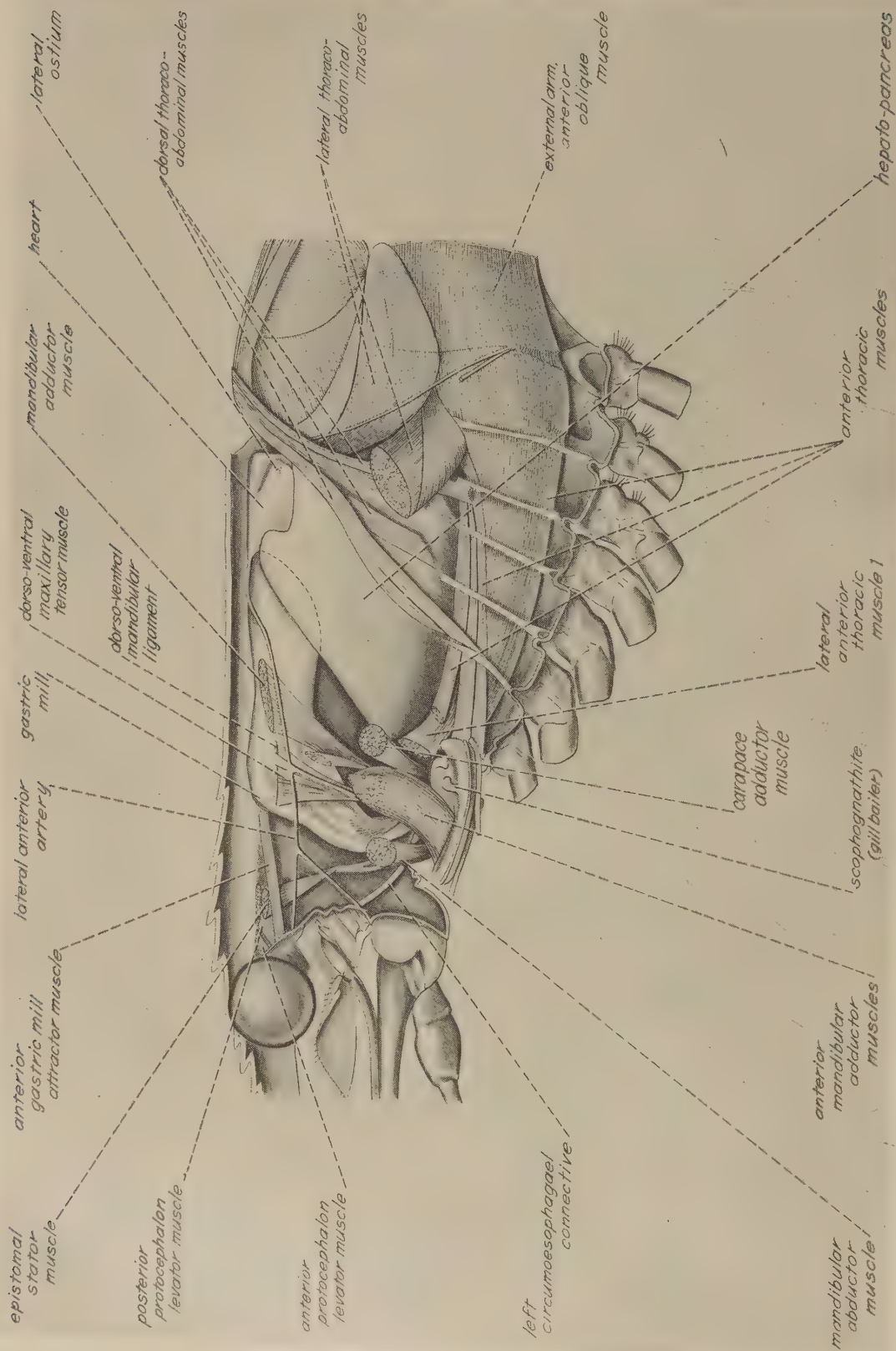


FIGURE 34.—Lateral view of left side of gnathothorax. Lateral muscles removed to show medial muscles and internal organs.

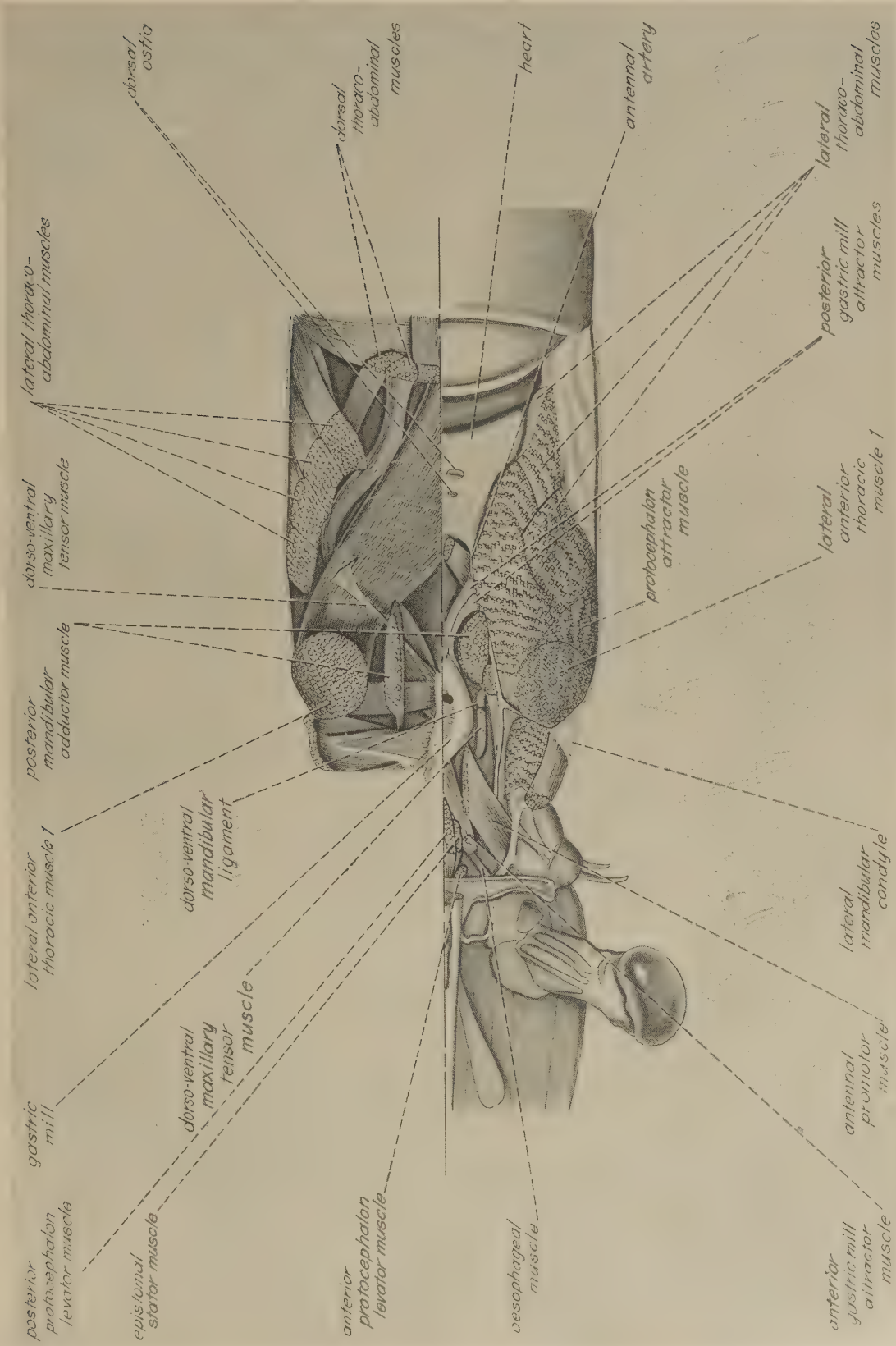


FIGURE 35.—Dorsal views of protocephalon and gnathothorax. Lower section, dorsal muscles. Upper section ventral muscles.

The median anterior thoracic muscles (fig. 36) also originate in segmentally arranged fasciae on the ventral surface. Joined by the other anterior thoracic muscles, the median anterior thoracic muscles pass caudad into the abdominal muscles.

The anterior thoracic muscles play an important part in the powerful flexion of the abdomen on the thorax made by the white shrimps when the animals withdraw suddenly from danger. The anterior thoracic musculature is similar to that in *Pandalus* and *Astacus*, except that in the crawfish these muscles are somewhat smaller. The anterior thoracic muscles are fully homologous in all the forms mentioned here.

VENTRAL HEAD LIGAMENTS

FIGURES 37, 38

The ventral head ligaments are small structures attached between the lateral wings of the epistome and the mandibular endosternite (fig. 37). Apparently they hold the endosternite in position anteriorly. Schmidt (1915) calls these ligaments the ventral head muscles in *Astacus*. Grobben (1917), however, denies the presence of muscle fibers in the structures and suggests the name ligament for muscle. Berkeley (1928) describes the organs as ventral head muscles in *Pandalus*.

CARAPACE ADDUCTOR MUSCLE

FIGURES 33, 34, 37, 38

The carapace adductor muscle originates on the carapace just ventrad of the most ventral part of the protocephalon attractor muscle (fig. 33). The origin point of the carapace adductor is slightly dorsal to the horizontally turned pleural plates, above the gill pump, or scaphognathite. The muscle runs directly mesad to insert on the postero-dorsal midline of the endosternite (fig. 37). The carapace adductor muscle functions as the major position retainer of the endosternite. It may also play a part in necessary distortions of the carapace associated with feeding, molting, and the like.

The carapace adductor muscle appears in many crustacean groups (Grobben 1917). Schmidt (1915) describes it in *Astacus* as the musculus dorsoventralis posterior and Berkeley (1928) has adopted his terminology for the muscle in *Pandalus danae*. Grobben (1917) considers the carapace adductor a useful phyletic character, because of its frequent occurrence, and we are indebted to this worker for the name. Grobben described the

carapace adductor muscle (Schalenschliessermuskel) in species of *Penaeus*, *Palaemon*, *Leander*, *Pandalus*, *Galathea*, and *Athanas*.

DORSAL MUSCLES

DORSAL THORACOABDOMINAL MUSCLES

FIGURES 34, 35

Inserting on the anterodorsal tergum of the first abdominal segment and running forward and down to the laterotergal brachia of the thorax are four pairs of long, slender muscles, the dorsal thoracoabdominal muscles (figs. 34, 35). In dorsal view (fig. 35) the area of attachment may be seen on the dorsal part of the first abdominal segment. The muscles divide around the heart and hepatopancreas as they go to the lateral wall of the thorax. Each muscle originates on its own pleural arm, suggesting that the muscles each belong to specific thoracic segments. Upon contraction the dorsal thoracoabdominal muscles extend the first abdominal segment with respect to the thorax, in opposition to the action of the anterior thoracic muscles.

The dorsal thoracoabdominal muscles are evidently the same muscles as those designated as the dorsal thoracoabdominal muscles in *Pandalus* and *Astacus*, although they appear to be relatively larger in size in the caridean shrimp.

LATERAL THORACOABDOMINAL MUSCLES

FIGURES 34, 35

From an area laterad of the heart and the dorsal thoracoabdominal muscles, the lateral thoracoabdominal muscles (figs. 34, 35) originate along a diagonal line just above the dorsal edge of the inner branchiostegal fold and run ventrally and caudad to junctions with abdominal muscles. Very likely the thoracoabdominal muscles are in reality abdominal muscles. Unlike the anterior thoracic muscles, the lateral thoracoabdominal muscles are not segmented. At least four pairs of these muscles covering a broad area laterally are found in *Penaeus*. Functionally, the lateral thoracoabdominal muscles are involved in the flexing of the abdomen, reinforcing the action of the anterior thoracic muscles. Berkeley (1928) suggests that contractions of the muscles on one side may bend the abdomen laterally in *Pandalus*, and such a movement may also take place in *Penaeus*. The lateral thoracoabdominal muscles of *Penaeus* are

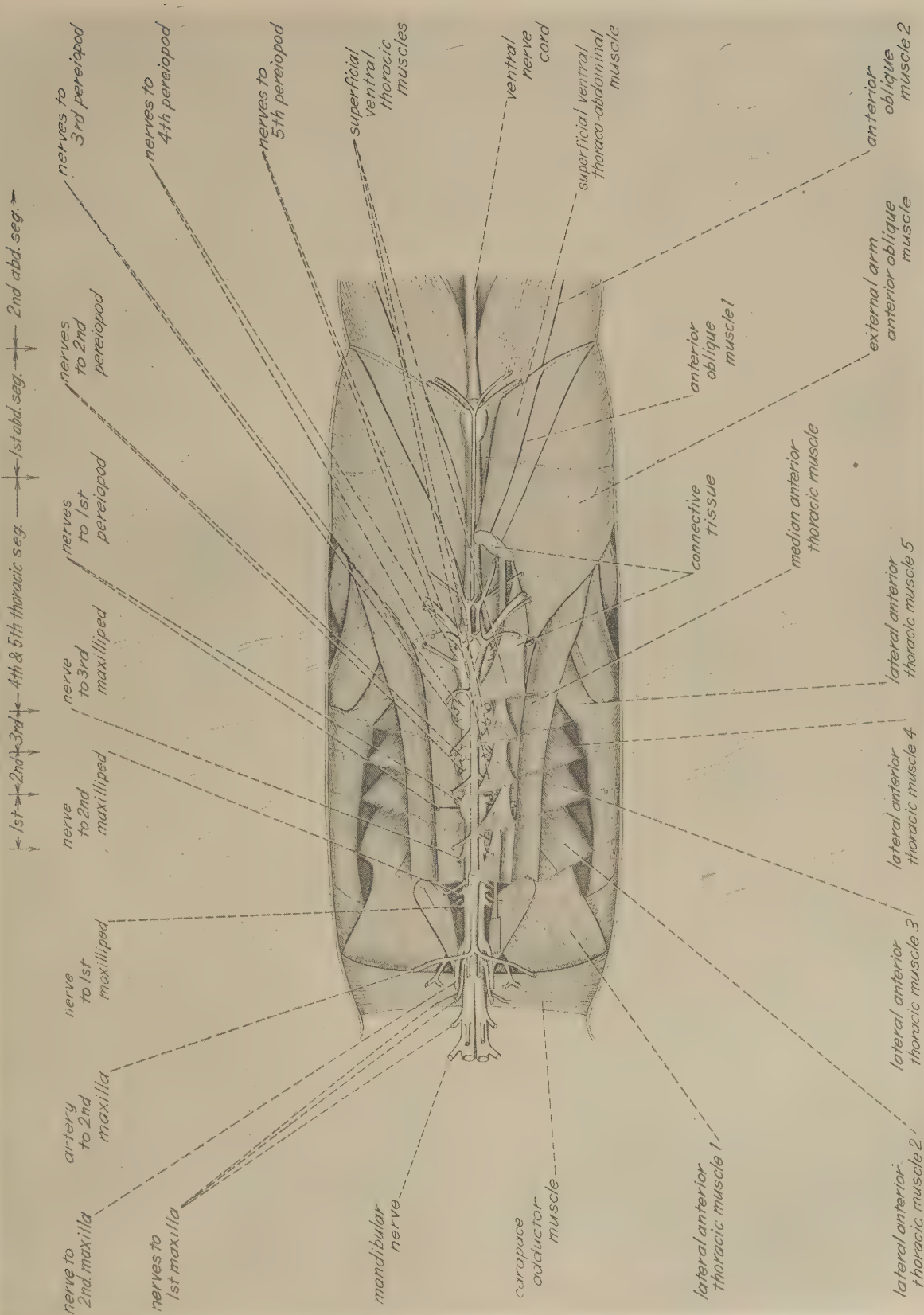


FIGURE 36.—Ventral view of gnathothorax showing muscles, nerves, and blood vessels.

undoubtedly homologous to the muscoli laterales thoracoabdominales of *Astacus* and *Pandanus*.

APPENDAGES OF THE GNATHOTHORAX

The generalized limb of Crustacea is thought to consist of a basal protopodite, made up of a proximal coxopodite and a distal basipodite, bearing two rami, the exopodite and endopodite. This simple arrangement has been retained in the pleopods of Crustacea, notably the Natantia. The typical gnathothoracic limb of *Penaeus*, if one can be said to exist, has the basal coxopodite and basipodite, the latter with a well-developed endopodite and a reduced exopodite. The coxopodite of the white shrimp is attached to the limb base by means of diametrically placed condyles, the axis of the condyles varying in accordance with the location and function of the appendage. The basipodite is attached to the coxopodite by dicondyles whose axis is at right angles to that of the coxal condyles. Distally, the endopodite is divided into the typical five articles: the ischiopodite, meropodite, carpopodite, propodite, and dactylopodite. The coxopodite may give rise to lateral epipodites and in some cases gills.

The exopodite may be modified to long, frond-shaped filaments (second and third maxillipeds) or reduced to a small finger projecting from the basipodites (fig. 31). The gnathal appendages, excluding the second and third maxillipeds, tend to be modified substantially from the typical plan given above.

1. MANDIBLES

SKELETAL ELEMENTS

Of the gnathothoracic appendages, the mandibles are perhaps the most difficult to understand. For one thing the dorsal and internal manifestations of this and other gnathal segments are coalesced or obliterated. In addition, the mandible is complicated by the presence of a true endoskeleton (figs. 37, 38), lying transversely in the gnathothorax above the nerve cord and supporting the gastric mill. Nevertheless, Snodgrass (1951b) adduces strong evidence in support of the evolution of the mandibles from a typical limb on the basis of a comparative study of the skeleton and muscles of the arthropod mandible.

The sclerotized parts of the mandibles in *Penaeus* are relatively simple. The strongly sclerotized incomplete tube of the mandible, with

its ventromedial incisor and molar surfaces and anterior palp, may be seen in ventral view (fig. 29, mandible). The incisor surfaces consist of 2 or 3 sharp ridges used in cutting and tearing food, whereas the flat molar surfaces are for grinding. Both kinds of surface are heavily impregnated with a hardening substance, possibly calcium carbonate, in the form of crystalline stones molded into the cuticle. The stones may be removed by dissection. The tubular part of the mandible, the body, is said to be the coxopodite, whereas the palp is thought to be part of the endopodite (Calman 1909). The subcylindrical body of the mandible is distinctly divided into a large basal portion and a distal lobe, immovably connected to the basal part. In the mandible of *Anaspides*, the basal part is called by Snodgrass (1952) the coxopodite and the distal lobe an endite of the coxopodite. It is interesting to note that the distal gnathal lobe is movable in some arthropods, notably the Diplopoda, and similar to the maxillary lacinia of insects.

The mandibular palp, a part of the endopodite, extends anteriorly from the body of the mandible in the form of a flat, setose lobe shaped to fit around the labrum laterally and anteriorly. The larger part of the broad palp arises from a basal segment, the latter being narrow at its posterior junction with the mandible and broad anteriorly.

The mandible of *Penaeus* is attached to the ventral skeleton by a medial condyle located at a point on the posterior arm of the epistome just laterad of the labrum (fig. 28), and by thin cuticle between the mandible and the lateral extension of the posterior epistomal arm. The mandible has also a lateral condyle on the side of the carapace. Among taxonomists the external manifestation of this lateral mandibular condyle is known incorrectly as the hepatic spine. Snodgrass (1951) erroneously shows two mandibular condyles in *Penaeus* on the laterally spread, posterior bars of the epistome, and does not mention the lateral mandibular condyle. This is curious since, as Snodgrass (1935, 1951b) has shown in earlier work, a significant phyletic series is evident from the study of the arthropod mandible, based upon the evolutionary response of the musculature to restriction of mandibular movements. In the course of the work, Snodgrass makes the fact amply clear that in no case is the site of the primary (lateral) condyle ever to be found on the epistome. Rather, the generalized arthropod man-

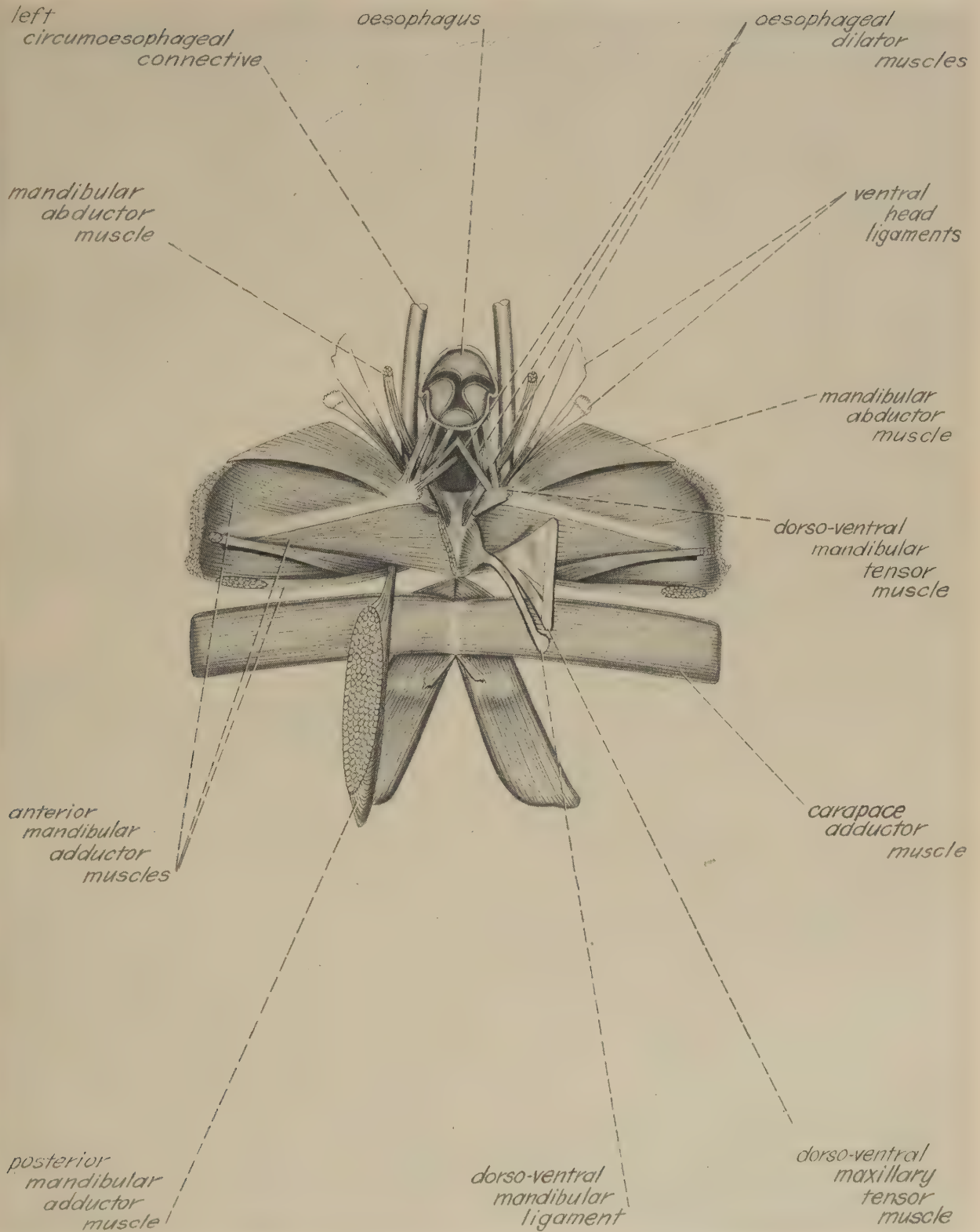


FIGURE 37.—Dorsal view of mandibular musculature.

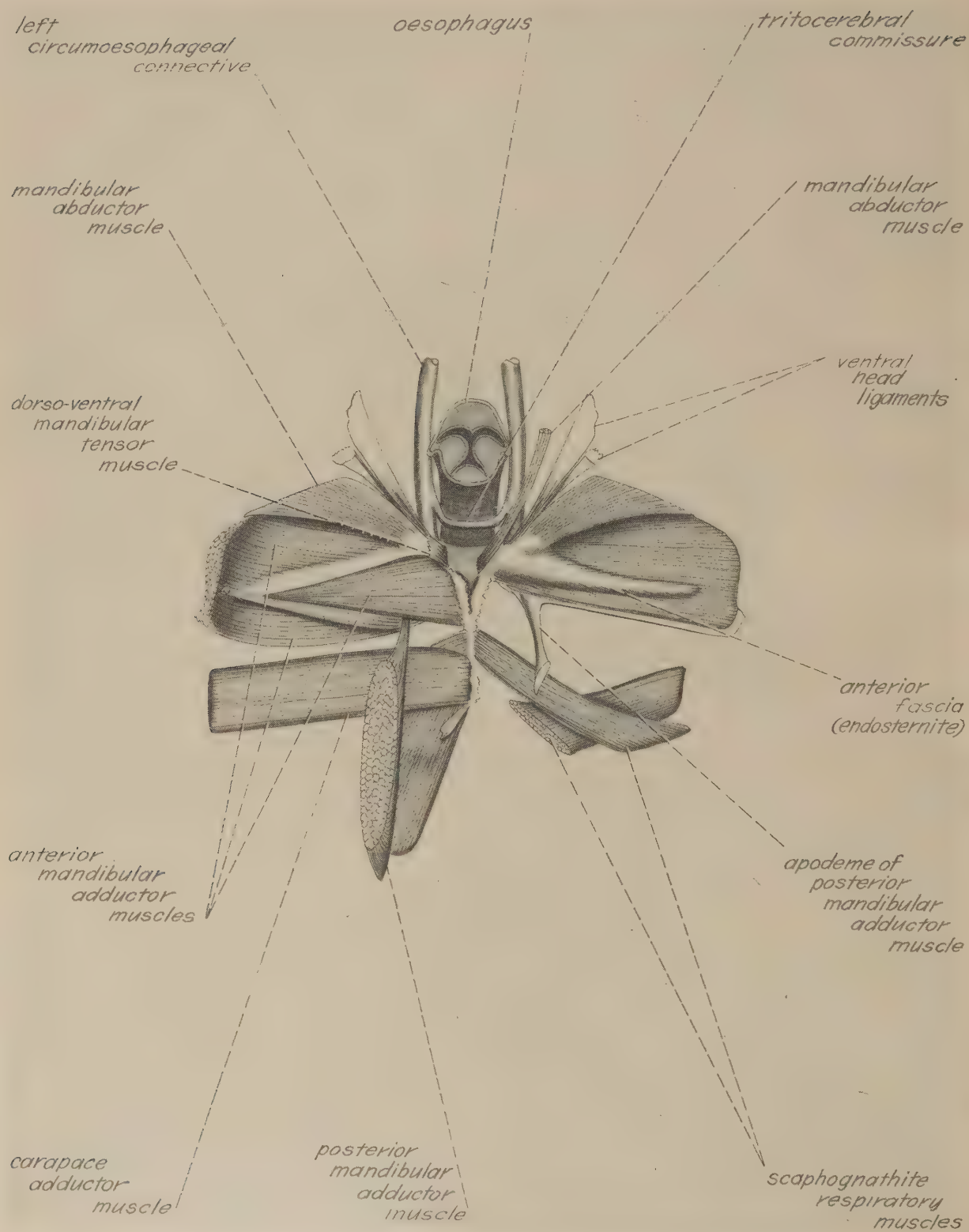


FIGURE 38.—Dorsal view of mandibular musculature, dorsal-most muscles removed.

ible articulates with the mandibular segment at a single point laterally, an arrangement which permits the varied movements of the organ seen at this stage of its evolution.

The monocondylic mandible is found in all the mandibulate arthropods except some higher Crustacea and the pterygote insects (Snodgrass 1935). In the latter forms, a secondary condyle on the epistome is added mesad of the primary lateral condyle. Mandibular movements now become restricted to those about the dicondylar axis. With simplicity, furthermore, comes strength. Snodgrass (1935, 1951b) has described the evolutionary simplification of mandibular musculature which follows in groups developing the dicondylar mandible.

A comparison of the mandibular hinges and musculature of the white shrimp with the account of Snodgrass indicates that *Penaeus* represents a transitional form; for, although the shrimp mandible is dicondylar, its musculature is strongly reminiscent of the monocondylic musculature in apterygote insects, myriapods, and lower Crustacea.

The endoskeleton of the mandibles, mentioned above, is a thick tendon situated between the mandibles and upon which the heavy ventral adductor and abductor muscles insert (figs. 37, 38). The substance of the endoskeleton is extremely tough connective tissue, not sclerotized. Such cuticular components of the structure as may exist are not hard. The mandibular endoskeleton of Crustacea Malacostraca has been variously interpreted. In forms having a well-developed endoskeletal system in the thorax, the mandibular element is considered to be a part of that system.

In *Astacus*, Schmidt (1915) refers to the structure as the endoskeleton, specifically, the head apodeme. Snodgrass (1935), referring to a similar structure in the mandibles of a diplopod, uses the term median ligament. Whether this worker attributes the median ligament to the endoskeleton is not clear. Berkeley (1928) designates the same material in *Pandalus* as the anterior fascia. She evidently considers it to be endoskeletal in nature, and the fusion product of the mandibular and first maxillary segments. Grobben (1917) apparently believes the mandibular tendon to be endoskeletal, naming it the median transverse mandibular tendon. Snodgrass (1952) refers to an intergnathal ligament in *Anaspides tasmaniae* Thomson and an endosternum in *Cambarus lon-*

gulus Girard, in speaking of the structure. In the course of a histological study of muscular and skeletal elements in various Crustacea, Debaisieux (1954) emphasizes that the mandibular endoskeleton is an endosternite, fully homologous in his opinion with that of Arachnida.

No further evidence is offered in the present study for the homology of the endosternite of Arachnida with the mandibular endoskeleton of Crustacea. However, from the work of Debaisieux (1954) on the structure in Crustacea, we can be fairly certain that the mandibular endoskeleton of all crustaceans mentioned above are homologous. In response to the plethora of names for the mandibular endoskeleton, the present writer sees no objection to the terms "endosternite" or "endosternum" given by Debaisieux (1954) and Snodgrass (1952). These names are used here as equivalent to one another and to the expression, "mandibular endoskeleton." Whether an endoskeleton and an endosternite are morphologically equal is not made clear in the literature.

As a final word on the composition of the endoskeleton, it will be recalled that Schmidt (1915) describes two small medial muscles between the mesophragsms of the endosternite, the endophragmal compressor muscles. Grobben (1917) denies the existence of muscle fibers in this material in the crawfish and establishes that the area is composed of connective tissue.

MUSCLE ELEMENTS

MANDIBULAR ABDUCTOR MUSCLES

FIGURES 33, 37, 38

At least three mandibular abductor muscles (figs. 33, 37, 38) are found in *Penaeus*. The smallest is the most anterior. This muscle arises in connective tissue in the anterior region of the gnathothorax, lateral to the esophagus and dorsal to the circumesophageal connective, and runs posteriorly and ventrad to insert on the anteromedial part of the mandibular endoskeleton. Contractions of these slender muscles aid in opening the mandibles. No counterpart of the muscle is described for the other crustaceans to which reference has been made.

The large and important mandibular abductors originate in connective tissue on the laterotergal plates (fig. 33) and run posteromedially to insertion areas on the mandibular endoskeleton (figs. 37, 38). Their contractions serve to open the

gnathal elements of the mandibles. These two mandibular abductor muscles are fully homologous with the major and minor mandibular abductor muscles in *Astacus* and with the mandibular abductor muscle in *Pandalus*. Presumably the homology holds for the major and minor mandibular abductors of *Callinectes*.

DORSOVENTRAL MANDIBULAR TENSOR MUSCLE

FIGURE 37

The long tendon of the dorsoventral mandibular tensor muscle (fig. 37) originates in connective tissue on the dorsal carapace by an extremely thin band mesad of the origin of the antennal promotor muscle. The thin apodeme runs ventrad to the muscle body which inserts on the anterior part of the mandibular endoskeleton, slightly laterad of the small mandibular abductor muscle. The muscle pulls the mandibular endoskeleton dorsad, possibly as a minor adjustment of the mandible in feeding. The muscle was named by Grobben (1917), who described it in a number of crustaceans, including species of *Penaeus*, *Palaemon*, *Leander*, *Pandalus*, and *Nebalia*. The description by Berkeley (1928) of *Pandalus dange* does not include the dorsoventral mandibular tensor muscle, although it is almost certainly present in that species. The muscle is missing in *Astacus*.

ANTERIOR MANDIBULAR ADDUCTOR MUSCLES

FIGURES 33, 34, 37, 38

The anterior mandibular adductor muscles (figs. 33, 34, 37, 38) are the largest occupants of the subcylindrical mandibular body. At least three muscles are evident in *Penaeus*, although a study of the nerves may show that the muscle groups are actually parts of the same muscle. The adductors originate laterally throughout the body of the mandible and insert extensively over the tissues of the mandibular endosternite. Contractions of the anterior mandibular adductors draw the gnathal lobes of the mandibles together. The action is direct and efficient. The anterior mandibular adductor muscles of *Penaeus* are the homologs of the musculus adductor anterior mandibulae in *Astacus* and *Pandalus*.

DORSOVENTRAL MANDIBULAR LIGAMENT

FIGURES 33, 34, 35, 37

The dorsoventral mandibular ligament (figs. 33, 34, 35, 37) originates in the thick connective tissue

of the dorsal carapace by a narrow, thin, apodeme between the dorsal edges of the protocephalon attractor muscle and the gastric mill (fig. 35). The apodeme runs anteroventrally to a thin, fan-shaped muscle inserting on the dorsal surface of the endosternite. It crosses over the tendon of the dorsoventral maxillary tensor muscle medially. The muscle is closely appressed to the fibers of the dorsoventral maxillary tensor, but may be separated readily from the latter in the sagittal plane. The dorsoventral mandibular ligament (or muscle) aids in retaining the endosternite in position.

The structure was found in a species of *Penaeus* and of *Palaemon* and named by Grobben (1917). Since the dorsoventral mandibular ligament actually ends in a small muscle, Grobben's name should be replaced with the name, "dorsoventral mandibular muscle." A muscle in *Pandalus*, designated by Berkeley (1928) as the musculus dorsoventralis anterior 2, is probably the dorsoventral mandibular ligament.

POSTERIOR MANDIBULAR ADDUCTOR MUSCLE

FIGURES 30, 33, 34, 35, 37, 38

Taking origin in an elongate ovoid on the dorsal carapace is a large, wedge-shaped muscle, the posterior mandibular adductor muscle (figs. 35, 37). Growing narrower as it passes ventrad, the posterior adductor attaches to a broad apodeme (figs. 29, B; 30, apodeme, mandibular adductor muscle). The adductor apodeme arises from the posterior margin of the semitubular mandibular body. The apodeme is so placed that a dorsal pull of the powerful muscle brings the gnathal lobe to the midline. The posterior mandibular adductor muscles are widely represented in the Arthropoda.

MANDIBULAR PALP FLEXOR MUSCLE

FIGURE 29

The muscles operating the mandibular palp are situated either in the gnathal lobe of the mandible or in the basal segment of the palp. The distal lobe of the palp contains no muscles. The mandibular palp flexor muscle (fig. 29) originates in the proximal region of the basal palp segment near the foramen between the gnathal lobe and the palp base. The muscle runs distally, becoming broad and flat, and inserts on the posteroventrad margin of the distal palp lobe. In action, the mandibular palp flexor turns the distal lobe ventrad. An ap-

parently similar muscle in the palp of *Astacus* is called by Schmidt (1915) a palp flexor also. Berkeley (1928) describes both a palp flexor and a palp extensor in the body of the mandible of *Pandalus*.

MANDIBULAR PALP ADDUCTOR MUSCLES

FIGURE 29

The proximal, and larger, mandibular palp adductor muscle originates on the posterior wall of the gnathal lobe of the mandible and inserts on the medial margin of the basal palp foramen. The smaller palp adductor originates dorsad of the insertion of the larger palp adductor on the medial margin of the gnathal lobe foramen. Contractions of these muscles turn the palp segments toward the midline. The proximal mandibular palp adductor muscle is probably homologous with the *musculus flexor a mandibulae* of *Astacus* and either the palp flexor or extensor of *Pandalus*.

2. PARAGNATHA

The paragnatha (figs. 29, 39) are two rounded lobes suspended from small foramina in the ventral skeleton immediately posterior to the gnathal lobes of the mandibles. Their cuticle is very thin, except for a slightly thickened ridge along the posterior surface of each paragnath. No intrinsic muscles are found in the body of the paragnath in *Penaeus*; however, a small muscle, the paragnathal muscle (fig. 39), inserts on the lateral margin of the paragnath. The paragnathal muscle moves the paragnath laterally and anteriorly against the gnathal lobe of the mandible.

Classically, the paragnatha have been interpreted as a secondary development of the mandibular segment, on grounds of their embryonic development and because of their innervation by mandibular nerves. Also, the view that they belong to the maxillae has been expressed. The idea that the paragnatha are reduced true appendages has in general been discounted, despite the presence of movable terminal lobes in the paragnatha of Tanaidacea (Crustacea). Snodgrass (1935) mentions the similarity of the insectan superlinguae to the crustacean paragnatha. This worker feels, apparently, that there is no evidence that the paragnatha are appendicular or that the superlinguae and paragnatha are homologous structures. He does indicate (Snodgrass 1952) in a study of *Cambarus*, that the paragnatha are

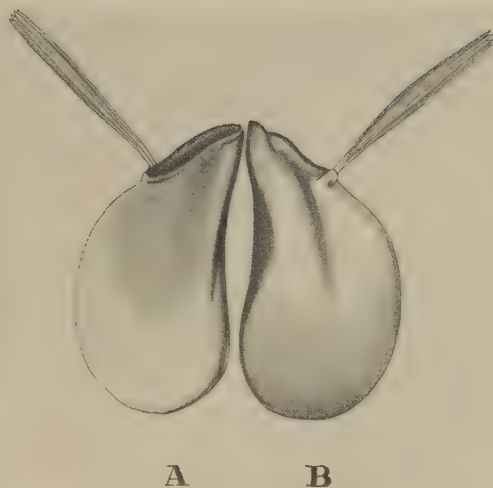


FIGURE 39.—Paragnatha. A. Anterior view. B. Posterior view.

mandibular entities. In a comparative study of arthropod nerves, Henry (1948a) assigns the paragnatha to the mandibular segment by virtue of their innervations. She invariably places the paragnathal nerves in a position posterior to those of the mandibles.

Recently, Chaudonneret (1955, 1956), in a detailed study of the gnathal nerves of *Orconectes limosus* (Rafinesque) (= *Cambarus affinis* Say), takes issue with Henry (1948a) and advances the idea that the paragnathal nerves are in fact anterior to those of the mandibles and entirely distinct from the mandibular nerves. On other grounds, furthermore, this worker suggests that the paragnathal foramina are anterior to the mandibular foramina and holds that their relative positions with respect to the mouth are constant in the Malacostraca. In the opinion of Chaudonneret (1956), the facts make difficult the interpretation of the paragnatha as either epithelial lobes or parts of the mandibles. Instead, this investigator thinks that the paragnatha may belong to a reduced premandibular, paragnathal segment homologous to the insectan superlingual segment.

If the paragnatha are indeed premandibular and homologous to the superlinguae, an interpretation which Snodgrass (1935) seems to consider possible, certain aspects of the morphology of the insect hypopharynx will need review. Careful study of the paragnathal nerves in *Penaeus* tends to support the view of Chaudonneret (1956) that this nerve is slightly anterior to the mandibular nerve. However, the gross anatomy will have to

be verified by histological work before a decision on this interesting point can be made.

3. MAXILLAE

3a. First Maxillae

SKELETAL ELEMENTS

The first maxillae articulate with the ventral gnathal skeleton at relatively large foramina slightly caudad and laterad of the paragnathal foramina (fig. 28, A). The medial lobes of these accessory feeding organs fit closely to the posterior surfaces of the paragnatha (fig. 30), and thus project anteroventrally over the mouth from the gnathal framework.

Although the first maxillae is true appendages, they are much modified for functional ends. The body of the first maxilla is produced into several lobes and a flagellum (figs. 29, 40). The flat medial lobes are the proximal coxopodite and the distal basipodite. The medial edges of these lobes are covered with stiff hairs or spines, those on the basipodite margin being especially strong. The spines function to hold food particles. Laterally, the coxopodite is produced into a rounded lobe from which a tuft of large, plumose, sensory setae project. Endites of the basipodite, including an anterior three-jointed flagellum, extend anterolaterally from the basipodite. Schmidt (1915) considers the endites the endopodite. Various lobes of the basipodite endites bear sensorial hairs. A single large seta projects anteriorly from the base of the flagellum.

MUSCLE ELEMENTS

The musculature of the first maxilla in *Penaeus* appears substantially similar to that of other Decapoda. Groups of muscles function to bring the spinous gnathal margins of the appendage to the midline in feeding. Other muscles open the opposing gnathal parts and make various position adjustments. *Penaeus* appears to have at least 10 muscles and muscle groups in the first maxilla, against 9 each for *Pandalus*, *Astacus*, and *Callinectes*.

COXOPODITE PROMOTOR MUSCLE OF FIRST MAXILLA

FIGURE 40

The first maxilla coxopodite promotor muscle (fig. 40) originates on a large sternal apodeme

arising from the brachia between the first and second maxillary foramina. This apodeme penetrates the substance of the endosternite. The promotor muscle runs ventrolaterally to insert in the lateral lobe of the coxopodite. Upon contraction, the muscle turns the first maxilla forward and upward. The first maxilla coxopodite promotor muscle in *Penaeus* is homologous with the musculus promotor I maxillae of *Pandalus*, *Astacus*, and *Callinectes*. Berkeley, Schmidt, and Cochran state that the promotor muscle originates on the head apodeme, or endosternite, in the above three crustaceans, whereas in *Penaeus* the area of origin of the promotor and other lateral muscles is not directly on the endosternite.

COXOPODITE REMOTOR MUSCLE OF FIRST MAXILLA

FIGURE 40

Taking origin on the sternal apodeme some distance ventrad of the origin of the coxopodite promotor muscle, the first maxilla coxopodite remotor muscle (fig. 40) passes laterad to the lateral lobe of the coxopodite. Contractions of the muscle draw the first maxilla posteroventrad. The coxopodite remotor in *Penaeus* is the homolog of the musculus remotor *a* or *b* I maxillae of *Pandalus*, *Astacus*, and *Callinectes*. In the latter three forms, two remotor muscles are described.

LATERAL COXOPODITE ADDUCTOR MUSCLE OF FIRST MAXILLA

FIGURE 40

The first maxilla lateral coxopodite adductor muscle (fig. 40) is a long, slender muscle originating on the laterotergal wall and running anteroventrally to a point of insertion on the medial region of the coxopodite. Contractions of the muscle raise the first maxilla and turn the gnathal lobes towards the midline. Berkeley and Schmidt maintain that the lateral coxopodite adductors in *Pandalus* and *Astacus* originate on the lateral carapace. Despite this difference, the lateral coxopodite adductors in the latter forms are homologous with those of *Penaeus*. Cochran (1935) finds in *Callinectes* a muscle termed by her the posterior adductor muscle. This muscle is a possible homolog of the lateral coxopodite adductors in *Penaeus*.

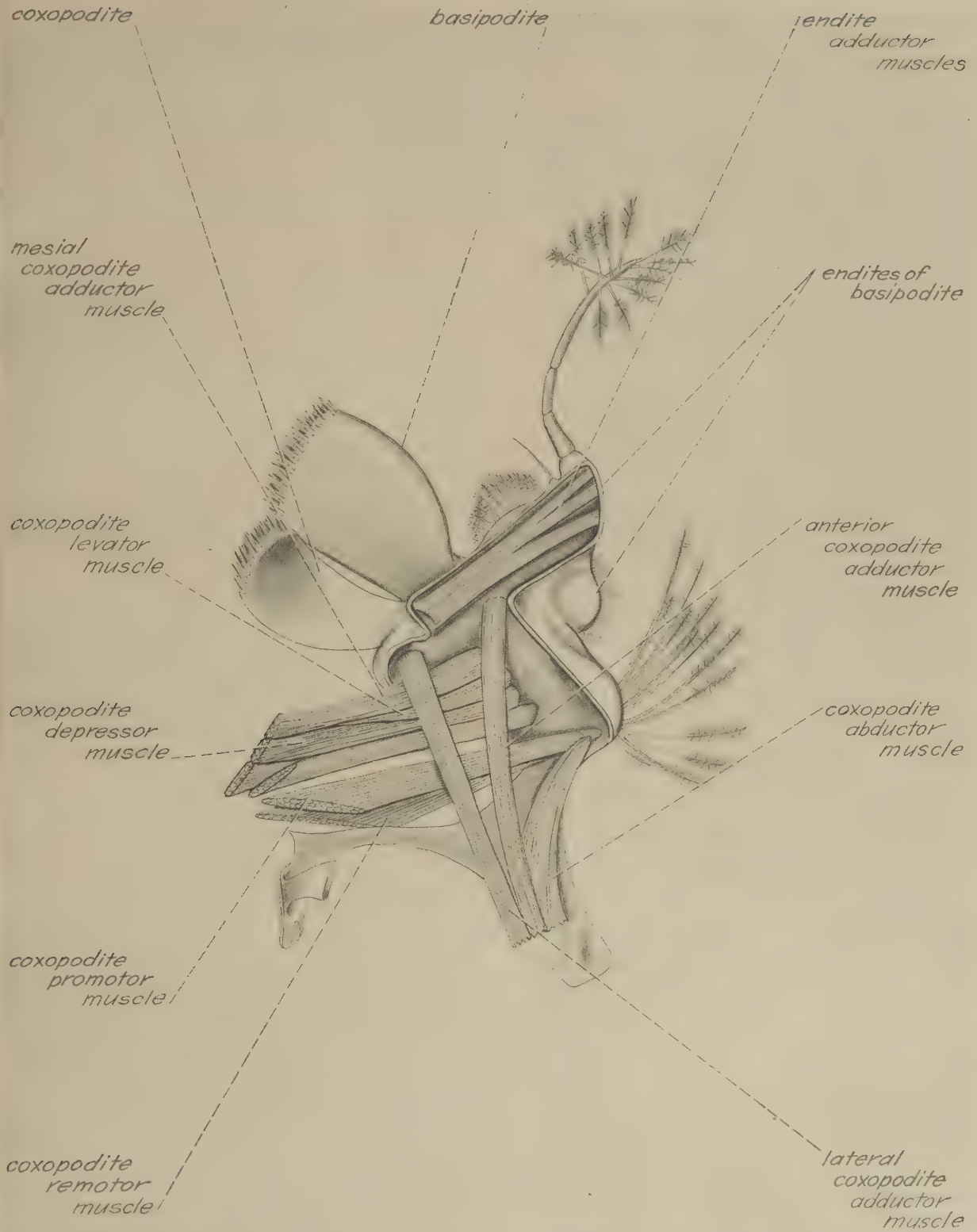


FIGURE 40.—Dorsal view of right first maxilla. Dorsal cuticle removed to show muscles.

MESIAL COXOPODITE ADDUCTOR MUSCLE OF FIRST MAXILLA

FIGURE 40

Arising from the large sternal apodeme of the coxopodite promotor muscle, the first maxilla mesial (=medial) coxopodite adductor muscle (fig. 40) inserts in the coxopodite. The muscle functions to turn the first maxilla to the midline, thus bringing the opposing gnathal lobes together. The first maxilla mesial coxopodite adductor muscle in *Penaeus* is the same muscle as the musculus adductor medialis coxopoditis I maxillae in *Astacus* and *Pandalus*. *Callinectes* does not appear to have this muscle.

ANTERIOR COXOPODITE ADDUCTOR MUSCLE OF FIRST MAXILLA

FIGURE 40

The first maxilla anterior coxopodite adductor muscle (fig. 40) originates on the laterotergal plate adjacent to the origin of the lateral coxopodite adductor muscle. The muscle passes antero-ventrally, diverging from the lateral adductor, and inserts somewhat anteriorly of the insertion of the lateral adductor. Together with the medial and lateral coxopodite muscles, the anterior adductor closes the gnathal lobes of the first maxilla. The anterior adductor is not evident in *Astacus* or *Pandalus*, but does appear in *Callinectes*.

COXOPODITE ABDUCTOR MUSCLE OF FIRST MAXILLA

FIGURE 40

The first maxilla coxopodite abductor muscle (fig. 40) originates with the anterior and lateral adductor muscles on the laterotergal wall. The muscle runs anteriorly and inserts in the lateral lobe of the coxopodite. Upon contraction, the muscle pulls the first maxilla away from the midline, opening the gnathal lobes. The coxopodite abductor muscle is found in *Pandalus*, *Astacus*, and *Callinectes*.

COXOPODITE LEVATOR MUSCLE OF FIRST MAXILLA

FIGURE 40

The first maxilla coxopodite levator muscle (fig. 40) is attached to the sternal apodeme upon which the medial adductor and the promotor and

remotor muscles originate. It inserts on the coxopodite. The angle of the muscle attachment is such that its contractions raise the first maxilla. The same muscle as the coxopodite levator in *Penaeus* appears in *Astacus*, *Pandalus*, and *Callinectes*.

COXOPODITE DEPRESSOR MUSCLE OF FIRST MAXILLA

FIGURE 40

The first maxilla coxopodite depressor muscle (fig. 40) originates on the base of the sternal apodeme mentioned above. The muscle runs to its insertion on the coxopodite so that its contractions draw the first maxilla ventrad, thereby lowering the gnathal lobes away from the mandibles and paragnatha. The coxopodite depressor muscle exists in the other crustaceans referred to above.

ENDITE ADDUCTOR MUSCLE OF FIRST MAXILLA

FIGURE 40

Intrinsic to the endite of the basipodite is a group of muscles, the first maxilla endite adductor muscle (fig. 40), which pass across the proximal neck of the gnathal lobe of the basipodite to the base of the endite flagellum. The muscle bends the flagellum mesad. The endite adductor muscle is common to the first maxilla of *Penaeus*, *Pandalus*, *Astacus*, and *Callinectes*.

DORSOVENTRAL MAXILLARY TENSOR MUSCLE

FIGURES 33, 34, 35, 37

This muscle originates by a broad, fan-shaped apodeme in the connective tissue of the dorsal carapace just laterad of the gastric mill (fig. 35). The apodeme runs ventromedially to the small dorsoventral maxillary tensor muscle (fig. 37) lying laterad of the muscle of the dorsoventral mandibular ligament and closely applied to it. Schmidt (1915) described the muscle as the anterior dorsoventral muscle in *Astacus* and Berkeley (1928) has followed this terminology in her work on *Pandalus danae*. Grobben (1917) studied the muscle in species of *Penaeus*, *Palaemon*, *Leander*, and *Pandalus*, and concluded that, on grounds of its innervation by nerves of the first maxilla, the muscle should be renamed the "dorsoventral maxillary tensor muscle." The name given by Grobben is adopted here.

3b. The Second Maxillae

The second maxilla is one of the most extensively modified appendages of higher Crustacea. The structure serves a double function. The medial lobes participate in feeding while the large lateral part pumps water over the gills. Despite its complexity, the second maxilla is so remarkably uniform in structure and musculature throughout a broad phyletic spectrum of higher Crustacea that, as Calman (1909) has shown, the appendage is of limited value in the study of crustacean evolution.

SKELETAL ELEMENTS

The second maxilla projects laterally and ventrally from its attachment point on the ventral skeleton. The large foramen enters a deeply sculptured coxopodite from which two small, medial gnathal lobes arise (figs. 29, *D*; 41). Distal to the coxopodite lies the complex basipodite bearing two larger, medial gnathal lobes and an anterior endite (figs. 29, *D*; 41). Spines on the gnathal lobes aid in holding food. Lateral to the basipodite lies the flat, indented scaphognathite, or gill pump (fig. 41). The folds and grooves of the scaphognathite represent areas of muscle attachment and of articulation.

Many different interpretations of the components of the second maxilla are encountered in the literature. In *Astacus*, Schmidt describes the gnathal lobes as partly coxal and partly basal, as had been done in the present study of *Penaeus*. However, the former considers the basipodite endite the endopodite, and is followed by Berkeley in *Pandalus*. Calman (1909) refers to the endite as a palp. The position of Cochran (1935) in her study of *Callinectes* is rather inconsistent. When describing the second maxilla of the blue crab, this worker calls all of the gnathal lobes endites of the coxopodite, and terms the basipodite endite the endopodite. By contrast, in a description of the mouthparts of a number of crustaceans included as a subsection of the study of the blue crab musculature, Cochran labels the median gnathal lobes as basal and the anterior lobe as an endite of the basipodite, as has been done by the present writer in *Penaeus*, even though the second maxillae of *Callinectes* and *Penaeus* are very similar. To say the least, the

composition of the second maxilla needs clarification.

MUSCLE ELEMENTS

The principle function of the muscles of the second maxilla is the operation of the scaphognathite as a gill pump. As mentioned above, the gill pump lies in a narrow channel through which the water is moved that passes over the gills. If the body processes of a shrimp are reduced by chilling, and the branchiostegite is cut away, the gill pump may be observed in slow action. Two functionally interrelated but distinct oscillations of the scaphognathite occur. The more obvious is that taking place about the horizontal, lateral axis through the gill pump, and by which the scaphognathite is tipped back and forth, or rotated on its axis. The less obvious oscillation is the dorso-ventral movement of the lateral margin of the scaphognathite about the long axis of the structure. The marginal undulation tends to ramify the former oscillation with respect to water pumping.

Water is drawn into the pumping chamber by a forward tipping of the anterior end of the gill pump to the floor of the chamber. At the same time the posterolateral margins of the pump are raised to the top of the chamber. The posterolateral margins of the pump are now brought ventrad at which time the anterior end rises, and the whole organ rolls anteriorly along the floor of the chamber, forcing water out of the cephalic aperture of the pumping chamber.

Conflicting opinions about the skeletal nomenclature of the second maxilla have given rise to some confusion in the naming of the muscles. In addition, small but important differences in the number, arrangement, and in particular the functions of the second maxillary muscles of *Penaeus* are apparent when the second maxilla of the white shrimp is compared to that of *Astacus*, *Pandalus*, and *Callinectes*. The differences are of sufficient magnitude to make difficult the homology of all the muscles in *Penaeus* with those in the three crustaceans mentioned without knowing the details of comparative innervations. Under pain of causing further confusion of names in the literature, the present writer renames the muscles of the second maxillae of *Penaeus* in accordance with their functions.

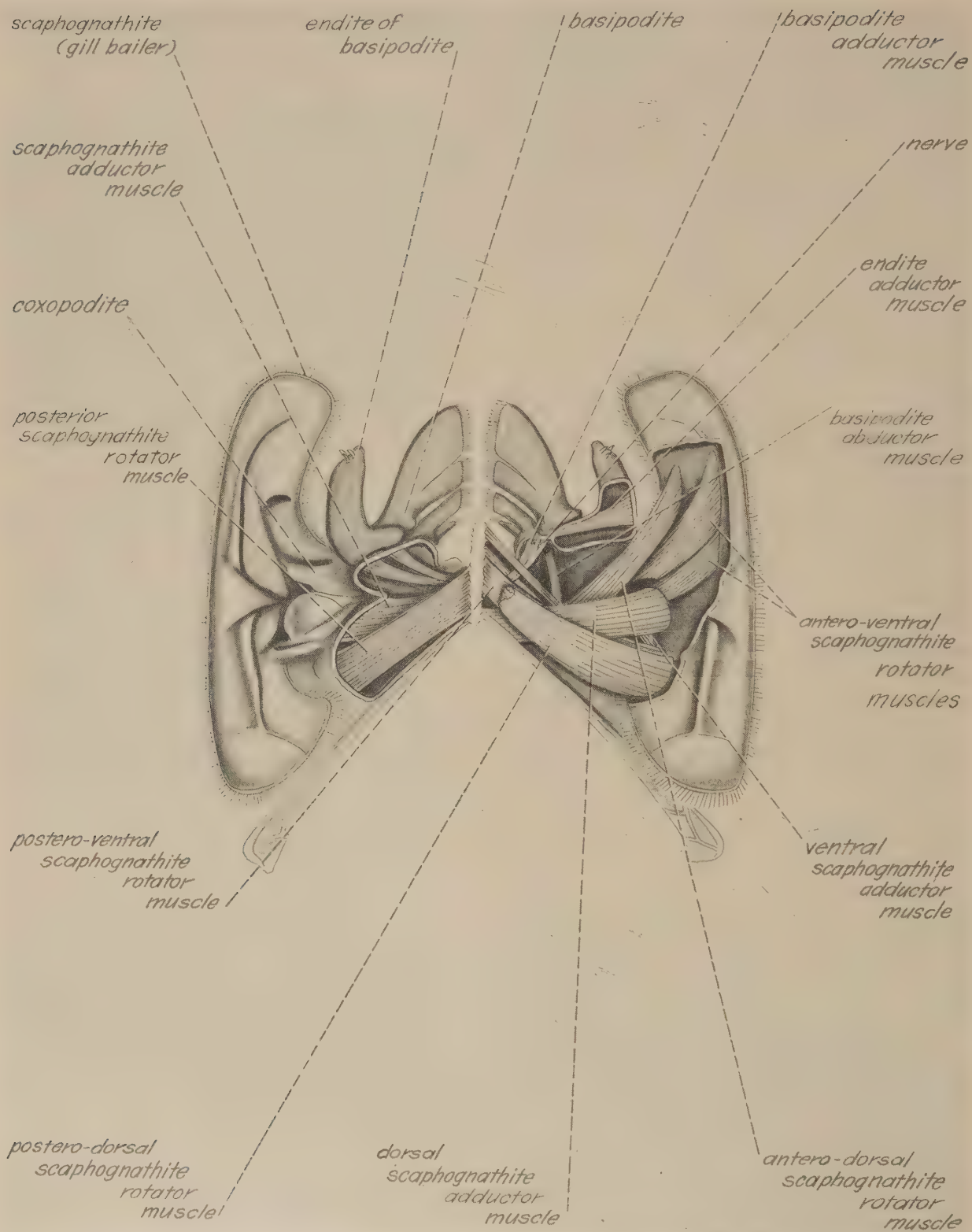


FIGURE 41.—Dorsal view of second maxilla. Left side, intact appendage. Right side, dorsal cuticle removed to show muscles.

BASIPODITE ADDUCTOR MUSCLE OF SECOND MAXILLA

FIGURE 41

This small muscle originates on the medial margin of the coxopodite and runs to the proximal region of the gnathal lobe of the basipodite. The second maxilla basipodite adductor muscle turns the basipodite lobes toward the midline. The only muscle in *Astacus* in a similar position is part of the musculus depressor II maxillae.

BASIPODITE ABDUCTOR MUSCLE OF SECOND MAXILLA

FIGURE 41

The basipodite abductor muscle runs from the posterior rim of the coxopodite to the proximal region of the basipodite gnathal lobes, laterad of the insertion of the basipodite adductor. Contractions of the basipodite abductor open the gnathal lobes. Like the basipodite adductor, the basipodite abductor may be the same muscle as part of Schmidt's musculus depressor II maxillae in *Astacus*.

ENDITE ADDUCTOR MUSCLE OF SECOND MAXILLA

FIGURE 41

The endite adductor muscles (fig. 41) arises in the proximal region of the basipodite gnathal lobes and passes laterad to insert on the lateral margin of the basipodite endite. Its contractions turn the endite mesad. The endite adductor of *Penaeus* is fully homologous with the endopodite adductor muscle of *Astacus*, *Pandalus*, and *Callinectes*.

POSTEROVENTRAL SCAPHOGNATHITE ROTATOR MUSCLE OF SECOND MAXILLA

FIGURE 41

The posteroventral scaphognathite rotator muscle, in company with several of the so-called respiratory muscles, originates on a large, sternal apodeme arising from the sternal brachia between the second maxilla and the first maxilliped. The dorsal, or distal, portion of this apodeme lies close to the dorsal end of the large sternal apodeme upon which muscles of the first maxilla are attached, but the second maxillary apodeme is not deeply

embedded in the substance of the endosternite as is the first maxillary apodeme. The posteroventral scaphognathite rotator runs posterolaterally to a point of insertion on the ventral surface of the scaphognathite. This muscle brings the posterior tip of the gill pump ventrad and removes the whole structure. The muscle is evidently the homolog of the musculus respiratorius *e* II maxillae in *Astacus* and the other forms referred to.

DORSAL SCAPHOGNATHITE ADDUCTOR MUSCLE OF SECOND MAXILLA

FIGURE 41

This muscle originates on the apodeme of the second maxilla and runs ventrolaterally to insert on the dorsal surface of the scaphognathite. The dorsal scaphognathite adductor muscle turns the posterolateral margin of the gill pump dorsad. Although far from clear, the muscle may be the same muscle as Schmidt's musculus respiratorius *b* II maxillae in *Astacus*.

POSTERODORSAL SCAPHOGNATHITE ROTATOR MUSCLE OF SECOND MAXILLA

FIGURE 41

The posterodorsal scaphognathite rotator muscle originates on the dorsal part of the sternal apodeme of the second maxilla and passes laterally to the dorsal surface of the scaphognathite. In action, the muscle aids the posteroventral rotator muscle in removing the whole scaphognathite, but opposes the posteroventral rotator by lifting the posterior tip of the gill pump. The posterodorsal rotator in *Penaeus* is almost certainly the homolog of the musculus respiratorius *d* II maxillae of *Pandalus*, *Astacus*, and *Callinectes*.

VENTRAL SCAPHOGNATHITE ADDUCTOR MUSCLE OF SECOND MAXILLA

FIGURE 41

The ventral scaphognathite adductor muscle originates in the ventral region of the coxopodite and inserts on the scaphognathite ventrad of the insertion of the posteroventral rotator muscle. The muscle turns the lateral margin of the gill pump ventrad, in opposition to the action of the dorsal adductor.

ANTERODORSAL SCAPHOGNATHITE ROTATOR MUSCLES OF SECOND MAXILLA

FIGURE 41

The anteroventral scaphognathite rotator muscles have separate origins and may well be two different entities. The posteriormost of the two is a long muscle extending from the ventral apodeme to an area well out in the anterior region of the scaphognathite. The more anterior rotator originates in the coxopodite and passes to a broad area of insertion in the anterolateral region of the gill pump. This muscle may be the counterpart in *Penaeus* of the scaphognathite flexor muscle of *Astacus*, *Pandalus*, and *Callinectes*. The posteriormost rotator of this pair in *Penaeus* apparently is not homologous with any of the muscles in the other crustaceans referred to. These muscles bring the anterior tip of the scaphognathite dorsad and promote the whole structure.

ANTERODORSAL SCAPHOGNATHITE ROTATOR MUSCLE OF SECOND MAXILLA

FIGURE 41

The anterodorsal scaphognathite rotator muscle takes origin on the endosternal apodeme of the second maxilla and runs anteroventrally to an area of insertion mesad of the anteroventral rotator muscles. The muscle reinforces the action of the anteroventral rotators in raising the anterior tip of the scaphognathite and promoting the structure. As nearly as can be determined, the anterodorsal rotator in *Penaeus* is the musculus respiratorius a II maxillae in *Astacus*, *Pandalus*, and *Callinectes*.

4. MAXILLIPEDS

4a. First Maxilliped

Like the second maxilla, the first maxilliped is a highly modified appendage having a dual function. Its strong gnathal lobes and sensory flagellum participate in feeding, while its flat, lateral lobes and small arthrobranchia play a part in breathing. Superficially, the first maxilliped appears to be as widely modified from the plan of the typical appendage as is the second maxilla, but the muscles indicate otherwise. And whereas the second maxilla is a relatively stable phylogenetic entity in the Crustacea, the form of the first maxilliped is variable.

SKELETAL ELEMENTS

The skeleton of the first maxilliped is comprised of a series of lightly sclerotized lobes or plates. The appendage articulates with the ventral skeleton by a transversely elongate foramen. The foramen enters the fused coxopodite and basipodite (protopodite, fig. 42). Projecting anteroventrally from the protopodite is a set of three gnathal lobes, the large, thick distal lobe being the endite of the coxopodite (fig. 42). Together, the endites of the maxillipeds are cupped against the anterior mouthparts. Heavy spines directed mesad from the edge of the endites help to hold food. Laterad of the coxopodite endite and extending anteriorly is a slender lobe bearing on its medial surface the rudiment of the endopodite and the jointed flagellum of the exopodite. The exites of the coxopodite, lying laterally, are two large, flat sheets that close the gill pump chamber on the ventral surface. Posteriorly, a small, flattened gill may be seen. This gill is said to be an arthrobranchia, but on embryological grounds it might be as easily a podobranch.

The components of the first maxilliped are interpreted in different ways by different authors. The structure called the protopodite in *Penaeus* is termed the coxopodite in *Astacus* by Schmidt. Also, Schmidt considers the coxopodite endite the basipodite, and refers to the coxopodite exites as epipodites. If, as has been discussed earlier, the coxopodite exites develop from podobranch primordia, then no objection to the term "epipodite" can be offered. In passing, we may note that the first maxilliped of *Penaeus* bears two coxopodite endites compared to one in the first maxilliped of *Pandalus*, *Astacus*, and *Callinectes*.

MUSCLE ELEMENTS

Compared to the musculature of the second maxilla, that of the first maxilliped is very light. The muscles function almost entirely in feeding, since the part played by the exites in breathing is mostly passive. Wider variation in the functions of the muscles of the first maxilliped in different crustaceans makes necessary a variety of muscle names, but most of them can be homologized. The first maxilliped of *Penaeus* has 12 muscles, against 9 for *Astacus*, 13 for *Pandalus*, and 11 in *Callinectes*.

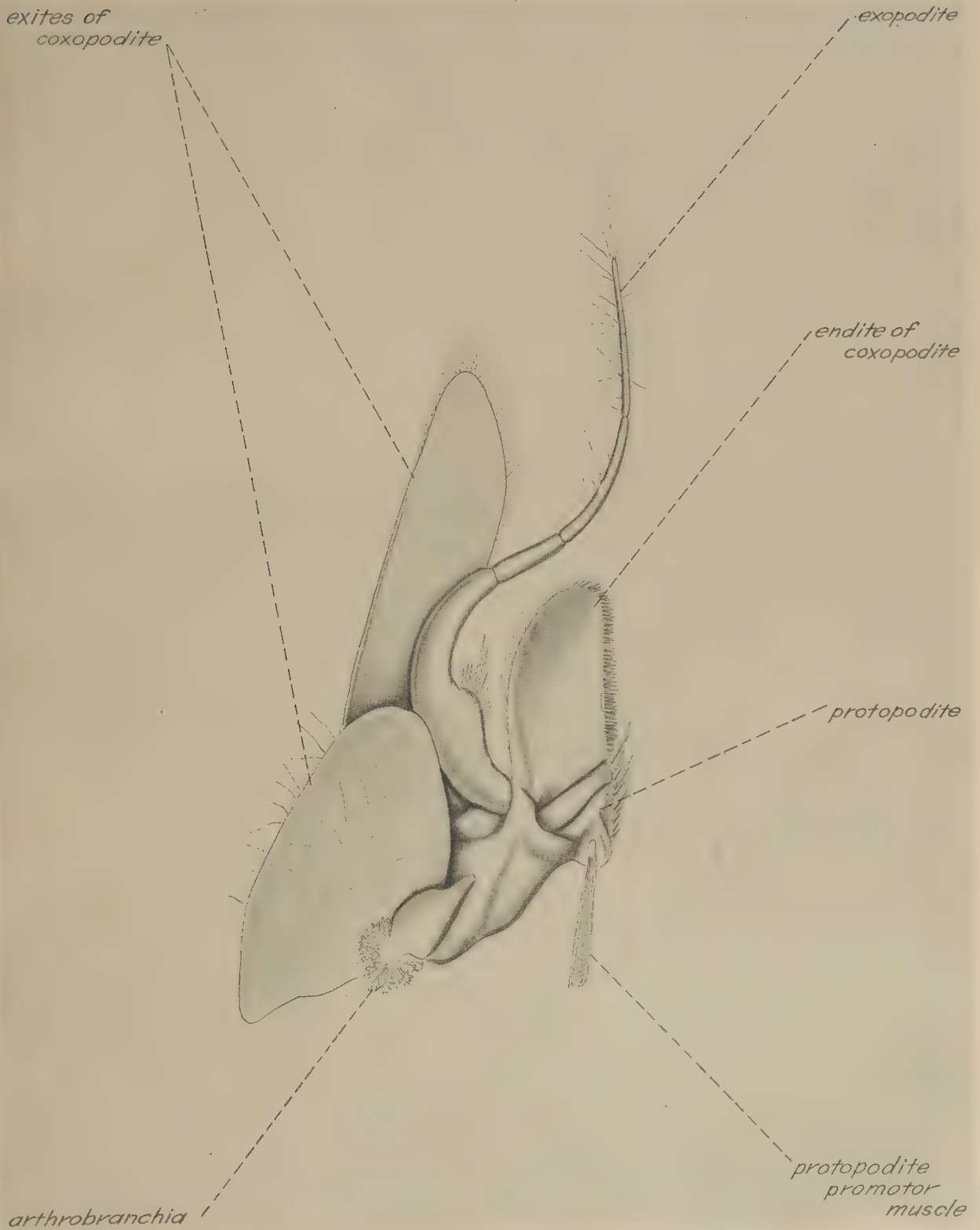


FIGURE 42.—Dorsal view of left first maxilliped.

PROPODITE PROMOTOR MUSCLES OF FIRST MAXILLIPED

FIGURES 42, 43

The propodite promotor muscles (fig. 43) originate on the small sternal apodemes (paraphrags) arising from the sternal brachia between the first and second maxillipeds. The longer, medial promotor inserts on an apodeme on the anterior wall of the propodite and the lateral promotor inserts in connective tissue ventrad of the medial promotor. Contractions of the promoters turn the first maxilliped forward and dorsad about a transverse axis. The propodite promoters of *Penaeus* are very likely homologous with the lateral and medial promotor muscles of *Astacus*, *Callinectes*, and *Pandalus*.

PROPODITE LEVATOR MUSCLES OF FIRST MAXILLIPED

FIGURE 43

Penaeus has at least three propodite levator muscles. All of them take origin on a small sternal apodeme overhanging the medial margin of the maxillipedal foramen. They fan out as they pass laterally to insert in connective tissue in the lateral part of the coxopodite. Contractions of the propodite levators lift the dorsal edges of the coxopodite exites dorsad. At least a part of this muscle group is homologous with the levator muscle of *Pandalus*, *Astacus*, and *Callinectes*.

COXOPODITE ADDUCTOR MUSCLE OF FIRST MAXILLIPED

FIGURE 43

Taking origin in the proximal region of the coxopodite, the coxopodite adductor muscle passes ventrad along the medial wall of the coxopodite to insert on the coxopodite endite. Upon contraction, the muscle turns the endite mesad. The coxopodite adductor in *Penaeus* appears to be the maxillipedal depressor muscle of *Astacus* and *Pandalus*, and possibly one of the small unknown muscles in the coxopodite of *Callinectes*.

COXOPODITE ABDUCTOR MUSCLES OF FIRST MAXILLIPED

FIGURE 43

Two coxopodite abductor muscles exist in the first maxilliped of *Penaeus*. The muscles originate in the lateral part of the coxopodite, near

the posterior exite, and run ventromedially to a point of insertion on the lateral margin of the coxopodite endite. Their contractions draw the endite laterad, in opposition to the coxopodite adductors. The endopodite reductor muscle of *Astacus* is a possible homolog of the coxopodite abductor in *Penaeus*.

EXITE ATTRACTOR MUSCLES OF FIRST MAXILLIPED

FIGURE 43

The first maxilliped of *Penaeus* contains at least two exite attractor muscles which pull the coxopodite exites caudad. The muscles originate on apodemes of the sternal brachia and insert on the medial margin of the posterior coxopodite exite. The epipodite attractor muscle in *Astacus* is very likely the homolog of the exite attractors of *Penaeus*.

EXOPODITE ADDUCTOR MUSCLE OF FIRST MAXILLIPED

FIGURE 43

The flagellum, or exopodite, of the first maxilliped is moved towards the midline by means of a short muscle, the exopodite adductor muscle, which originates at the base of the exopodite and runs distally within the structure. The homolog in *Astacus* is the exopodite adductor muscle.

EXOPODITE ABDUCTOR MUSCLE OF FIRST MAXILLIPED

FIGURE 43

The exopodite abductor muscle originates at the distal end of the exopodite adductor and runs distally in the flagellum. Its contractions tend to straighten the flagellum, thus turning the structure laterad. The muscle in *Penaeus* is in all probability the flagellum muscle of *Astacus*.

4b. Second Maxilliped

The structure of the second maxilliped is much more like that of the typical arthroappendage than the anterior gnathal appendages already treated. The typical number of appendage articles are found, albeit those of the basipodite and ischiopodite are fused. A large, flagellar exopodite is developed. A notable difference is that some of the endopodite articulations of the second maxilliped permit far more extensive movements than do the hinges of the anterior gnathal limbs.

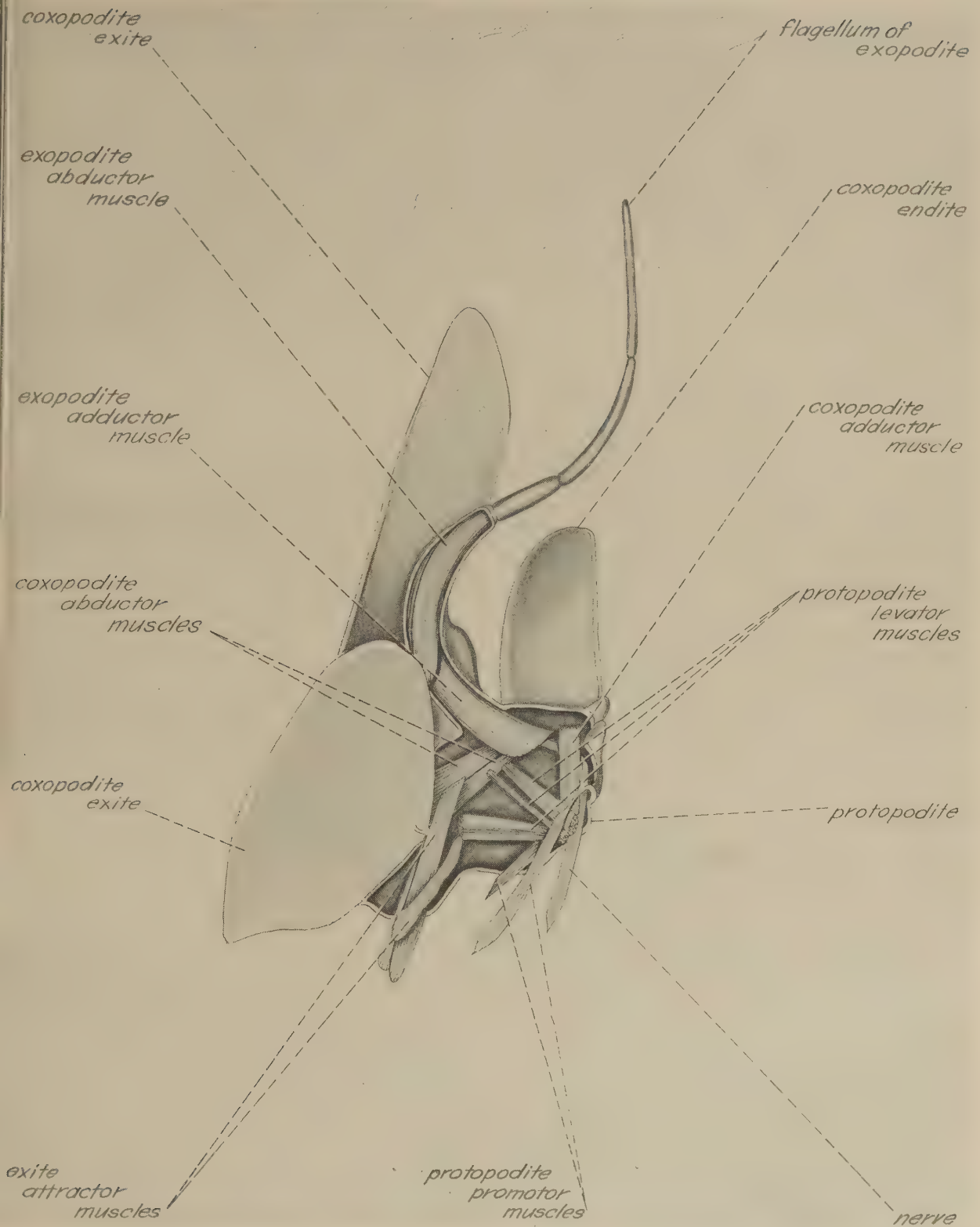


FIGURE 43.—Dorsal view of left first maxilliped, dorsal cuticle removed to show muscles.

The second maxilliped is an active participant in the process of feeding.

SKELETAL ELEMENTS

The coxopodite (fig. 44) of the second maxilliped projects ventrad from a region of the ventral skeleton that is much more heavily sclerotized than are the corresponding areas anteriorly. The foramen of the second maxilliped is surrounded by several large sternal and laterotergal apodemes upon which muscles originate. The whole structure is distinctly heavier than is that of the anterior gnathal appendages, in keeping with the greater movability and strength of the second maxilliped compared to that of the anterior accessory mouth-parts. Projecting laterally from the coxopodite is a small gill, the podobranchia (fig. 44), and a mitten-shaped mastigobranchia, or epipodite. (See fig. 32.) The articular membrane dorsal to the coxopodite bears two arthrobranchiae.

The short, curved carpopodite is connected by dicondyles between the meropodite and the propodite placing the propodite laterad of the distal end of the meropodite. The propodite is a short, square article containing muscles operating the heart-shaped dactylopodite on its distal end. The dactylopodite lies laterad of the proximal part of the meropodite, its apex nearly touching the ischiopodite. The condyles between the carpopodite and propodite and between the propodite and dactylopodite are rotated 90° from the axis of the condyles between the meropodite and the carpopodite. Thus the movements of the distal segments are at right angles to those of the proximal segments.

The coxopodite articulates with the basipodite (fig. 44) by dorsoventral condyles which permit extensive lateral movements. The coxopodite and basipodite are said to be fused in *Pandalus* and *Astacus*. To the basipodite is articulated the exopodite, a long, annular, plumose flagellum (fig. 44) that extends anteriorly and then curves gracefully laterad. The ischiopodite (fig. 44) is immovably fused to the basipodite in *Penaeus*, as in *Callinectes*, but a fine line of light cuticle clearly distinguishes the two articles. The meropodite (fig. 44), the longest article of the endopodite, is attached to the ischiopodite so as to allow the distal segments limited movements from side to side as well as up and down. This article projects anteriorly from the ischiopodite to a position be-

neath the mandibles. Stiff spines on the medial side of the meropodite opposing those on the lateral side of the dactylopodite produce a functional chela or claw.

MUSCLE ELEMENTS

The musculature of the second maxilliped of *Penaeus* is substantially similar to that of *Pandalus*, *Astacus*, and *Callinectes*, although *Penaeus* has a larger number of discrete muscles. The second maxilliped of *Penaeus* contains 14 types of muscles including 23 muscles. *Astacus* has 15 muscle types with 17 muscles. The second maxilliped of *Pandalus* has 14 types of muscles and 16 discrete muscles, whereas *Callinectes* has 16 muscle types and 17 muscles. The classical muscle nomenclature has been changed slightly here as elsewhere to conform to the appendage article in which the muscles insert.

COXOPODITE PROMOTOR MUSCLE OF SECOND MAXILLIPED

FIGURES 44, 45

Taking origin on a paraphragmal apodeme on the lateral pleural wall, the coxopodite promotor muscle passes mesad to insert in connective tissue on the medial wall of the coxopodite. The muscle turns the coxopodite, and with it the distal segments, anterior and dorsad. The musculus promotor II pedis maxillaris of *Astacus*, *Pandalus*, and *Callinectes* is homologous with the coxopodite promotor muscle of *Penaeus*.

COXOPODITE REMOTOR MUSCLE OF SECOND MAXILLIPED

FIGURES 44, 45

The coxopodite remotor muscle originates on a paraphragmal apodeme above the posterolateral margin of the foramen and runs to an insertion in connective tissue on the posterior wall of the coxopodite. Its contractions turn the coxopodite caudad and the distal elements ventrad. The coxopodite remotor muscle in *Penaeus* is fully homologous to the second maxilliped remotor muscle in the other crustaceans mentioned.

BASIPODITE LEVATOR MUSCLES OF SECOND MAXILLIPED

FIGURES 44, 55

The basipodite levator muscles originate at two different points. The medial portion is attached

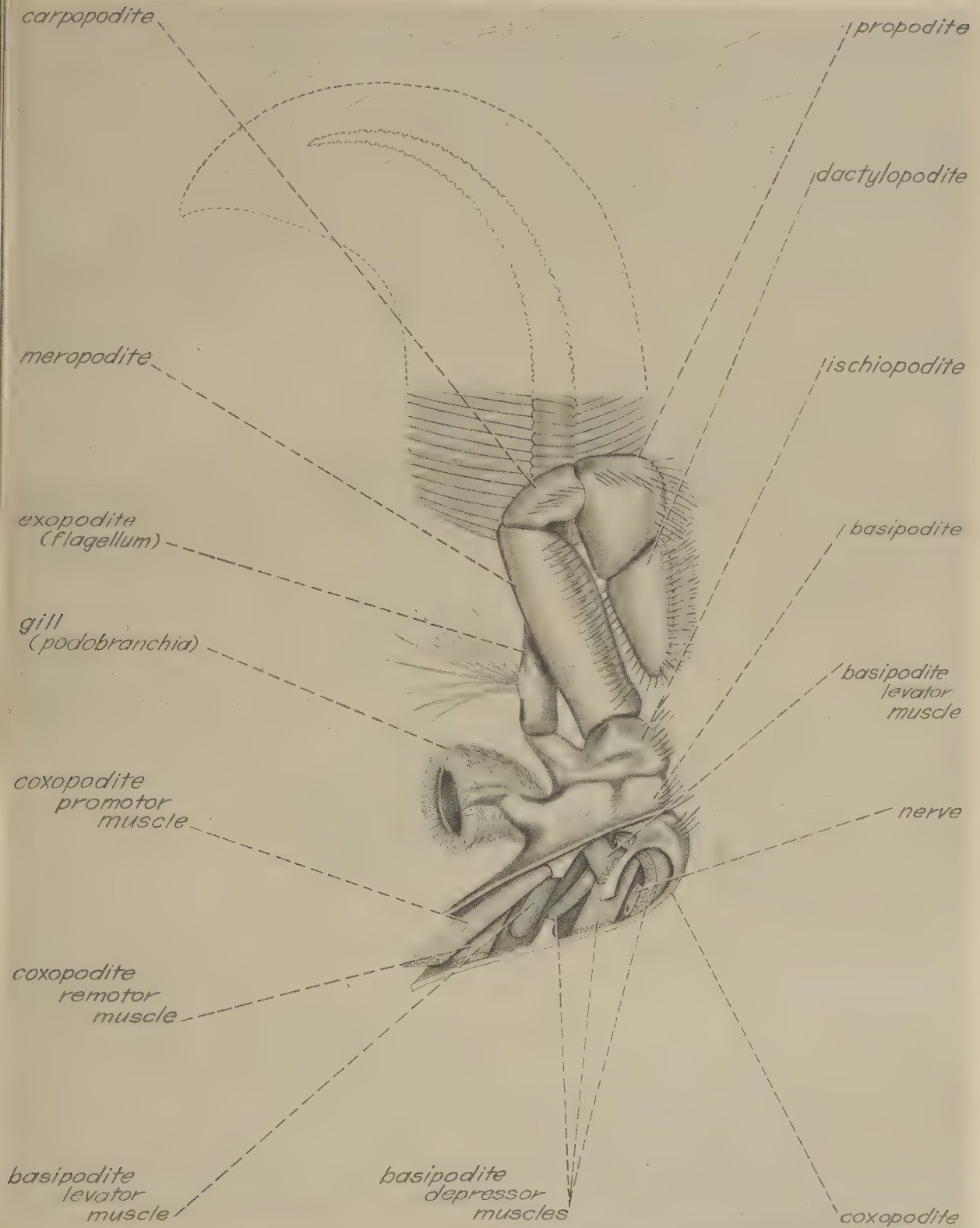


FIGURE 44.—Dorsal view of left second maxilliped.

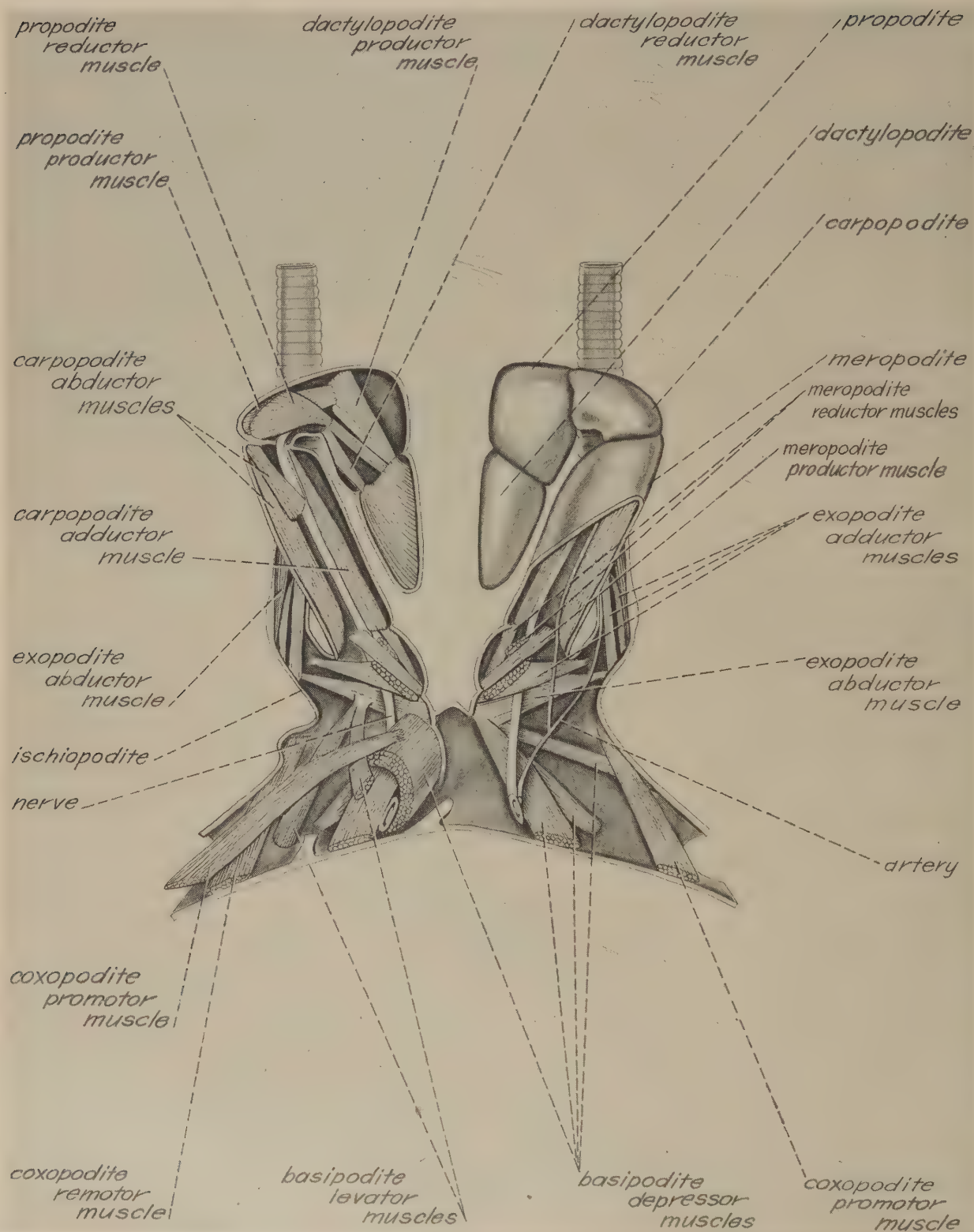


FIGURE 45.—Dorsal view of second maxilliped, cuticle removed to show muscles.

to a large sternal apodeme overhanging the anteromedial part of the foramen. The muscle runs beneath the coxopodite promotor and inserts together with the lateral basipodite levator on the anterior margin of the basipodite foramen, slightly laterad of the dorsal coxobasipodite condyle. The lateral levator muscle originates on the posterior wall of the coxopodite and runs ventrad to join the medial levator. The muscles raise the distal articles and also abduct them. The muscles are homologous with the levator muscles *a* and *b* of *Pandalus* and *Astacus*, and with the single levator in *Callinectes*.

BASIPODITE DEPRESSOR MUSCLES OF SECOND MAXILLIPED

FIGURES 44, 45

The second maxilliped of *Penaeus* contains four basipodite depressor muscles. The largest is the medial depressor, a short, strong, semicylindrical muscle which, in a manner of speaking, lines the medial curvature of the coxopodite. This muscle originates from the medial margin of the dorsal coxopodite foramen and inserts with the other depressors on the ventromedial margin of the coxopodite, mesad of the ventral condyle between the coxopodite and basipodite. Two other basipodite depressors arise, the larger from the posterior margin of the coxopodite foramen, the smaller from the posterior wall of the coxopodite. The fourth depressor muscle originates on the ventrolateral wall of the coxopodite, beneath the coxopodite remotor, and runs across the coxopodite to join the other basipodite depressors. The contractions of the depressor muscles turn the basipodite and thus the distal maxillipedal elements ventrad. In addition, as a consequence of their insertion mesad of the condylic axis, the depressor muscles turn the basipodite and the distal elements towards the midline.

In fact, from the arrangement of the basipodite levators and depressors with respect to the basipodite condyles, the true function of the muscles may be as abductors and adductors, rather than as levators and depressors. The latter names, of course, derive from *Astacus* in which the coxopodite and basipodite of the second maxilliped are fused. On functional grounds, then, Schmidt's names are at least partially incorrect when applied to *Penaeus*. The basipodite depressor muscles of *Penaeus* are nonetheless homologous with

the depressor muscles *a* and *b* in *Pandalus*, *Astacus*, and *Callinectes*.

EXOPODITE ABDUCTOR MUSCLES OF SECOND MAXILLIPED

FIGURE 45

Two exopodite abductor muscles exist in the second maxilliped of *Penaeus*. The medial abductor is extrinsic, originating broadly on the medial wall of the basipodite and inserting on the posterior edge of the exopodite base. The extrinsic abductor turns the exopodite laterad. Intrinsic to the exopodite is a long abductor muscle which originates by a fine tendon on the base of the exopodite and extends distally for some distance in the exopodite flagellum. Its contractions bend the flagellum laterad. The intrinsic exopodite abductor muscle in *Penaeus* is homologous with the exopodite abductor muscle of *Astacus* and *Callinectes*, and the extrinsic abductor is probably the same muscle as the flagellum abductor in *Astacus*.

EXOPODITE ADDUCTOR MUSCLES OF SECOND MAXILLIPED

FIGURE 45

The second maxilliped of *Penaeus* has at least three exopodite adductor muscles. The extrinsic exopodite adductor originates in a broad fan across the dorsomedial wall of the basipodite and inserts on the anterior edge of the exopodite base. When it contracts, the exopodite is turned towards the midline. Within the exopodite are two exopodite adductors which function to straighten out the exopodite flagellum, in opposition to the action of the intrinsic exopodite abductor muscle. No homolog of the extrinsic exopodite adductor muscle is evident in *Astacus* or *Callinectes*. Berkeley (1928) illustrates an extrinsic adductor of the exopodite, but does not discuss the muscle. The intrinsic exopodite adductors of *Penaeus* apparently have no homologs in the crustaceans referred to here.

MEROPODITE PRODUCTOR MUSCLE OF SECOND MAXILLIPED

FIGURE 45

The types and arrangement of the second maxilliped endopodite muscles are remarkably uniform in many higher Crustacea, in accordance

with the evolution of this versatile accessory feeding mechanism. The meropodite productor muscle originates on the medial side of the ischiopodite, near the exopodite adductor, and inserts on a small apodeme on the dorsal surface of the meropodite. The muscle lifts the meropodite and other distal elements towards the ventral surface of the mandibles. The same muscle appears in *Pandalus*, *Astacus*, and *Callinectes*.

MEROPODITE REDUCTOR MUSCLES OF SECOND MAXILLIPED

FIGURE 45

Penaeus has two meropodite reductor muscles. The medial reductor originates on the medial wall of the ischiopodite, ventrad of the meropodite productor, and inserts on an apodeme on the ventral margin of the meropodite. Another reductor muscle originates on the anterodorsal surface of the ischiopodite and passes to the same apodeme as the medial reductor. The muscles turn the meropodite ventrad. The meropodite reducers of *Penaeus* have partial counterparts in the second maxilliped of *Astacus*, *Pandalus*, and *Callinectes*, but in the latter only one reductor has been described.

CARPOPODITE ABDUCTOR MUSCLES OF SECOND MAXILLIPED

FIGURE 45

Two carpopodite abductor muscles are found in the second maxilliped of *Penaeus*. The smaller of these originates on the dorsal side of the meropodite and inserts on the common abductor apodeme on the lateral edge of the carpopodite. The larger abductor takes origin on the lateral side of the meropodite, proximally. The muscle runs out along the lateral side of the meropodite and attaches to the abductor apodeme on the carpopodite. The two muscles turn the small carpopodite laterad. However, with respect to the gnathal surface of the dactylopodite, the carpopodite abductors cause functional adduction, in consequence of the hooked shape of the endopodite. The large carpopodite abductor muscle is the same muscle as the carpopodite abductor in *Astacus*, *Pandalus*, and *Callinectes*. The small abductor in *Penaeus* is not described in any of the foregoing crustaceans.

CARPOPODITE ADDUCTOR MUSCLE OF SECOND MAXILLIPED

FIGURE 45

The carpopodite adductor muscle runs from the proximal end of the meropodite to an adductor apodeme on the medial side of the carpopodite. The muscle is less powerful than the carpopodite abductors. The adductor turns the carpopodite mesad, but as a result of the distal hook, the gnathal surface of the dactylopodite is moved laterad. The carpopodite adductor of *Penaeus* is fully homologous with the carpopodite adductor in the other crustaceans to which reference has been made.

PROPODITE PRODUCTOR MUSCLE OF SECOND MAXILLIPED

FIGURE 45

The propodite productor muscle is a short, thick structure originating on a proximal, ventral part of the carpopodite, and running to a broad apodeme on the ventrolateral side of the propodite. Its contractions turn the propodite and dactylopodite ventrad. The propodite productors in *Penaeus* are homologous with the propodite productor in *Astacus*, *Pandalus*, and *Callinectes*.

PROPODITE REDUCTOR MUSCLE OF SECOND MAXILLIPED

FIGURE 45

The propodite reductor muscle occupies the dorsal part of the carpopodite in the shape of a fan. The broad portion originates in the proximal region of the carpopodite. The muscle becomes narrow as it inserts on a little apodeme on the dorsal side of the propodite. The muscle turns the propodite and dactylopodite dorsad, about the horizontal axis through the condyles. The propodite reductor muscle of *Penaeus* is represented in the second maxilliped of the three crustaceans to which frequent reference has been made.

DACTYLOPODITE PRODUCTOR MUSCLE OF SECOND MAXILLIPED

FIGURE 45

The dactylopodite productor muscle is a small, fan-shaped muscle that originates proximally in the propodite and inserts on the ventral margin of the dactylopodite. It serves to straighten the

dactylopodite and turn the distal article ventrad, in opposition to the dactylopodite reductor. The same muscle as the dactylopodite productor in *Penaeus* is found in *Pandalus*, *Astacus*, and *Callinectes*.

DACTYLOPODITE REDUCTOR MUSCLE OF SECOND MAXILLIPED

FIGURE 45

Attached between the proximal part of the propodite and the dorsal edge of the dactylopodite is the small dactylopodite reductor muscle. Its function is to turn the dactylopodite dorsad, opposing the action of the productor. The muscle is homologous with the dactylopodite reductor of *Pandalus*, *Astacus*, and *Callinectes*.

4c. Third Maxilliped

The third maxilliped is the first accessory feeding appendage which lacks the jawlike characteristics of the anterior gnathal limbs. Far more than any of the other appendages, the third maxillipeds function to grasp large food particles passed up by the chelate legs and hold them next to the mouthparts for further reduction in size and for swallowing. Structurally, the third maxilliped is closely similar to the pereopods.

SKELETAL ELEMENTS

The body hemocoel is confluent with that of the third maxilliped by means of a ventral skeletal foramen whose fringes are strongly sclerotized and from which phragmal apodemes project over the opening. The sternal plate between the foramina is wider at this point than that between the anterior mouthpart foramina. The heavily sclerotized coxopodite articulates with the ventral skeleton by a dorsal, laterotergal condyle and a ventral sternal condyle, the typical situation in the white shrimp limb. The axis through the condyles is about 45° from the vertical, with respect to the transverse plane. Since the distal elements are anterior, movements about these coxopodite condyles raise and lower the appendage, as well as promote and remote it. The basipodite articulates with the coxopodite by typical dicondylar connections, the axis of the condyles being horizontal. Thus the basipodite accounts for most of the depression and levation of the distal elements, an arrangement that is common to the third maxil-

iped and all of the walking legs. A large, tapering, annulated exopodite articulates with the basipodite laterally.

The ischiopodite is fused to the basipodite immovably, although the line of demarcation is clear. The ischiopodite is the longest article of the third maxilliped. The meropodite is connected to the distal end of the ischiopodite by two condyles whose axis permits both flexion of the distal articles towards the midline and reduction of the distal elements. The movements at this joint are extensive. Due to the bending at this joint the shrimp is able to grasp food with the third maxilliped. The carpopodite articulates with the distal end of the meropodite by two condyles. The axis of these dicondyles is vertical, permitting the carpopodite to flex on the meropodite.

The axis through the condyles of the joint between the carpopodite and the propodite is also vertical allowing the propodite to be flexed upon the carpopodite. The small dactylopodite is similarly articulated with the propodite.

The distal elements, beginning with the ischiopodite, bear rows of stout spines on their medial sides for holding food particles. The exopodite has long, plumose setae. The third maxilliped has a branchial arrangement similar to that of the first three walking legs. From the pleural plate arises a pleurobranchia. Two arthrobranchiae project from the dorsal articular membrane, and a bilobed mastigobranchia arises from the coxopodite.

MUSCLE ELEMENTS

The musculature of the third maxilliped of *Penaeus* is typical of that seen in the walking legs. Some variation, however, is evident when the third maxilliped of different crustaceans is compared, especially in the musculature of the exopodite and the distal articles of the endopodite. In *Penaeus*, most of the basipodite and coxopodite muscles originate on the laterotergal, pleural plates rather than on phragmal elements as in the anterior gnathal appendages. Pleural origins of these muscles are typical of all the posterior appendages, thoracic and abdominal alike.

The third maxilliped of *Penaeus* is operated by twenty muscles comprising 12 functional types. That of *Astacus* contains 17 muscles, including 14 types. The third maxilliped of *Pandalus* is somewhat modified, containing 13 muscles of 9 functional types, whereas the same appendage in

Callinectes has 16 muscles grouped into 15 muscle types.

COXOPODITE PROMOTOR MUSCLE OF THIRD MAXILLIPED

FIGURES 46, 47, 48

The coxopodite promotor muscle is a strong, lozenge-shaped muscle (fig. 47, *B*) originating on the pleural plate along the anterodorsal margin of the muscle. The insertion is on the anterior margin of the coxopodite (fig. 46). The promotor turns the coxopodite forward and, since the distal parts extend directly anterior, the latter are raised against the other mouthparts. The promotor muscles of *Pandalus*, *Astacus*, and *Callinectes* are the counterparts of the coxopodite promoters in *Penaeus*.

COXOPODITE REMOTOR MUSCLES OF THIRD MAXILLIPED

FIGURES 46, 47

The third maxilliped of *Penaeus* contains two coxopodite remotor muscles. They are much more powerful than the coxopodite promotor muscles which they oppose. The remotors originate on the the laterotergal plate dorsad (fig. 47) and insert on the posterior margin of the coxopodite. They turn the coxopodite caudad and thus the distal elements ventrad as well as laterally. The larger, lateral coxopodite remotor of *Penaeus* is fully homologous with the remotors in the three crustaceans to which we have referred.

BASIPODITE LEVATOR MUSCLES OF THIRD MAXILLIPED

FIGURES 46, 47

The two basipodite levator muscles that have been found in *Penaeus* have different origins. The short, stout lateral levator takes origin on the anterolateral margin of the coxopodite and inserts on the anterior margin of the basipodite. The larger basipodite levator lies internal to the smaller muscle. The origin of the former is on the laterotergal plate and its insertion on the anterior rim of the coxopodite. The levators pull the anterior side of the basipodite upward and with it the distal maxillipedal elements. Three levators of the third maxilliped are described in *Pandalus*, *Astacus*, and *Callinectes*. Part of this group is very likely homologous with the levator pair in *Penaeus*.

BASIPODITE DEPRESSOR MUSCLES OF THIRD MAXILLIPED

FIGURES 46, 47, 48

Considerable variation from the pattern of the depressor musculature of *Penaeus* is seen in the third maxillipeds of *Astacus*, *Pandalus*, and *Callinectes*. Schmidt and Berkeley describe two depressors of the third maxillipeds of *Astacus* and *Pandalus*, whereas Cochran shows one depressor in *Callinectes*. By contrast, *Penaeus* has at least four basipodite depressor muscles in the third maxilliped, just as in the second maxilliped.

The lateral basipodite depressor takes origin on the posterolateral margin of the coxopodite (fig. 47) and inserts, together with the other depressors, on the large apodeme on the posterior rim of the basipodite. Interior to the lateral basipodite depressor is a long, two-part depressor muscle (fig. 47, *B*) originating on the laterotergal plate mesad of the coxopodite remotor muscle. This depressor joins the short lateral depressor on the basipodite depressor apodeme. Immediately mesad of the long depressor lies a small, flat basipodite depressor muscle (figs. 46; 47, *A*) which originates on a medial apodeme of the maxillipedal foramen and inserts on the basipodite apodeme. The most internal basipodite depressor muscle (figs. 46, 48) is a semicylindrical structure originating on medial phragmal apodemes and inserting on the posteromedial edge of the basipodite.

The total action of the basipodite depressor muscles is relatively powerful. By their contractions the basipodite and distal elements of the third maxilliped are turned ventrad. The homologies of these muscles with those of *Astacus*, *Pandalus*, and *Callinectes* are not entirely clear. The depressor muscle *a* of *Astacus* and *Pandalus* is the same muscle as the innermost, medial basipodite depressor muscle in *Penaeus*. Undoubtedly, the depressor muscles *a*₁ and *b* in *Astacus*, and *a*₁, *a*₂ and *b* in *Pandalus* have counterparts in the third maxilliped of *Penaeus*, but their exact relationships are difficult to determine.

EXOPODITE ABDUCTOR MUSCLES OF THIRD MAXILLIPED

FIGURES 46, 47

Similar to the arrangement in the second maxilliped, the third maxilliped exopodite of *Penaeus* is moved by 2 exopodite abductor muscles, 1 ex-

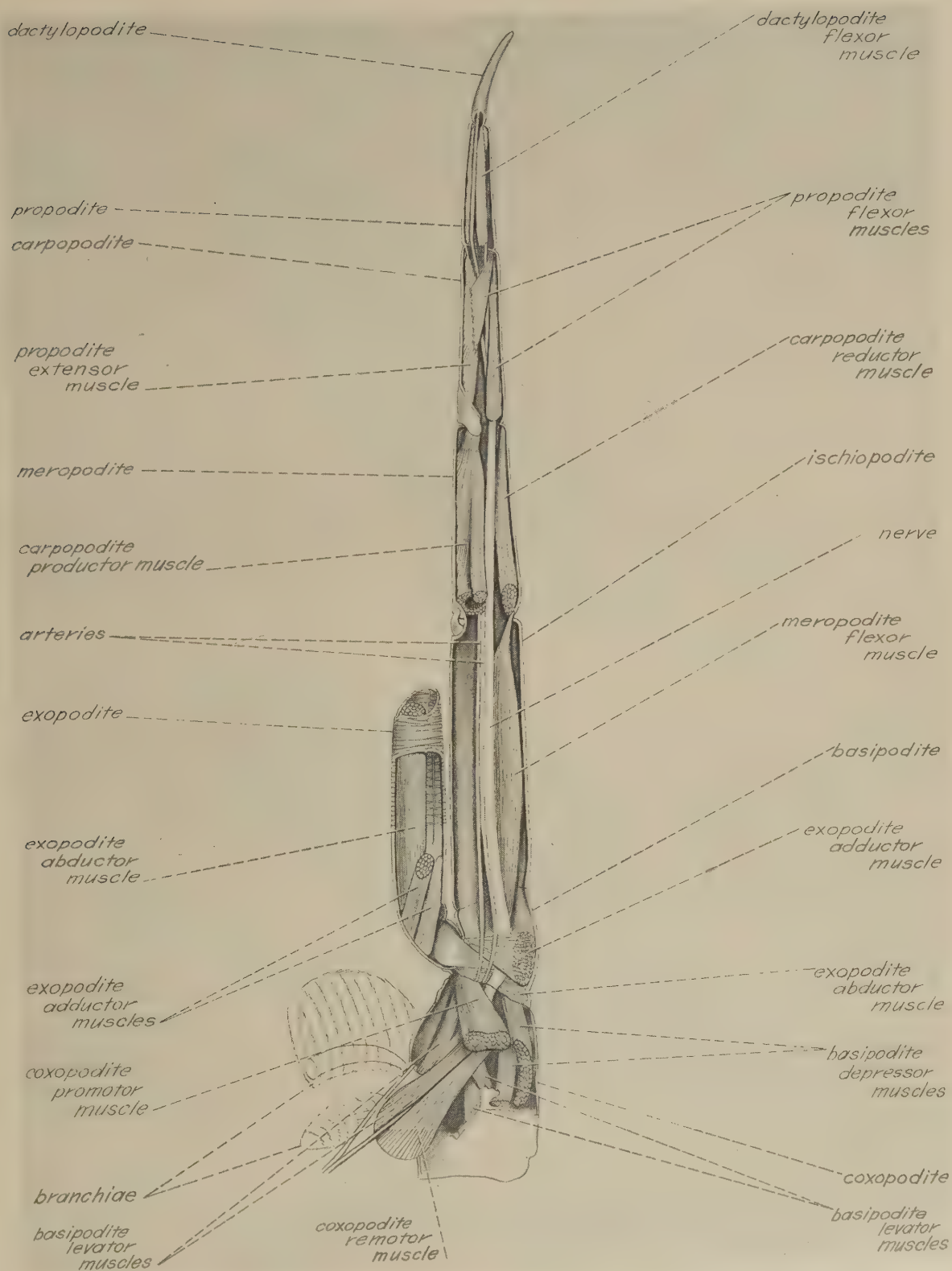


FIGURE 46.—Dorsal view of left third maxilliped, dorsal cuticle removed to show muscles.

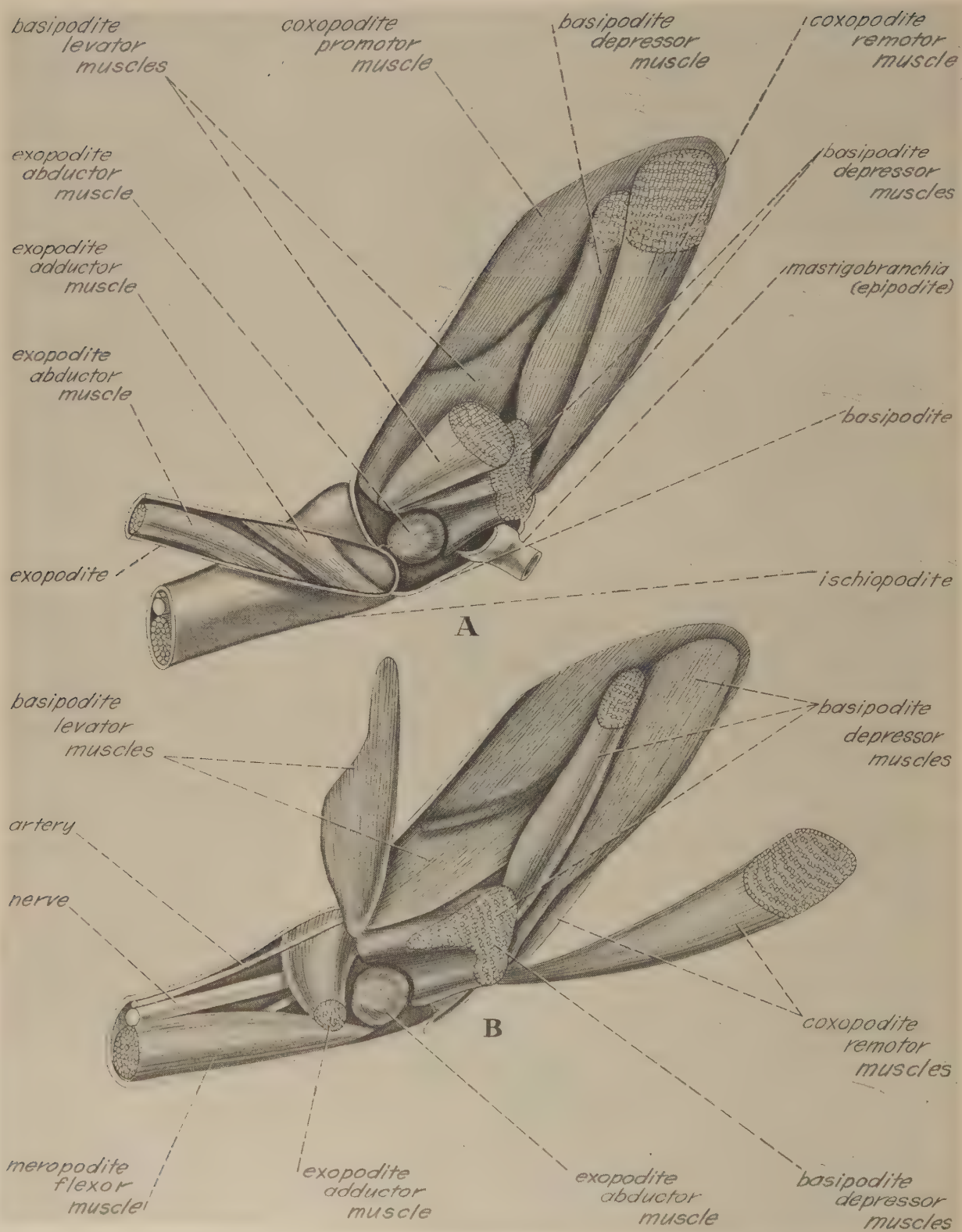


FIGURE 47.—Lateral view of leg base of left third maxilliped. Cuticle removed to show muscles. A. Superficial lateral and exopodite muscles. B. Lateral muscles, exopodite removed.

trinsic and 1 intrinsic to the structure. The wedge-shaped extrinsic abductor (figs. 46; 47, *B*) originates on the ventromedial wall of the basipodite. The muscle tapers to a point, laterally, inserting on the posterior margin of the exopodite foramen. Contractions of the exopodite abductor turn the exopodite laterad. Intrinsic to the flagellar exopodite is a long abductor muscle (fig. 46) which originates in the proximal region of the exopodite and runs distally along the lateral side of the flagellum. Its contractions increase the lateral curvature of the flagellum, thus turning it laterad. The extrinsic exopodite abductor muscle of *Penaeus* is represented under the same name in *Astacus* and *Callinectes*. The exopodite of *Pandalus* is extremely reduced. The intrinsic exopodite abductor probably appears as the exopodite flagellum muscle in *Astacus* and *Callinectes*.

EXPODITE ADDUCTOR MUSCLES OF THIRD MAXILLIPED

FIGURES 46; 47, *B*

Two exopodite adductor muscles in *Penaeus* turn the exopodite flagellum mesad. The conical extrinsic adductor originates on the dorsomedial wall of the basipodite, dorsad of the extrinsic exopodite abductor muscle (fig. 46), and inserts on the anterior rim of the exopodite foramen. The intrinsic exopodite adductor is comprised of a pair of small muscles originating on the posterolateral side of the flagellum base and attaching a short distance distally on the medial and dorsal wall of the flagellum. The extrinsic adductor has a counterpart in *Callinectes*, but not in *Astacus*. The little intrinsic adductor muscle in *Penaeus* possibly may be the exopodite flagellum abductor muscle of *Astacus* and *Callinectes*, although this is doubtful.

MEROPODITE FLEXOR MUSCLE OF THIRD MAXILLIPED

FIGURES 46, 47

The meropodite flexor muscle is a long, spindle-shaped muscle arising in the basipodite and passing distally along the lateral wall of the ischiopodite to an apodemal insertion on the proximal end of the meropodite. The apodeme of insertion is located ventromedially. When the meropodite flexor contracts, the meropodite and distal articles of the endopodite are turned sharply mesad and,

due to the arrangement of the condyles, ventrad. The ischiopodite-meropodite junction is the major functional joint in the third maxilliped. The meropodite flexor muscle of *Penaeus* is fully homologous with the meropodite flexor muscle of *Astacus* and *Callinectes*. The third maxilliped endopodite of *Pandalus* has lost parts by fusion, making difficult the homologies of its muscles.

CARPOPODITE PRODUCTOR MUSCLE OF THIRD MAXILLIPED

FIGURE 46

The carpopodite productor muscle takes origin over an extensive area along the lateral side of the meropodite (fig. 46) and inserts on an apodeme of the carpopodite. Its contractions straighten the carpopodite with respect to the meropodite. The muscle is represented in the third maxilliped of *Astacus* and apparently in *Pandalus*. The productor muscle is referred to as an extensor by Cochran in *Callinectes* due to a difference in condylic orientation.

CARPOPODITE REDUCTOR MUSCLE OF THIRD MAXILLIPED

FIGURE 46

Opposing the action of the carpopodite productor muscle is the carpopodite reductor muscle. This muscle originates in the proximal region of the meropodite and passes distally along the medial side of the meropodite to insert on a ventromedial apodeme of the carpopodite. Upon contraction, the muscle turns the carpopodite and the distal articles ventrad. The carpopodite reductor muscle of *Penaeus* is the same muscle in *Astacus*, *Pandalus*, and *Callinectes*, although in the latter Cochran describes it as a flexor.

PROPODITE EXTENSOR MUSCLE OF THIRD MAXILLIPED

FIGURE 46

Originating for some length along the lateral margin of the carpopodite, the propodite extensor muscle inserts on a laterally located apodeme of the propodite. The muscle extends the propodite and dactylopodite directly anterior to the carpopodite. The same muscle is found in all the crustaceans to which reference has been made.

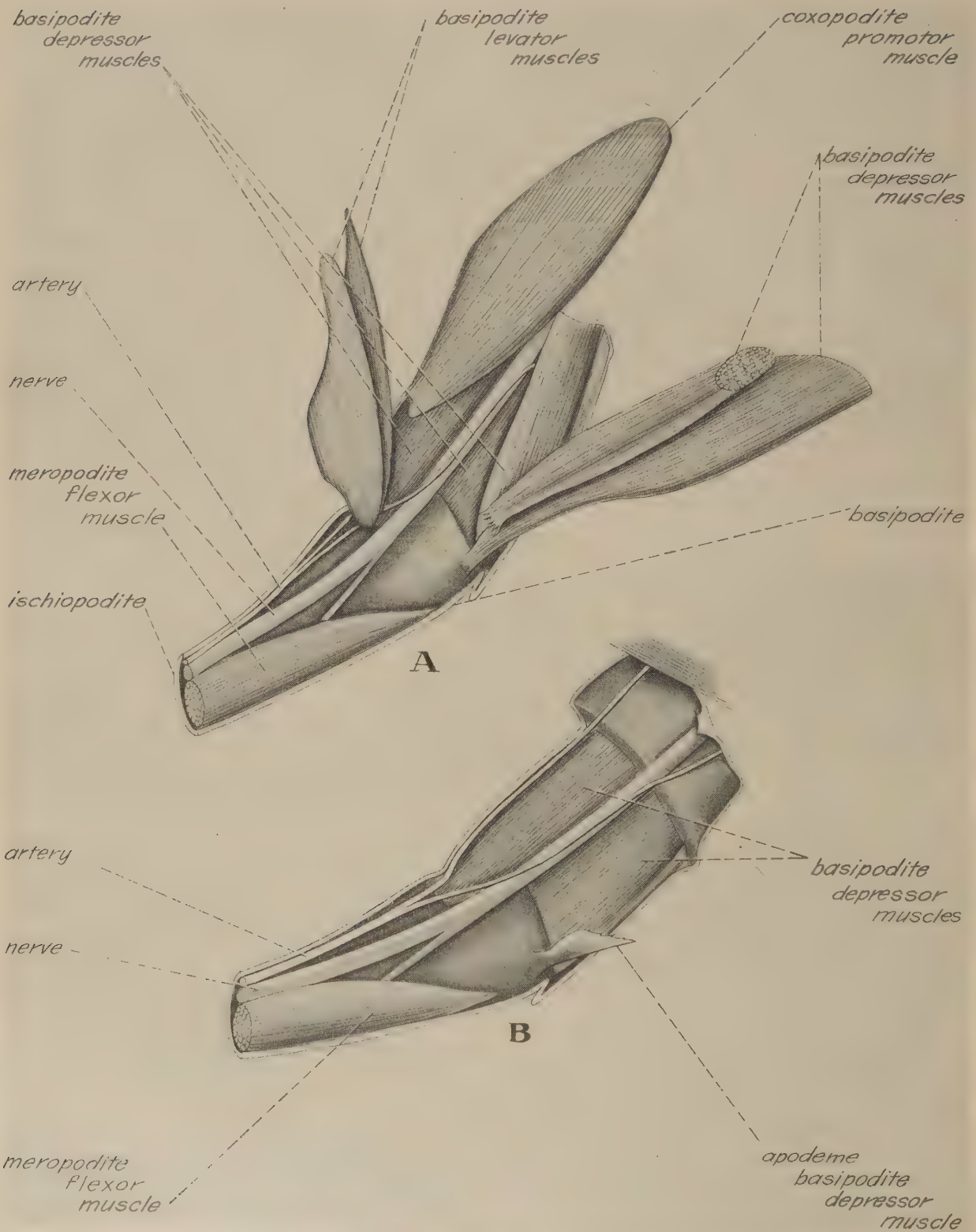


FIGURE 48.—Lateral view of leg base of left third maxilliped. A. Lateral and medial muscles. B. Medial muscles.

PROPODITE FLEXOR MUSCLES OF THIRD MAXILLIPED

FIGURE 46

The third maxilliped of *Penaeus* is unique by comparison with the same appendage of *Pandalus*, *Astacus*, and *Callinectes* in having two propodite flexor muscles. The larger originates in the carpopodite proximally and passes distally on the lateral side of the article to its insertion on an apodeme of the propodite. The smaller propodite flexor is a little triangular muscle (fig. 46) which has a common insertion with the larger flexor. The flexor muscles turn the propodite mesad on the carpopodite. The larger propodite flexor muscle is found in all the crustaceans referred to.

DACTYLOPODITE FLEXOR MUSCLE OF THIRD MAXILLIPED

FIGURE 46

The dactylopodite flexor muscle arises in the proximal portion of the propodite and is attached to an apodeme of the dactylopodite. The muscle turns the dactylopodite toward the midline. Dactylopodite flexors appear in the third maxilliped of *Astacus* and *Callinectes*. The distal articles are fused in *Pandalus* and the muscles thus lost. The third maxilliped of *Penaeus* has no dactylopodite extensor muscle, a structure described in *Astacus* and *Callinectes*.

5. PEREIOPODS

In the Crustacea Decapoda the last five pairs of thoracic appendages are usually referred to as walking legs, or pereopods. Although their length, size, and functional modifications are variable in the group, the walking legs are all fundamentally alike in structure. Exopodites, usually small, are either present or absent. The large propodite-endopodite is almost always composed of the typical seven appendage articles, although fusion of the basipodite with the ischiopodite has occurred in some groups. Some or all of the pereopods may be chelate. Incidentally, Dougherty (Steinberg and Dougherty, 1957) objects to the spelling of the word "pereopod" and offers good reasons for dropping the *i* in American usage. The common and perhaps incorrect spelling is used in the present work.

In *Penaeus*, the pereopods are all relatively long and slender as befits a lightly sclerotized form whose body weight on the walking legs is not

great. The first three pereopods bear small chelae with sharp cutting edges. The last two walking legs are subchelate. Small exopodites are found on all five pairs of pereopods. The joint between the basipodite and ischiopodite is movable in the walking legs of *Penaeus*. The coxopodite and basipodite comprise the protopodite to which an endopodite with the typical five articles is attached. Associated with the pereopods are branchiae. With the third maxillipeds, the first, second, and third pereopods have a bilobed mastigobranchia (epipodite) arising from the coxopodites, 2 arthrobranchiae on the dorsal articular membrane, and 1 pleurobranchia on the laterotergal plate. The mastigobranchia and one arthrobranchia is missing on the fourth pereopod and the fifth pereopod has only a pleurobranch.

Due to pronounced serial similarity of parts in the walking legs, only the first and fifth pereopods will be considered in detail here.

5a. First Pereiopod

The first pereiopod is considered as an example of a chelate limb. It is ordinarily treated as the first walking leg. Whereas in the Reptantia the first pereiopods are modified into huge chelate chelipeds, those of the Penaeidae, while chelate, are similar in size to the remaining four pairs of walking legs. In point of fact, the first pereiopod of *Penaeus* is not a functional walking leg. That is to say, none of the body weight is supported on the appendage. The first pereiopods are carried horizontally, directed anteriorly, ventrad of the third maxillipeds, and function to pass food particles to the latter.

SKELETAL ELEMENTS

The strongly sclerotized coxopodite articulates with the ventral skeleton by dorsoventral condyles. The axis through the condyles is such that when the coxopodite swings forward it also approaches the midline. The rearward motion, on the other hand, turns the coxopodites away from the midline. The angular attitude of the condylic axis of the first pereiopod is thus raised laterally with respect to the frontal plane and rostrad with respect to the transverse plane. Proceeding caudad, the angular attitude of the axis through the coxopodite condyles rises laterally with respect to the frontal plane in conjunction with the shift dorsad of the dorsal condyle and the increased width between the ventral sternal

condyles. To the coxopodite is attached the strong, curving tube of the basipodite. The axis of the condyles between the coxopodite and basipodite is rotated 90° from the axis of the condyles connecting the coxopodite and the ventral skeleton. Primarily by means of the basipodite, the distal elements of the pereopod are raised and lowered. A sharp process projects distally from the distomedial portion of the basipodite. A small, fingerlike exopodite bearing long setae projects laterad from the basipodite.

The ischiopodite is hinged to the basipodite to permit limited reduction of this article, and also some rotation of the distal elements due to the oblique angle of the condyles. Like the basipodite, the ischiopodite bears a sharp spine medially. The meropodite, one of the longer articles of the first pereopod, articulates with the distal end of the ischiopodite, allowing some flexion and extension, together with a little reduction, of the distal segments. The most movable joint in the first walking leg is that connecting the meropodite and the carpopodite. The axis of the joint is horizontal, and the carpopodite is thereby capable of deep flexion on the meropodite. An antennal cleaning brush composed of comb-like setae resides in a distomedial depression of the carpopodite.

The chela is freely movable on the carpopodite. The propodite component is made up of a base element and a distal process (fig. 49). The dactylopodite articulates with the distal part of the base of the propodite, opposing the distal propodite process. Extremely fine calcareous teeth on the inner margins of the propodite process and dactylopodite enhance the cutting ability of the chela. Chemosensory pits from which project chemoreceptor tufts of fine setae are distributed in rows about the propodite process and dactylopodite surface.

MUSCLE ELEMENTS

The musculature of the first pereopod is closely similar to that of the third maxilliped and the remaining walking legs. It is also similar to the pereopod musculature of other decapods. The first pereopod of *Penaeus* contains at least 21 discrete muscles arranged in 14 functional muscle types. *Astacus* has 19 muscles of 13 types, and *Callinectes* has 20 muscles functioning in 11 ways. The third pereopod of *Pandalus* contains 16 muscles of 13 types.

COXOPODITE PROMOTOR MUSCLES OF FIRST PEREIOPOD

FIGS. 49, 50, 51

The first pereopod of *Penaeus* possesses at least two coxopodite promotor muscles. They originate by broad regions along the anterodorsal margin of the pleural plate belonging to the segment and insert on the anterior rim of the coxopodite. Their contractions turn the coxopodite forward. The coxopodite promotors of *Penaeus* are represented in *Astacus*, *Pandalus*, and *Callinectes*.

COXOPODITE REMOTOR MUSCLE OF FIRST PEREIOPOD

FIGS. 49, 50, 51

The strong, spatulate coxopodite remotor muscle originates on the posterodorsal part of the laterotergal plate and attaches to a small apodeme on the posterior margin of the coxopodite foramen. The muscle turns the coxopodite rearward and in so doing brings the distal elements of the pereopod ventrad, in opposition to the coxopodite promotors. The movement is evidently weaker than that of promotion, since the single remotor is much smaller than the promotor muscles. The homologies of the coxopodite remotor muscle of *Penaeus* with the remotor muscle of *Callinectes* are fairly obvious. *Astacus*, however, has three remotors. Which of these is homologous with the coxopodite remotor of *Penaeus* is not certain.

BASIPODITE LEVATOR MUSCLES OF FIRST PEREIOPOD

FIGURES 49, 50, 51

A characteristic of pereopod musculature is the great weight and strength of the basipodite musculature, affording powerful levation and depression of that article and with it the distal elements of the leg. The situation in the Decapoda is reminiscent of the trochanteral musculature of the insects, except that the levators and depressors of the latter originate within the coxa, rather than on the pleural wall. Functionally, the basipodite depressors are the major support of the body between the walking legs.

The first walking leg of *Penaeus* has 4 basipodite levator muscles, of which 3 originate within the coxopodite and 1 dorsally on the pleural wall.

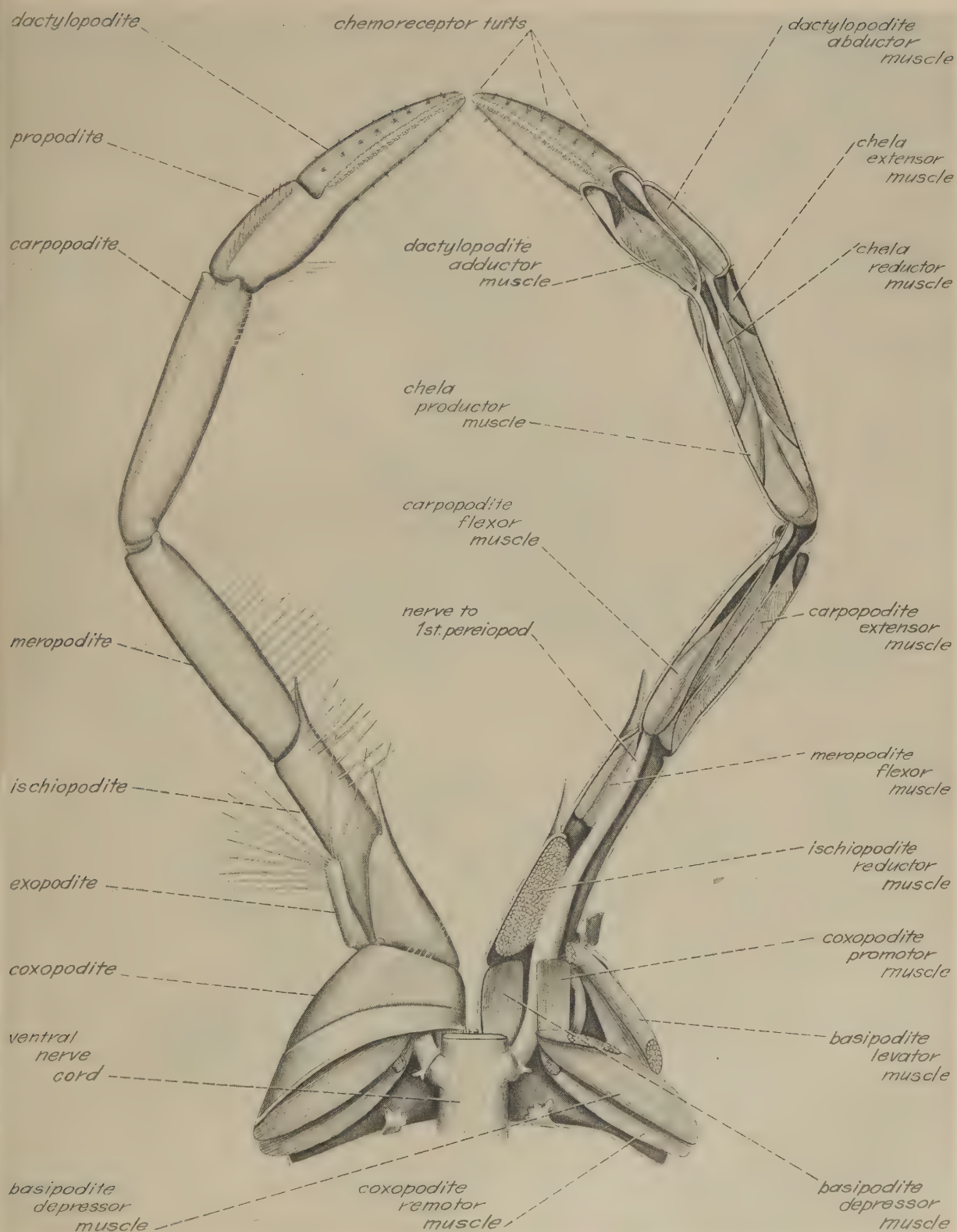


FIGURE 49.—Dorsal view of first pereiopod. Left side, intact appendage. Right side, dorsal cuticle removed to show muscles.

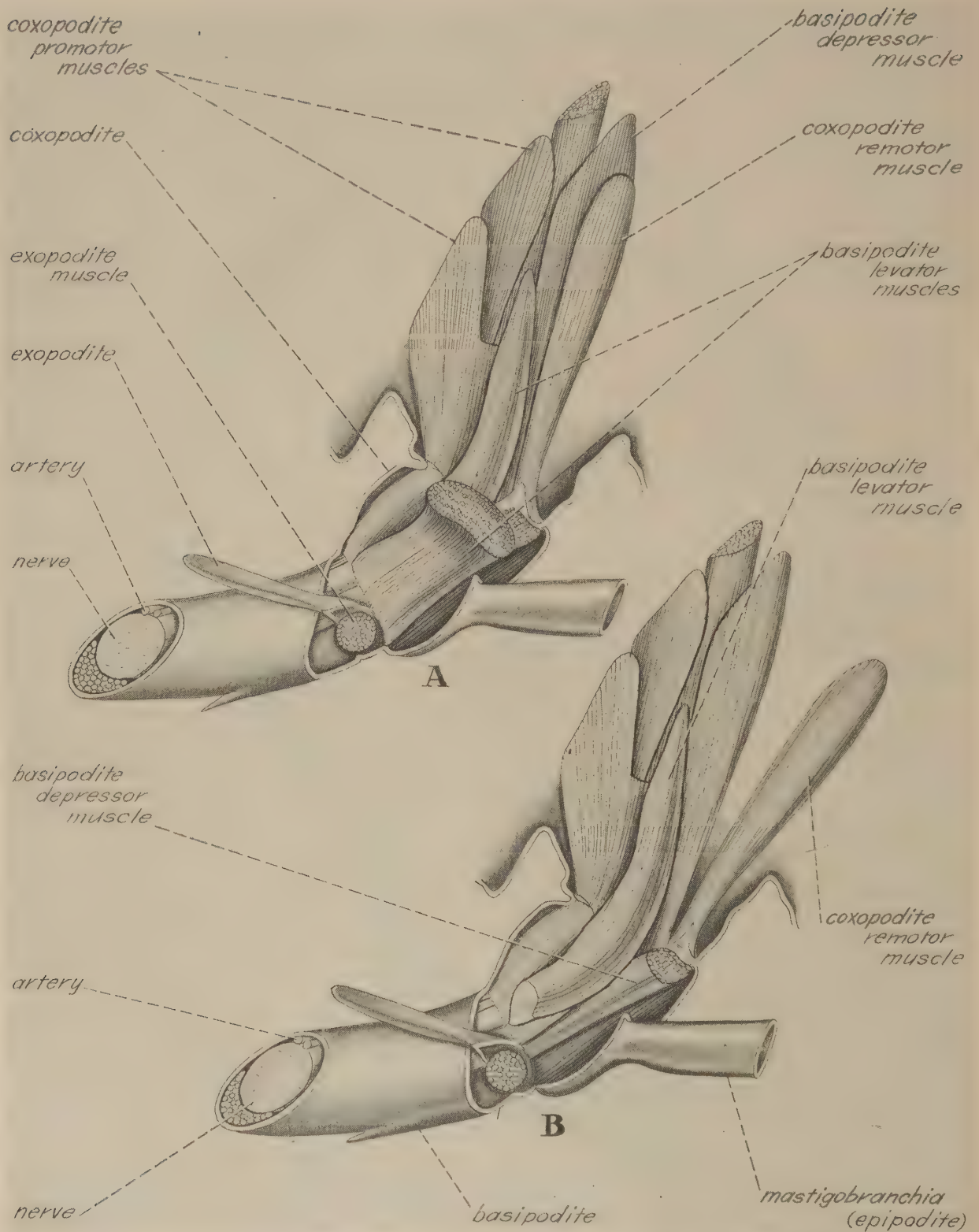


FIGURE 50.—Lateral view of leg base of left first pereopod. A. Superficial lateral muscles. B. Some lateral muscles removed to show deeper muscles.

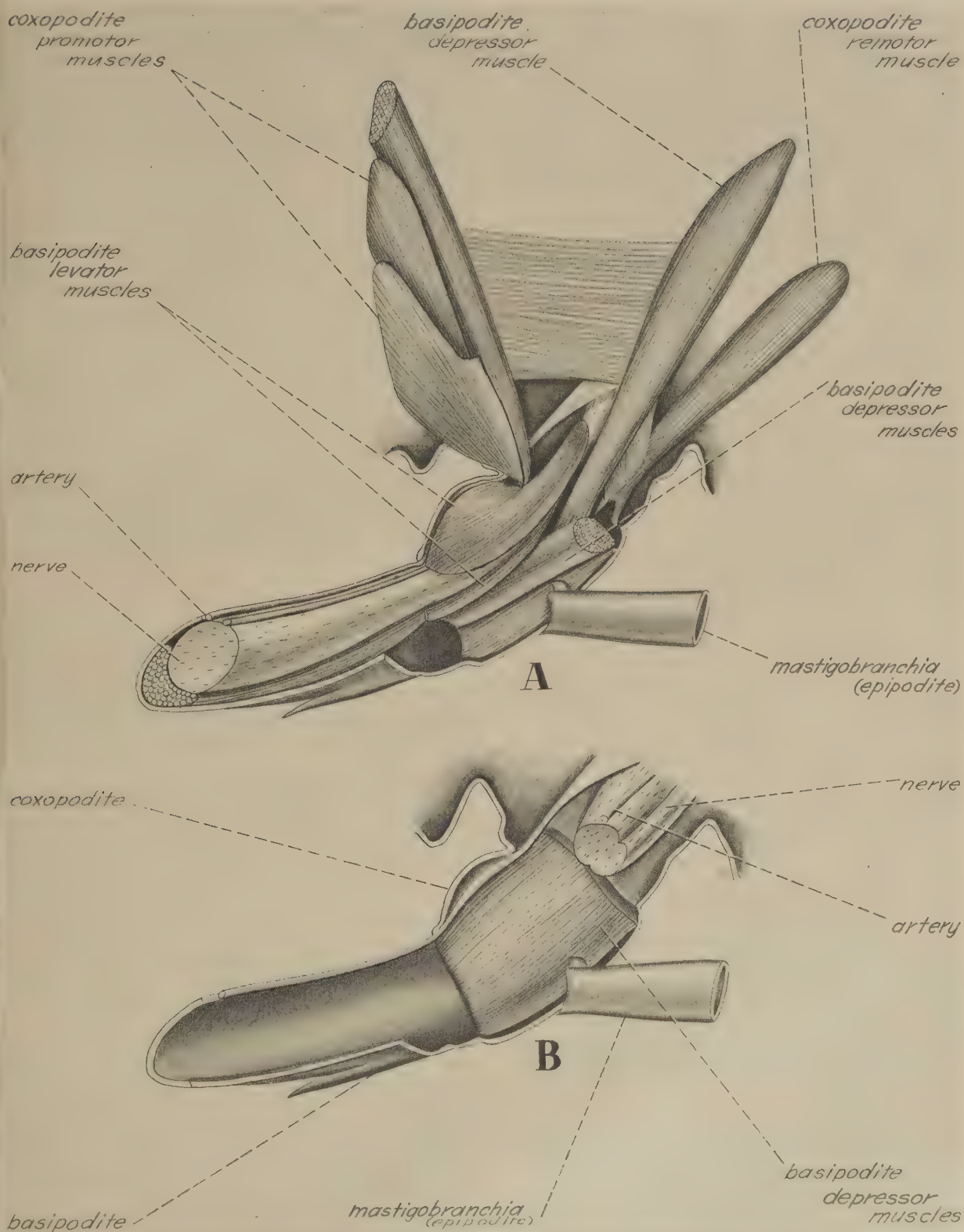


FIGURE 51.—Lateral view of leg base of left first pereopod. A. Lateral and medial muscles. B. Medial muscles.

The lateralmost levator (fig. 50, *A*) is a short, strong muscle taking origin on the lateral edge of the coxopodite and inserting on an anterolateral apodeme of the basipodite. Mesad of this levator lies a long, curving basipodite levator (fig. 50, *A*, *B*) which originates on the pleural plate and joins the former levator on the basipodite levator apodeme. The third basipodite levator is a short, rounded muscle situated mesad of the previously mentioned levators (fig. 50, *A*). The third levator originates in the coxopodite and inserts on the anterior rim of the basipodite. The medial, fourth levator, also originating on the coxopodite, inserts on the anteromedial edge of the basipodite (fig. 51, *A*). The basipodite levator muscles turn the basipodite and the distal leg article dorsad.

While the three walking leg levator muscles found in *Astacus* and *Callinectes* are undoubtedly homologous with some or all of the basipodite levators in *Penaeus*, the details of their relationships are uncertain.

BASIPODITE DEPRESSOR MUSCLES OF FIRST PEREIOPOD

FIGURES 49, 50, 51

The first walking leg of *Penaeus* has four basipodite depressor muscles. Two of the depressors originate in the coxopodite and two on the pleural wall. The lateralmost member of the group originates on the lateral side of the coxopodite (fig. 51, *A*) and passes ventromedially to insert on the common basipodite depressor apodeme. The longer, second depressor originates dorsad, on the pleural wall, and runs ventrad to the common apodeme. Mesad of the second basipodite depressor lies a short third depressor of the basipodite which is attached between the dorsomedial rim of the coxopodite and the basipodite apodeme. The fourth basipodite depressor muscle lines the medial side of the coxopodite. Unlike the other depressors, the fourth basipodite depressor inserts for some length along the posteromedial margin of the basipodite.

The basipodite depressor muscles turn the basipodite ventrad, thus accomplishing body support on the limbs. Considerable variation in basipodite musculature exists. The depressor muscle *a* of *Astacus* is very likely the homolog of the long, second basipodite depressor of *Penaeus*, whereas the depressor muscle *b* of *Astacus* is probably homologous to the lateral, first basipodite depressor muscle in *Penaeus*.

The relationships in *Penaeus* to the seven depressor muscles of the blue crab are difficult to ascertain. The depressor muscle *b* in the third pereopod of *Pandalus* looks much like the medial, fourth basipodite depressor muscle in the first pereopod of *Penaeus*.

EXOPODITE MUSCLE OF FIRST PEREIOPOD

FIGURE 50

The little, spindle-shaped exopodite muscle originates on the wall of the basipodite, mesad, and passes across the basipodite to a point of insertion at the base of the exopodite. Presumably its contractions move the exopodite, which, while very reduced, is connected to the basipodite by an articular joint. Nothing similar to the exopodite muscle is described in the first pereopod of the other crustaceans referred to. Berkeley describes an attractor of the mastigobranch in the pereopods of *Pandalus*, but illustrates the muscle as a coxopodite component.

ISCHIOPODITE REDUCTOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

The ischiopodite reductor muscle originates over an extensive area on the dorsomedial surface of the basipodite. The muscle inserts on a proximal ischiopodite apodeme located on the ventromedial rim of the ischiopodite. The reductor pulls the ischiopodite and with it the distal articles ventrad. The ischiopodite reductor muscle of *Astacus* is the same as that of *Penaeus*. The muscle is missing in the blue crab, in which the basipodite and ischiopodite are fused.

MEROPODITE FLEXOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

The meropodite flexor muscle is a two-part structure whose short fibers insert at an angle on an elongate apodeme projecting from the ventromedial edge of the meropodite, proximally. The apodeme divides the muscle approximately in half on its long axis. The muscle fibers originate about a wide area of the ventral and medial surface of the ischiopodite. Their contractions turn the meropodite mesad and to a lesser extent ventrally. The meropodite flexor muscle apparently is subdivided into two parts in *Astacus*, in which

Schmidt describes them as reducers. A single meropodite reductor is shown in the study of *Callinectes*. The muscle is unopposed by an extensor or productor in *Penaeus* and the other forms considered here.

CARPOPODITE EXTENSOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

The fibers of the carpopodite extensor muscle take origin over very nearly the whole of the lateral side of the meropodite. The apodeme upon which they insert extends proximally from the lateral edge of the carpopodite and runs almost the length of the meropodite. The carpopodite extensor straightens the carpopodite on the meropodite in a horizontal plane. The muscle is represented in *Astacus* and *Callinectes* as the carpopodite abductor muscle.

CARPOPODITE FLEXOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

Somewhat like the carpopodite extensor, the carpopodite flexor muscle is a long muscle inserting on a long apodeme. The muscle origins occur about the medial and ventral surfaces of the meropodite. The carpopodite flexor apodeme arises from the proximal margin of the carpopodite, mesad of the dicondylic axis. A pull on this apodeme turns the carpopodite and chela deeply on the meropodite, for this joint is a free one. The same muscle is found in the first pereopod of *Astacus* and *Callinectes*, but named the carpopodite adductor muscle.

CHELA PRODUCTOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

The fibers of the chela productor muscle arise from lateral, proximal, and medial regions of the dorsal part of the carpopodite and insert on the long productor apodeme of the propodite. Since the joint between the carpopodite and propodite is a free one, tension on the productor apodeme tends to straighten the chela with respect to the carpopodite and even to levate the chela. The propodite productor muscle of *Penaeus* is found in much the same form in *Astacus* and *Callinectes*, under the name of propodite productor muscle.

CHELA REDUCTOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

Directly ventrad of the chela productor muscle lies the chela reductor muscle, a structure also attaching by several slips to a long apodeme. The reductor apodeme arises from the ventral rim of the propodite. The chela reductor muscle turns the chela ventrad. The same muscle appears in the first pereopod of *Astacus* and *Callinectes*, and in the second pereopod of *Pandalus*, as the propodite reductor muscle.

CHELA EXTENSOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

As mentioned above, the joint between the carpopodite and propodite affords free movements of the latter on the former. The chela extensor muscle straightens the chela with respect to the carpopodite in the horizontal plane. The muscle originates in the distolateral part of the carpopodite and inserts on an apodeme projecting proximally from the lateral margin of the propodite. Exact counterparts of the chela extensor are missing in the first pereopod of *Astacus* and *Callinectes*. Berkeley describes, however, 2 extensors and 2 flexors of the propodite in the second pereopod of *Pandalus*. Homologies of these muscles without information about their innervations is not feasible.

DACTYLOPODITE ADDUCTOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

The dactylopodite adductor muscle originates throughout the medial part of the pod-shaped propodite and inserts on a heavy apodeme of the dactylopodite. Contractions of this large muscle close the dactylopodite on the distal gnathal process of the propodite. The dactylopodite adductor muscle is found in the first pereopod of *Astacus* and *Callinectes*.

DACTYLOPODITE ABDUCTOR MUSCLE OF FIRST PEREIOPOD

FIGURE 49

Functioning in opposition to the dactylopodite adductor muscle, the fibers of the dactylopodite abductor muscle originate about the lateral surfaces of the propodite and insert on the abductor

apodeme attached to the lateral margin of the dactylopodite. The muscle opens the jaws of the chela. The dactylopodite abductor muscle of *Penaeus* is fully homologous with the muscle of the same name in the first pereopod of *Astacus* and *Callinectes*. The first pereopod chela of *Pandalus* is reduced.

5b. Fifth Pereiopod

The fifth pereopod has been chosen as an example of a nonchelate limb. It is truly ambulatory, slender and very long, much longer than the first pereopod, mainly due to pronounced lengthening of the meropodite and carpopodite. Dorsad of the fifth pereopod projects a single pleurobranchia.

SKELETAL ELEMENTS

The coxopodite (fig. 54) is a rounded box whose lateral corner projects dorsally to a point. This point makes contact with the dorsal condyle of the article. The ventral condyle is located on the sternal plate between the limbs. The arrangement of these coxopodite dicondyles permits this article great freedom of movement. A large region of thin cuticle, the articular cuticula (fig. 54), lying caudad of the coxopodite, presents little resistance to posterior movements of the segment. A second articular cuticula associated with basipodite movements is found on the lateral surface of the coxopodite.

The strong basipodite (fig. 54) is hinged to the ventral surface of the coxopodite in the typical way, allowing for extensive movements of this article and of the distal segments in the vertical plane. With the dorsolateral surface of the basipodite is articulated a small exopodite. The ischiopodite (fig. 52) makes an oblique connection with the basipodite. The joint is slightly movable. The ischiopodite may be bent ventrad on horizontally located condyles. The articulation point between the short ischiopodite and the long meropodite is a transverse one capable of limited motion. By means of the joint the meropodite may be turned ventrad.

The carpopodite articulates with the meropodite by means of a complex "knee" joint made up of a pair of heavy condyles so arranged that the joint has considerable freedom of action. Similarly oriented, but not so freely movable, is the articulation between the carpopodite and propo-

dite. The propodite is not so long as the carpopodite. The slender, tapering dactylopodite makes contact with the propodite by a simple dicondylic joint. Chemoreceptor tufts and other sensory and mechanical setae are arranged in rows on the dactylopodite and to some extent on the propodite.

MUSCLE ELEMENTS

The arrangement of the muscles of the fifth pereopod is typical of the true supporting limbs of the thorax. The ambulatory muscles are light by comparison with those of the heavy reptant crustaceans and the terrestrial arthropods. In response to the support function of this appendage, the depressor musculature of the basipodite is especially well developed. The only anatomical account of the fifth pereopod of a crustacean known to the present writer is that of Cochran on *Callinectes*. However, the fifth leg of the blue crab is the swimming leg and hence its muscles, particularly the basal ones, have diverged from the typical pattern. A remarkable uniformity of endopodite musculature between *Penaeus* and *Callinectes* still remains, however. The fifth pereopod of *Penaeus* is operated by 24 muscles grouped into 13 functional types.

COXOPODITE PROMOTOR MUSCLES OF FIFTH PEROPOD

FIGURES 52, 53, 55 to 57

Two coxopodite promotor muscles are found in the fifth pereopod of *Penaeus*. The smaller lateral promotor originates by its broad, dorsal, fan-shaped portion on the laterotergal plate of the segment (figs. 55, 56) and inserts on the anterior rim of the coxopodite. Lying medial to the latter muscle is a larger promotor muscle mass originating somewhat dorsad of the smaller promotor on the pleural region. Contractions of these muscles turn the coxopodite forward on its condyles.

COXOPODITE REMOTOR MUSCLES OF FIFTH PEROPOD

FIGURES 52, 53, 55, 56

The fifth pereopod of *Penaeus* has two coxopodite remotor muscles. These are large, flat muscles (fig. 55) situated beneath the thin material of the articular cuticula (fig. 54). Both originate by broad margins on the pleural plate and insert on the caudal margin of the coxopodite.

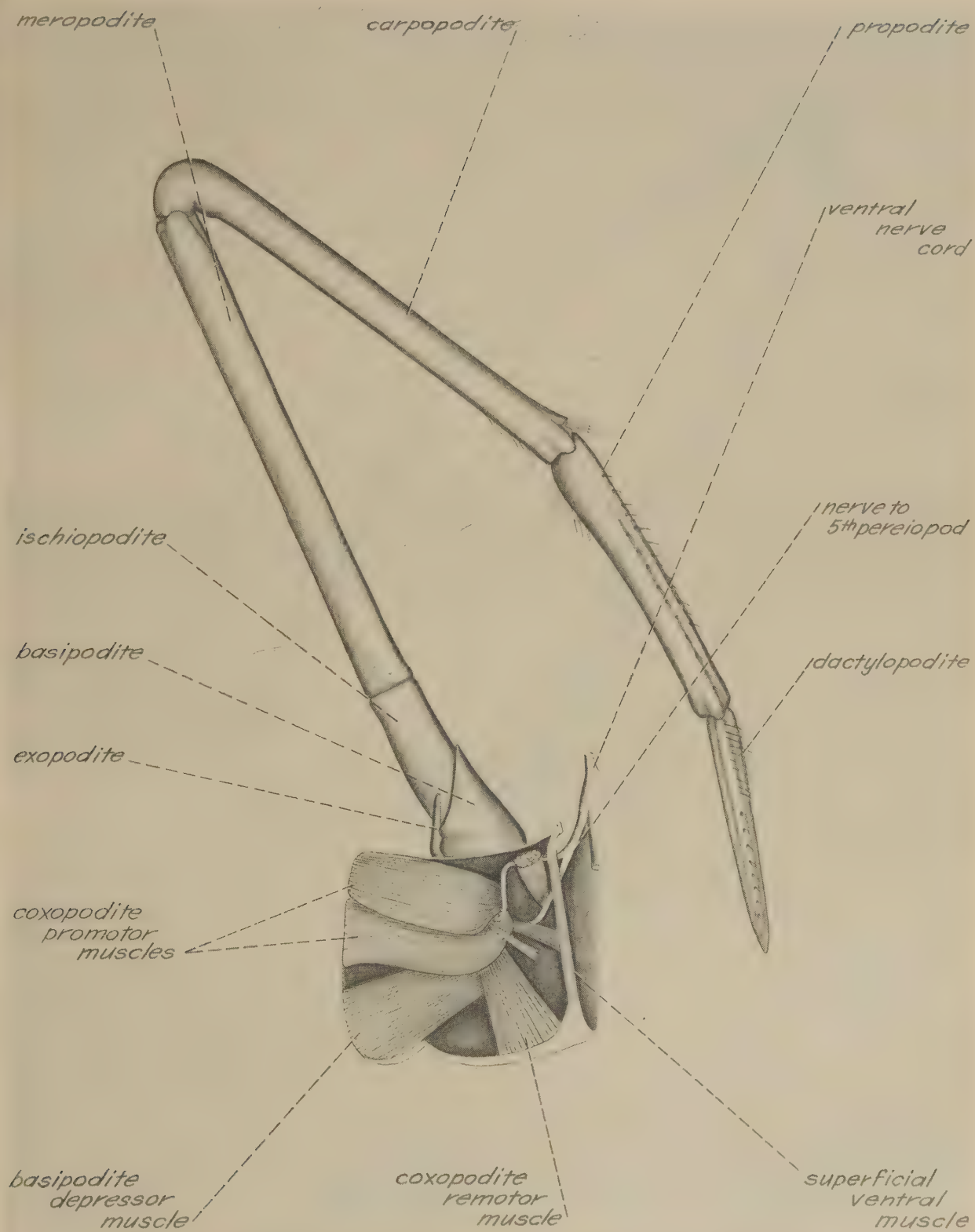


FIGURE 52.—Dorsal view of left fifth pereiopod showing protopodite muscles.

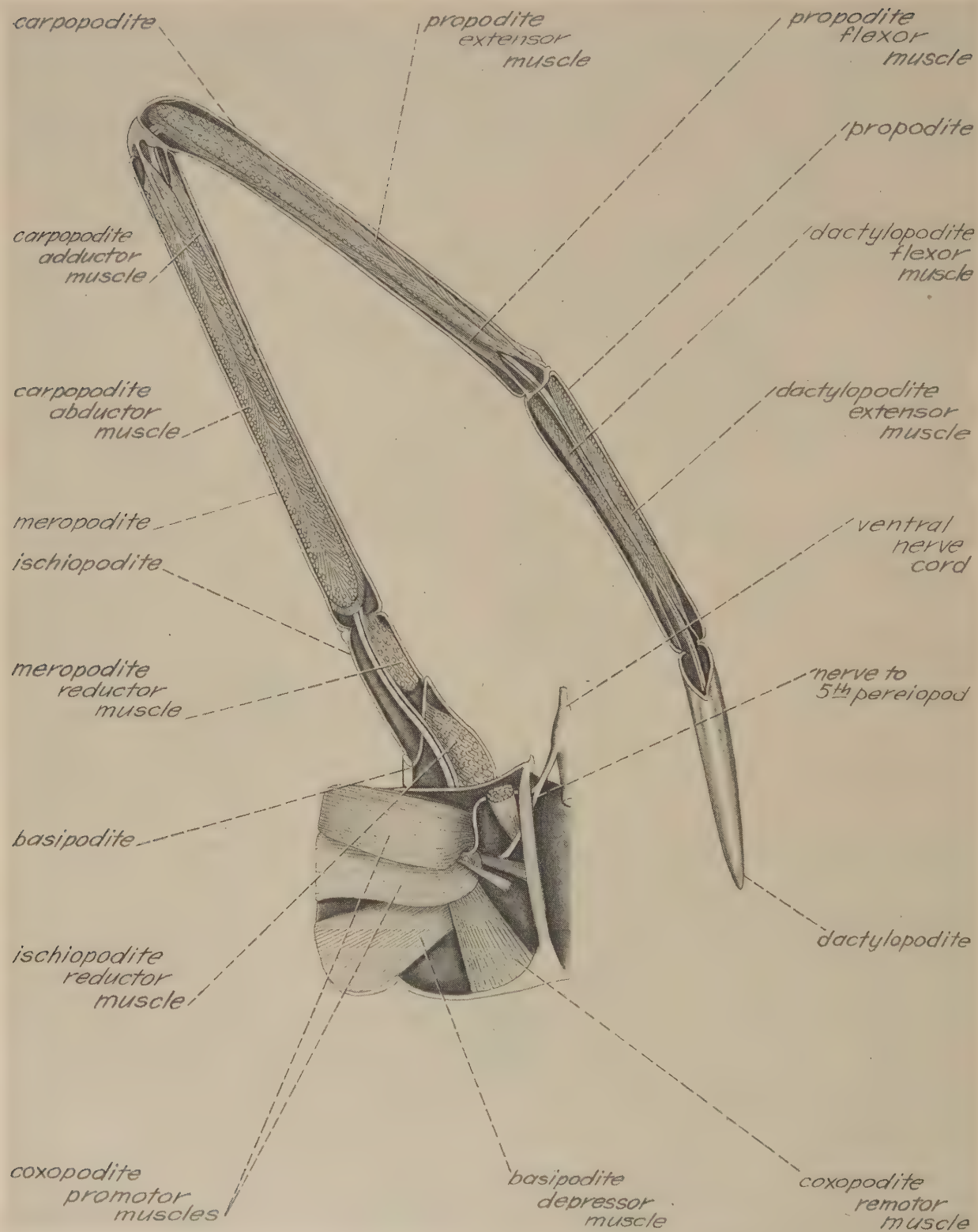


FIGURE 53.—Dorsal view of left fifth pereiopod. Dorsal cuticle removed to show muscles of propodite and endopodite.

The coxopodite remotor muscles act with some power to turn the coxopodite rearward.

COXOPODITE DEPRESSOR MUSCLE OF FIFTH PEREIOPOD

FIGURE 58

The coxopodite depressor muscle arises from phragmal material on the medial margin of the ventral skeletal foramen entering the coxopodite and runs to the medial margin of the coxopodite (fig. 58). Apparently the muscle is able to lift slightly the medial margin of the coxopodite, and for this reason the structure has been named the coxopodite depressor muscle.

BASIPODITE LEVATOR MUSCLES OF FIFTH PEREIOPOD

FIGURES 55, 56

Four basipodite levator muscles are seen in the fifth pereopod of *Penaeus*. The two lateral levators constitute together a broad fan (fig. 55) originating along the caudal margin of the coxopodite. They become narrow as they run to their insertions on the lateral rim of the basipodite. Two longer levators lying mesad of the lateral levators originate in the dorsal apex of the coxopodite and run to heavy apodemal material common to the levator muscles. The basipodite levator muscles raise dorsally the basipodite and with it the distal elements of the limb.

BASIPODITE DEPRESSOR MUSCLES OF FIFTH PEREIOPOD

FIGURES 52, 53, 55 TO 58

The most important muscles of the fifth pereopod of *Penaeus* are the depressors of the basipodite. Seven basipodite depressor muscles exist in the limb. The first and second depressor muscles are lateral (fig. 56). They take origins on areas of the caudal margin of the coxopodite and insert on the large common depressor apodeme of the basipodite. The third depressor (figs. 55, 56) is a large fan which originates broadly on the laterotergal plate of the body segment. The fourth basipodite depressor (fig. 57), also fan-shaped but narrower than the third, lies just mesad of the third depressor muscle. It, too, has a broad origin on the pleural area. The fifth depressor is a rather small muscle (fig. 58) which takes its origin on medial phragmal material of the ven-

tral skeleton. All of the foregoing five muscles insert on the basipodite depressor apodeme. The sixth and seventh depressor muscles (fig. 58) arise on phragmal material on the medial side of the ventral skeletal foramen and insert for some length along the posteromedial rim of the basipodite. The function of the basipodite depressor muscles is to turn the fifth pereopod ventrad, providing support for the body.

ISCHIOPODITE REDUCTOR MUSCLE OF FIFTH PEREIOPOD

FIGURE 53

The ischiopodite reductor muscle has multiple origins over a substantial area on the dorsal and medial parts of the basipodite. It inserts on an apodeme projecting from the ventral surface of the ischiopodite. The muscle bends the ischiopodite ventrad slightly and to some extent rotates it, due to the oblique angle by which the basipodite and ischiopodite are connected (fig. 52). The muscle is not opposed by any other muscle.

MEROPODITE REDUCTOR MUSCLE OF FIFTH PEREIOPOD

FIGURE 53

Arising by multiple origins on the dorsal and medial half of the ischiopodite, the meropodite reductor muscle fibers insert on a small apodeme on the ventral surface of the long meropodite, proximally. The meropodite reductor bends the meropodite ventrally a short distance.

CARPOPODITE ABDUCTOR MUSCLE OF FIFTH PEREIOPOD

FIGURE 53

The fibers of the carpopodite abductor muscle originate over most of the dorsal half of the meropodite and insert on an extremely long apodeme running nearly the whole length of the meropodite along the midline. The length of pull of this muscle and of its opponent, the carpopodite adductor, is very great. The long apodeme to which it attaches arises from the proximal end of the carpopodite, lateral to the condylic axis. The muscle thus turns the carpopodite to a position in which the axes of the meropodite and carpopodite are in line. The carpopodite abductor muscle easily could be described as an extensor.

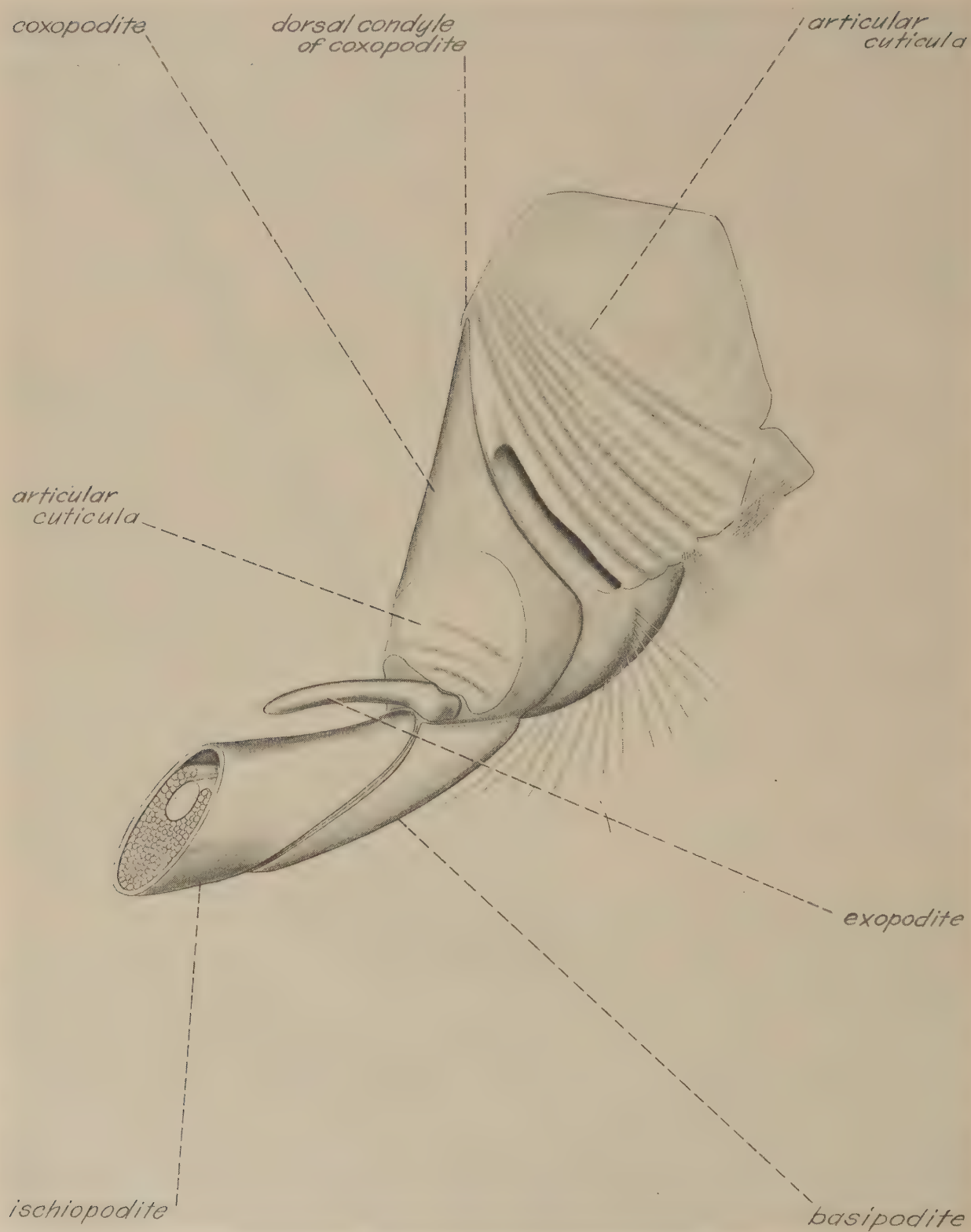


FIGURE 54.—Lateral view of leg base of left fifth pereopod.

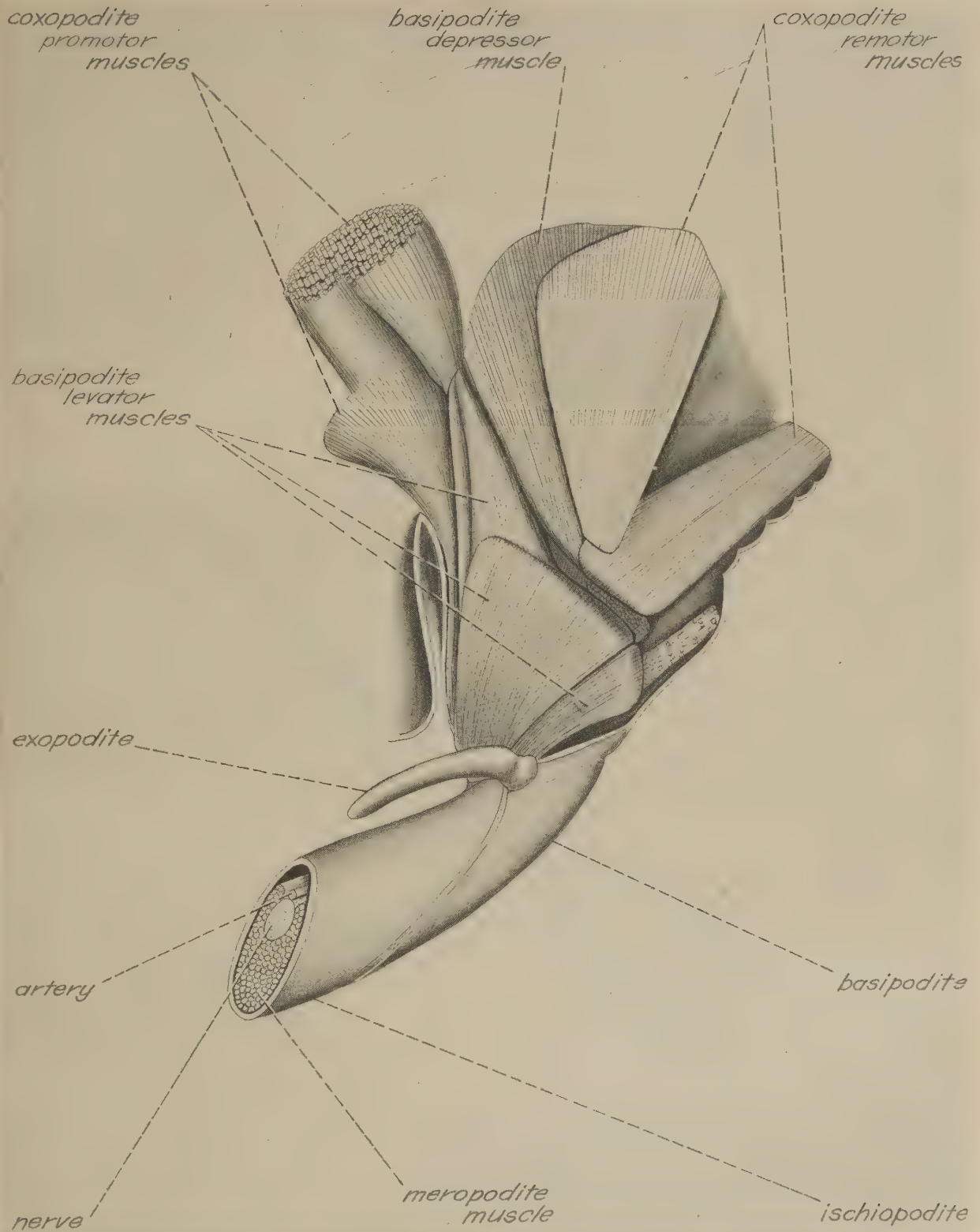


FIGURE 55.—Lateral view of leg base of left fifth pereopod. Lateral cuticle removed to show superficial lateral muscles.

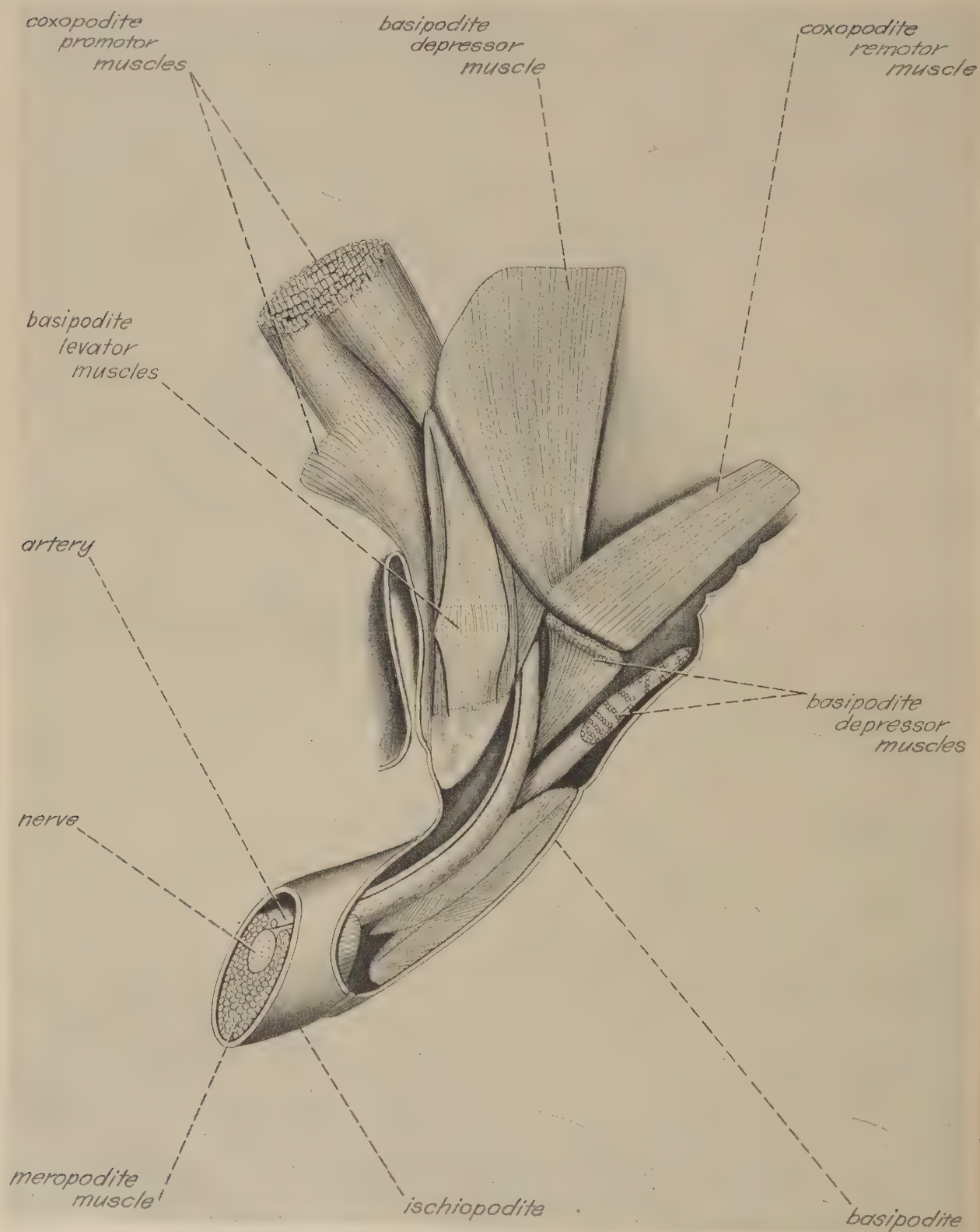


FIGURE 56.—Lateral view of leg base of left fifth pereopod. Some superficial lateral muscles removed.

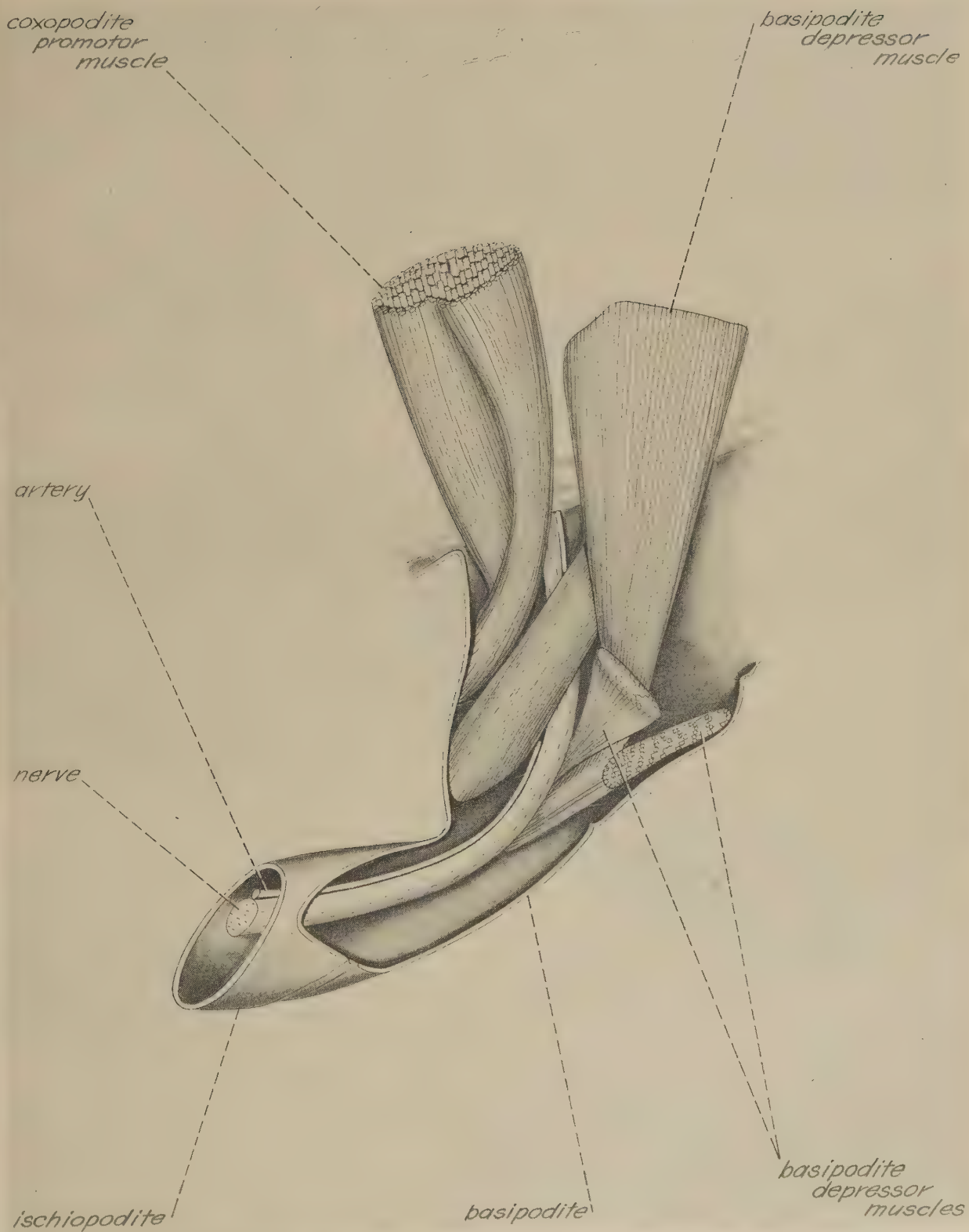


FIGURE 57.—Lateral view of leg base of left fifth pereopod. Lateral muscles removed to show medial muscles.

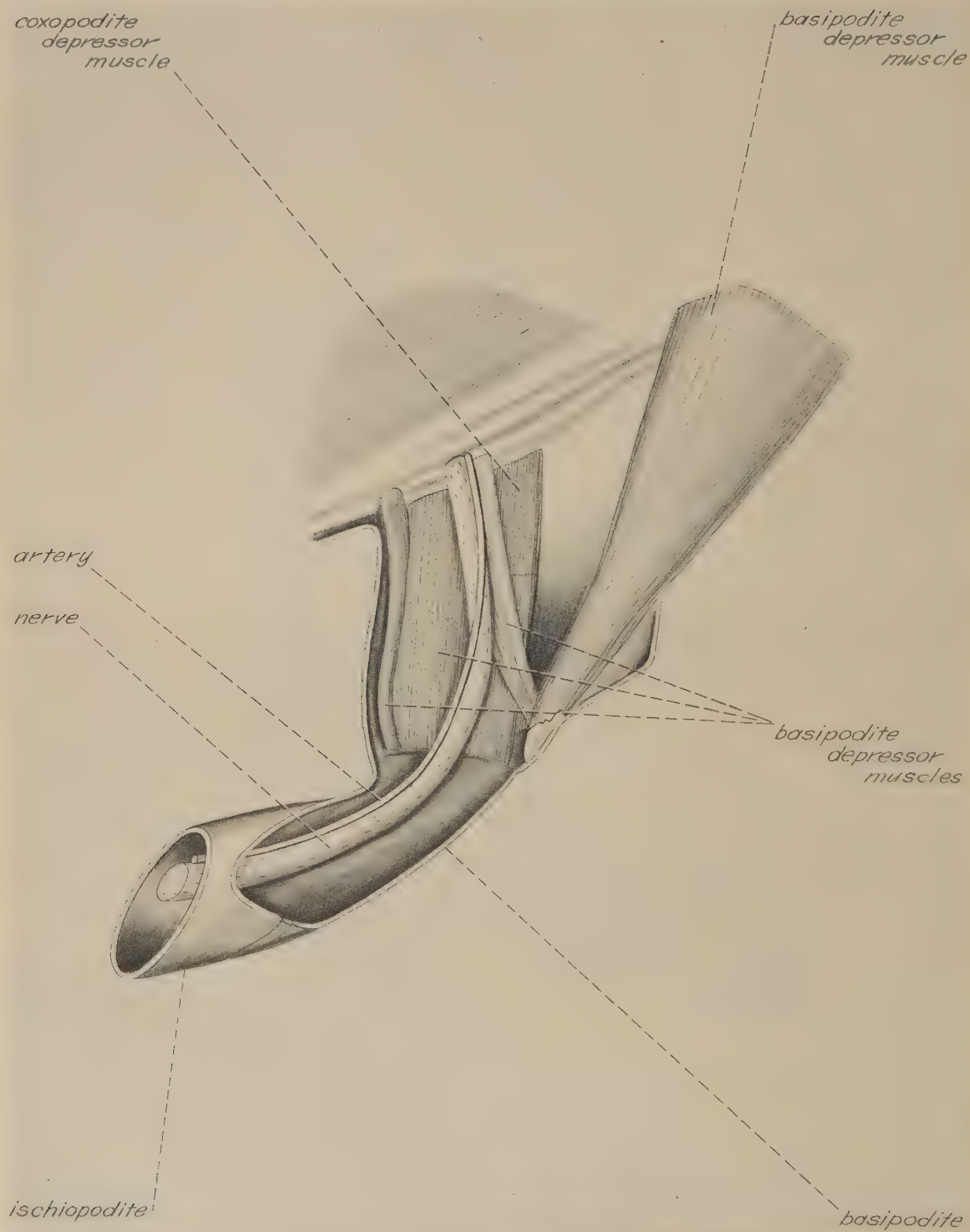


FIGURE 58.—Lateral view of leg base of left fifth pereopod. Medial muscles.

CARPOPODITE ADDUCTOR MUSCLE OF
FIFTH PEREIOPOD

FIGURE 53

In like fashion to the carpopodite abductor, the carpopodite adductor muscle fibers originate on a large area of the meropodite, but on the ventral half of the article. The muscle inserts on a long apodeme arising from the proximal portion of the carpopodite, medial to the axis of the dicondyles. Carpopodite adductor muscle contractions serve to turn the carpopodite toward the body, and in fact deeply on the meropodite. The muscle might better be considered a flexor of the carpopodite.

PROPODITE EXTENSOR MUSCLE OF
FIFTH PEREIOPOD

FIGURE 53

The arrangement of the propodite muscles in the carpopodite is very similar to that of the carpopodite muscles within the meropodite. The propodite extensor muscle originates over much of the dorsal part of the carpopodite and inserts on a long apodeme projecting proximally from the base of the propodite. The apodeme is so connected to the propodite that a pull on it extends the propodite with respect to the carpopodite.

PROPODITE FLEXOR MUSCLE OF
FIFTH PEREIOPOD

FIGURE 53

The propodite flexor muscle opposes the action of the propodite extensor muscle. The fibers of the flexor muscle arise from the ventral surface of the carpopodite and, like those of the extensor, attach to a long apodeme of the propodite. This apodeme arises from a position opposite to that of the extensor apodeme. Contractions of the propodite flexor muscle flex the propodite upon the carpopodite.

DACTYLOPODITE EXTENSOR MUSCLE OF FIFTH
PEREIOPOD

FIGURE 53

The dactylopodite extensor muscle originates along the lateral side of the propodite and inserts on a long apodeme arising from the proximal end of the dactylopodite. The muscle straightens the dactylopodite on the propodite.

DACTYLOPODITE FLEXOR MUSCLE OF FIFTH
PEREIOPOD

FIGURE 53

The dactylopodite flexor muscle bends the dactylopodite upon the propodite. Like the extensor muscle, the flexor has multiple origins upon the medial surface of the propodite. The muscle inserts on the long flexor apodeme of the dactylopodite.

C. Abdomen

Unlike the head and gnathothorax, the abdomen is almost entirely devoted to the propulsion of the white shrimp. Except for slender components of the gut, the gonads, and the nervous and circulatory systems, the space within the abdominal skeleton is filled with muscles, most of them concerned with the powerful flexion of which the animal is capable. The abdomen consists of six segments, all of which bear appendages, and a posterior telson which does not. The abdominal segments are attached to one another by deep folds of thin articular cuticle which allow each segment great freedom of movement with respect to its neighbors and with the thorax. Intersegmental connections in the abdomen are of several types. The simplest and perhaps most movable is that between the thorax and first abdominal segment (fig. 59). Here, cuticular folds of great depth reinforced by heavy muscles internally make ventral flexion possible between these body tagmata. The junction is without special, restrictive condyles, allowing extensive lateral movements of the abdomen on the thorax at this point.

The junctions of the first and second, and of the second with the third abdominal segments (fig. 59) are identical. Motion at these joints is limited to flexion and extension by lateral condyles of similar design. In contrast, the joint between the third and fourth abdominal segment differs from those between the first three segments. This connection is much simpler. The condyles are rather loosely connected, affording flexion and extension and a certain amount of lateral motion. The fourth, fifth, and sixth abdominal segments articulate with one another by means of two pairs of identical condyles similar in their rigidity to those between the first, second, and third segments, but different in structure (see enlargements of condylic structure, detail of articulation, fig. 59).

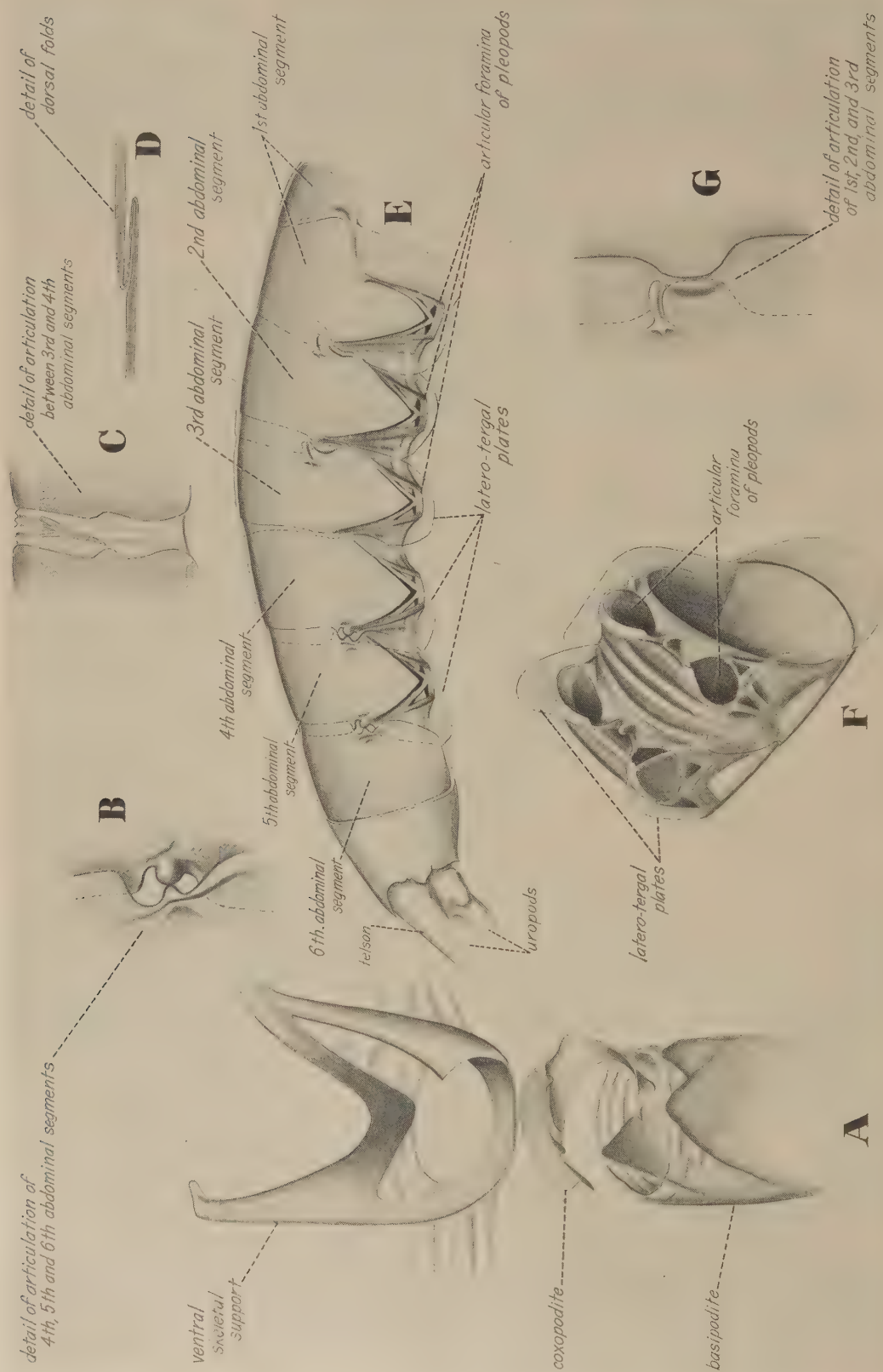


FIGURE 59.—Details of abdominal skeletal elements.

The posterior joints permit free extension and flexion, but probably resist lateral motion. We have, then, 4 kinds of intersegmental hinges functioning in 2 different ways. No reason for this multiplicity of structure in the hinges is readily apparent.

SKELETAL ELEMENTS

The abdominal segment of arthropods is comprised of a ring of cuticle. This ring, however, may be subdivided in parts. The abdominal somite in the Crustacea has usually been said to consist of an arched dorsal tergum and a flat, concave, or convex ventral sternum connected to the tergum by two lateral pleura, the latter often produced ventrad in a fold. The lateral pleuron has been variously interpreted. Snodgrass (1935) describes the pleural areas of arthropods as "typically membranous" to permit the movement of appendages arising from the pleuron. Pleural sclerites, when present, represent a contribution of the proximal parts of the appendages, according to this worker. He sets forth the fact also that the appendages articulate with the ventral margins of the pleural sclerites and with the lateral edges of the sternal sclerites. That this typical situation may not always obtain is shown by Hart (1952) who found that the first pleopod in *Cambarus longulus longulus* Girard, 1852, is connected to the abdominal venter entirely by sternal components.

Snodgrass (1935) apparently was of the opinion that the arthropod pleuron represented a distinct region of the segment. In a study of the Crustacea made at a later date, Snodgrass (1952) describes in *Anaspides tasmaniae* Thomson, 1892, a generalized malacostracan, a clearly demarked pleural sclerite with which the limb basis articulates. In the same study he refers to the pleural sclerite as a "laterotergal pleural plate" and states that the pleuron of the crustacean belongs to the tergum. Support of the more recent opinion of Snodgrass (1952) that the pleuron of crustaceans is tergal in origin may be found by a consideration of the abdominal segment of *Penaeus setiferus*.

The sternum and tergum are distinct regions in the white shrimp, but no line or suture can be seen which distinguishes a pleural component of the abdominal segment (fig. 59). From this we may conclude that the pleura are not needed in a morphological construct of the abdominal segment of the crustacean. That the term, "pleuron,"

for the laterotergal plates has become embedded ineradicably in the literature of crustacean systematics is therefore most regrettable.

Beyond what has been brought out about the general structure and articulation of the abdominal segments, little need be said further except for a brief mention of the modified posterior end of the sixth abdominal segment. Unlike the five anterior abdominal segments from which light appendages project, the sixth abdominal segment must be strong enough to bear the tail fan. For this reason the posterior end of the sixth segment is heavily sclerotized. In addition, the contents of the sixth segment are largely devoted to mediating the flexions of the tail fan which project caudad from the segment rather than ventrad as is the case with the preceding abdominal segments. The result is the visible difference in shape of the sixth segment compared to that of the typical abdominal segment. More will be said about the sixth abdominal segment when the tail fan is considered.

MUSCLE ELEMENTS

The characteristic of the abdominal muscles, apart from their great mass, is the unusual manner in which they are laid out. These muscle bodies are extraordinarily heavy and so intertwined with one another that their separation for study is difficult. In accordance with the system of Daniel (1931c), the abdominal muscles are divided into the following four functional muscle groups: (1) The superficial ventral muscles, (2) the lateral muscles, (3) the dorsal muscles, and (4) the main ventral muscles. They will be treated in that order.

SUPERFICIAL VENTRAL ABDOMINAL MUSCLES

FIGURES 61, 63

The superficial ventral abdominal muscles of *Penaeus* are attached between yokes of thin cuticle (fig. 63) lying transversely across the posterior portion of each abdominal segment beneath the ventral nerve cord. These yokes are associated with the folds of articular cuticle on the ventral surface, and they are so arranged that the superficial ventral muscles run from the posterior region of one abdominal segment to the posterior region of the next. The superficial ventral abdominal muscles are of course a continuous sys-

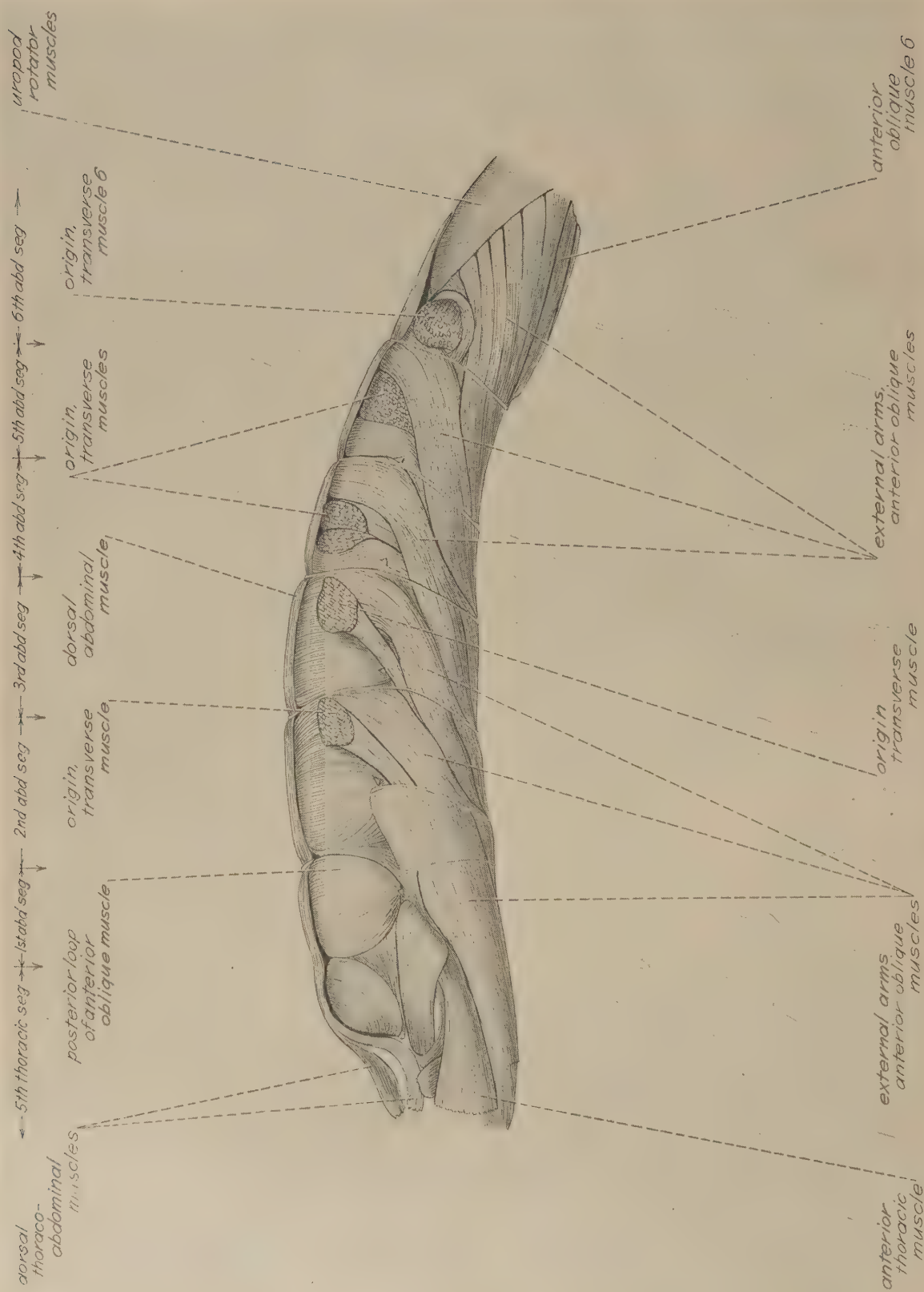


FIGURE 60.—Lateral view of left side of abdomen showing superficial abdominal muscles.

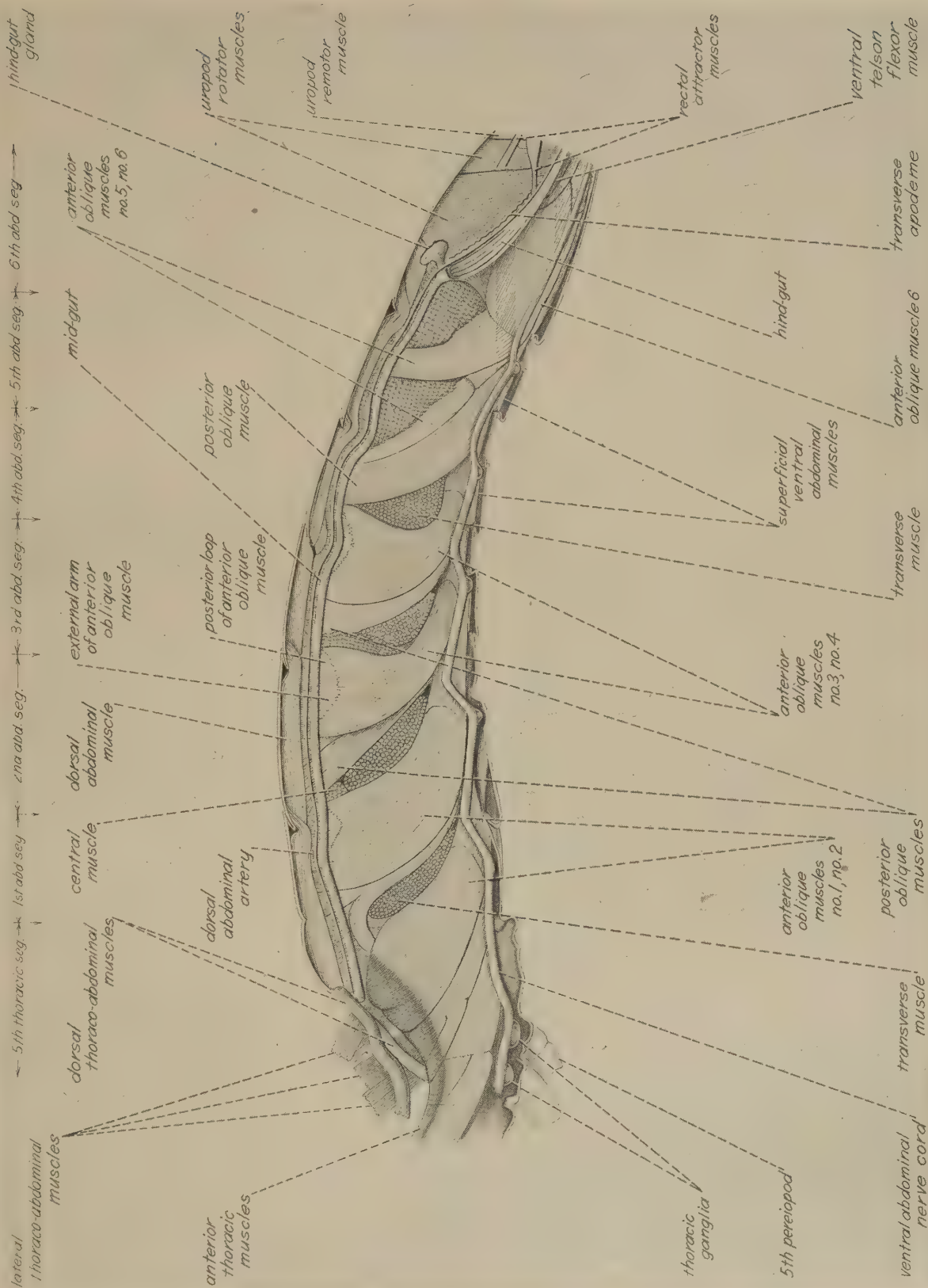


FIGURE 61.—Median sagittal cut through abdomen showing medial muscles.

tem with the superficial ventral thoracic and the superficial ventral thoracoabdominal muscles considered above in the section on the gnathothorax.

Like their thoracic counterparts, the superficial ventral abdominal muscles are very thin. Two of them are found on each side of the ventral nerve cord between the first and second and between the second and third abdominal segments. Only one is found on each side thereafter. Except for the ventral muscle that passes between the fifth and sixth abdominal segments, and which is ventrad of the nerve cord, all of them arise anterolaterally and run mesad. In this respect, the situation in *Pandalus* is much more like that of *Penaeus* than is *Astacus*. The superficial ventral muscles presumably function to hold the articular cuticle between the abdominal segments in place.

The superficial ventral muscles of *Pandalus*, *Astacus*, and *Callinectes* are probably homologous with the same muscles in *Penaeus*. Some differences exist. All of the superficial ventral muscles of the former crustaceans are single on a side, except for the last two abdominal segments of *Astacus* in which the muscles are double.

SUPERFICIAL LATERAL ABDOMINAL MUSCLES

Although these muscles are not illustrated here, their centers are indicated by small triangles on figure 60. The triangles represent apodemal depressions. Each superficial lateral abdominal muscle is attached to broad, tough apodemal material arising at the triangles. The muscles are very thin, fan-shaped structures, and apparently function to retain the position of the cuticle during movements of the white shrimp.

SUPERFICIAL DORSAL ABDOMINAL MUSCLES

FIGURE 62

Lateral to the midline on the dorsal surface of the abdomen lie the thin superficial dorsal abdominal muscles. The muscles arise in superficial connective tissue in the abdominal segments and pass to apodemal material at the anterior margins of the succeeding segments. Those inserting on the fourth, fifth, and sixth abdominal segments are square, flat muscles, situated laterad of the main dorsal abdominal muscles. The superficial dorsal abdominal muscle attached to the second abdominal somite is a long, thin muscle lying dorsad of the main dorsal muscles. Inserting on the third abdominal segment are two superficial dorsal ab-

dominal muscles. The medial one is long and slender, while the lateral muscle is rectangular and has a lateral anterior projection. The superficial dorsal muscles probably hold the articular cuticle in position.

The superficial dorsal abdominal muscles have counterparts in *Pandalus* and *Astacus*. Those of *Pandalus*, however, are all single muscles on each side of the midline. In addition Berkeley illustrates a sixth superficial dorsal muscle in *Pandalus* inserting on the telson. In *Astacus*, 6 pairs of strong superficial dorsal muscles are found lateral to the midline and a single seventh muscle attached to the telson.

MAIN DORSAL ABDOMINAL MUSCLES

FIGURES 60, 61, 62

The main dorsal abdominal muscles function as abdominal extensor muscles in opposition to the action of the huge ventral abdominal muscle mass. The dorsal abdominal muscles make apodemal connection with the dorsal thoracoabdominal muscles (fig. 61) and as such represent a functional abdominal continuation of the latter. The dorsal abdominal muscles may be divided into two groups, the dorsolateral abdominal muscles, and the dorsomedial abdominal muscles. Both groups are easily distinguished from the underlying ventral muscles. Their removal exposes the midgut and various dorsal circulatory and nervous elements. The arrangement of the dorsolateral and dorsomedial muscles of *Astacus* appears to be similar to that of *Penaeus*, in that the two groups of muscles are distinct. In *Pandalus*, on the other hand, these muscle groups are so intertwined that Berkeley refers to the lateral and medial parts as slips of the same muscle.

In *Penaeus* the dorsolateral and dorsomedial abdominal muscles are markedly segmental and readily separable from one another, except for the dorsal abdominal muscle of the second segment (occupying the dorsum of the first abdominal segment (fig. 62)). Here, the muscles are fused. The medial portion is connected with the dorsal thoracoabdominal muscle and a lateral slip of fibers is attached to superficial connective tissue. The whole structure passes caudad to insert on the dorsal apodemal yoke dividing the first and second somites. The dorsolateral muscles occupying the second, third, fourth, fifth, and sixth abdominal segments all arise from the cuticular yokes

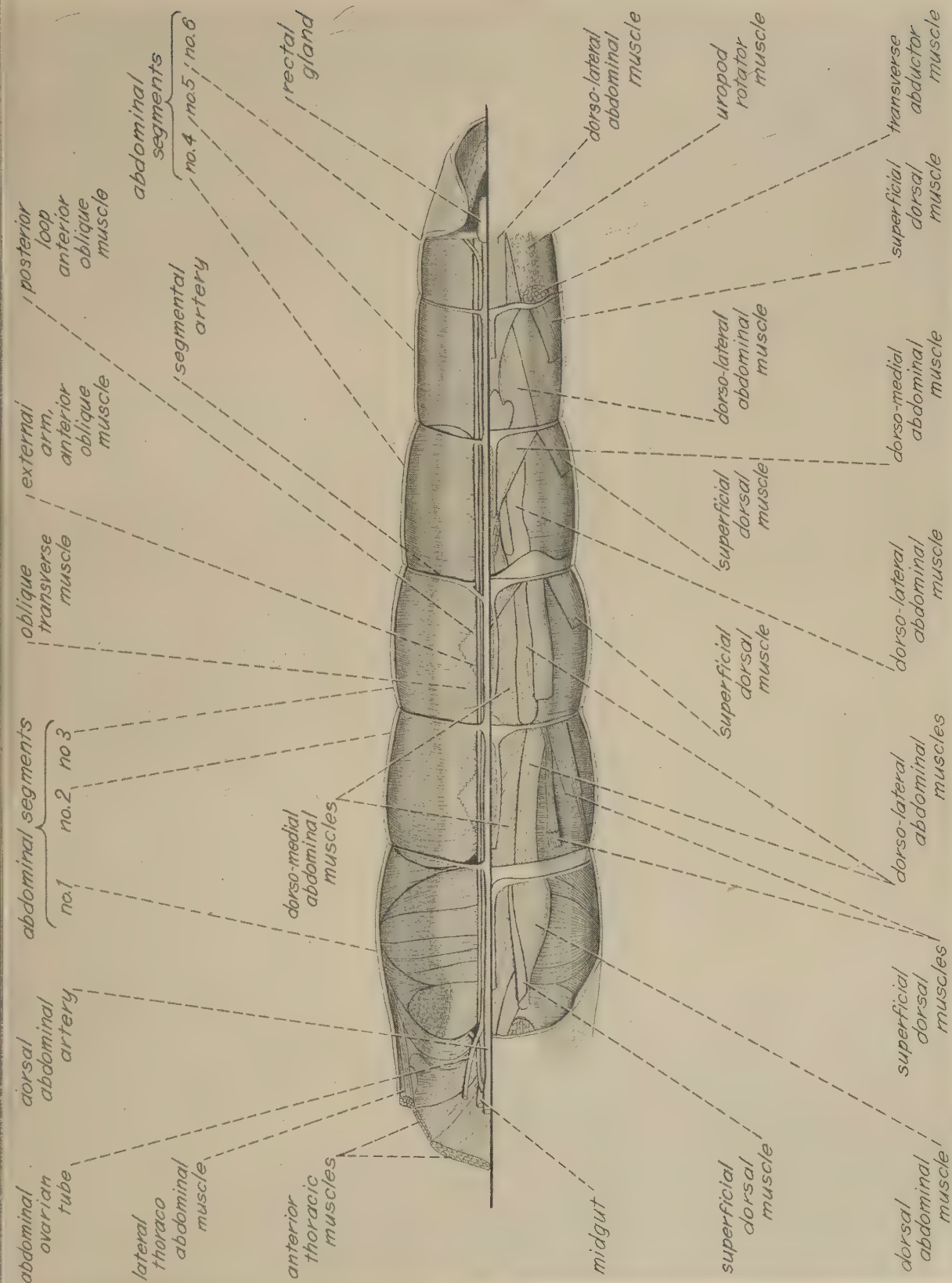


FIGURE 62.—Dorsal views of abdominal muscles. Lower section, intact musculature. Upper section, dorsal abdominal muscles removed.

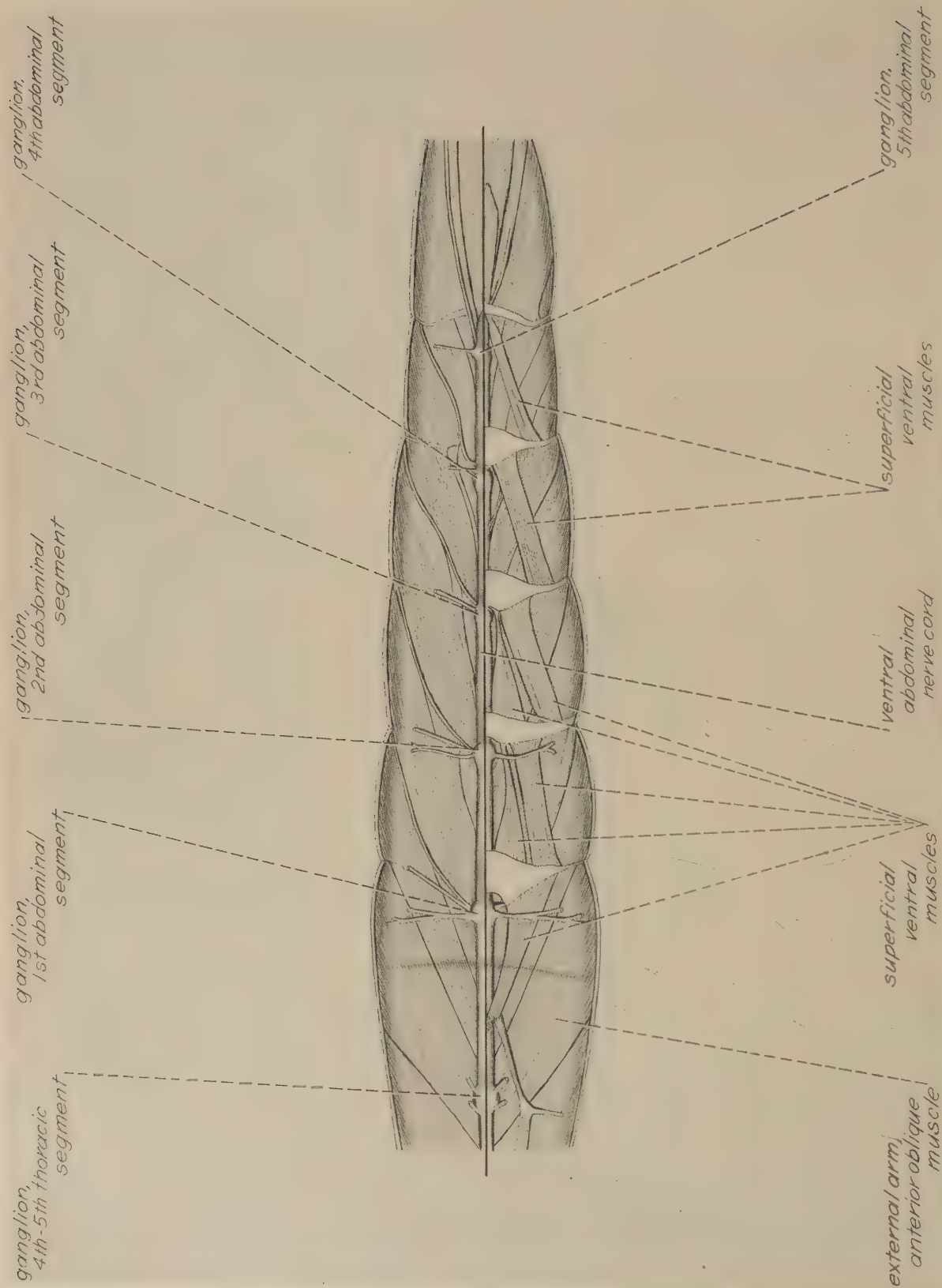


FIGURE 63.—Ventral views of abdominal muscles. Lower section, intact musculature. Upper section, superficial ventral muscles removed.

passing across the anterior part of each abdominal segment and from them run caudad to the next yoke. The axis of the fibers of the lateral muscles is approximately parallel with the long axis of the animal.

The fibers of the dorsomedial abdominal muscles are oblique to the long axis of the white shrimp. These muscles lie beneath the dorsolateral muscles. Each originates on a median sagittal apodeme within a segment and runs laterad to insert on an intersegmental apodeme, posteriorly. The arrangement in *Penaeus* is far simpler than that of the dorsal abdominal musculature of *Pandalus*.

Dorsad of the muscles under discussion, and very close to the middorsal line, are paired structures passing rostrad from the cuticular tissue dividing the abdominal segments (see lower half of fig. 62). The structures appear to be flattened tubes, soft and easily broken. They resemble vessels of the circulatory system more than anything else. Their function is unknown.

MAIN VENTRAL ABDOMINAL MUSCLES

The illustrations show that relatively few abdominal muscle types exist. However, the few that do are serially repeated and so extensively interwoven that their isolation is extremely difficult. Consequently no one should be surprised that an understanding of abdominal muscle function is of fairly recent derivation. Much of what we know is based upon the work of Daniel (1928, 1929, 1931a, 1931b, 1931c, 1931d, 1933) who first set forth the details of the comparative anatomy of abdominal muscles in higher Crustacea.

One of the most interesting features of abdominal anatomy is the contrast between the lightly sclerotized abdominal skeleton and the very heavy abdominal musculature. One would almost expect the light cuticle to be damaged by the powerful flexions of the abdomen. That such does not occur is explained by the unusual arrangement of the abdominal muscles. Some of them are attached to prevent extreme distortions that might otherwise take place. Some muscles, in a sense then, substitute for the skeleton. In addition, certain muscles act in the functional sense as great apodemes of other muscles, thereby freeing the abdominal skeleton of the requirement to produce all but the simplest apodemal material. These apodemal muscles, furthermore, act as fulcrum to

improve the mechanical advantage of long muscles, such as the anterior obliques, connected between two widely separated abdominal segments.

The terminology of the abdominal musculature of *Penaeus* has been adopted from the work of Daniel. For this reason, substantial differences in nomenclature will be found between the present study and the older morphological research. Daniel (1931c, 1932) has renamed some abdominal muscles and subdivided others. For the most part, however, he makes use of the classical abdominal muscle names.

CENTRAL MUSCLES OF ABDOMEN

FIGURE 61

The longitudinal central muscles are among the main fulcral muscles of the abdomen. They lie deep within the abdominal muscle mass. Six pairs of central muscles are found in the abdomen of *Penaeus*, joined together end to end in a wavy chain. Each central muscle takes origin on an apodeme of a dorsal branch of the preceding central muscle. These dorsal apodemes lie above the transverse muscles. As the central muscle passes caudad into the succeeding segment, it is first joined from above by the fibers of the oblique transverse muscle and then, in the neighborhood of the succeeding transverse muscle, the central muscle divides. The dorsal slip joins the apodeme of the next central muscle and the larger ventral part turns mesad along the anterior face of the transverse muscle. The central muscle apodemes dorsad of the transverse muscles also give rise to the posterior oblique muscles.

The counterparts of the central muscles of *Penaeus* in *Astacus* are not easy to determine. The only muscles in the European crawfish corresponding to the central muscles are the muscoli ventrales profundi of Schmidt. The central muscles function to support the oblique muscles during the contractions of the latter.

TRANSVERSE ABDOMINAL MUSCLES

FIGURES 60, 61, 64

The large transverse abdominal muscles attach dorsolaterally on the abdominal tergum (fig. 60) and run across the abdomen in association with strong fasciae (the fasciae arise at points indicated by small triangles in fig. 60). The abdomen of *Penaeus* contains six transverse muscles. A mid-sagittal view of the abdomen (fig. 61) shows the

transverse muscles crossing the midline. Their great size at the midline is partly due to the addition of fibers of the posterior loop of the anterior oblique muscle, as well as those of the ventral slip of the central muscle, mentioned above. The muscles function in the fulcral support of the central muscles, together with lateral compression of the abdomen. The transverse abdominal muscles of *Penaeus* are fully homologous with those of *Pandalus* and *Astacus*. Both of the latter forms have six of these muscles.

ANTERIOR OBLIQUE MUSCLES OF ABDOMEN

FIGURES 60 TO 64

The anterior oblique muscles are directly responsible for the strong abdominal flexions of which the white shrimp is capable, and as such are on functional grounds the most important longitudinal muscles of the abdomen. Each anterior oblique muscle is made up of several parts. The thickest part runs mesad of the central muscle and is closely applied to its opposite number at the median sagittal line (fig. 61). From the midline, the main part of the anterior oblique turns ventrally and curves caudad to an insertion area two segments to the rear. The area of its insertion is on the posteroventral margin of the abdominal segment concerned. *Penaeus* has 6 anterior oblique muscles, compared with 7 each for *Pandalus* and *Astacus*.

FIRST ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

The first anterior oblique muscle arises in connective tissue in the fifth or last thoracic segment, mesad of the lateral thoracoabdominal muscle (fig. 61) and runs ventrocaudally through part of the last thoracic and all of the abdominal segment to an area of insertion on the posterior edge of the first abdominal segment (fig. 64). The muscle functions to pull the ventral surface of the first abdominal segment forwards about its hinges with the fifth thoracic segment. The muscle appears in the abdomen of *Pandalus* and *Astacus*.

SECOND ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

This muscle (fig. 61) traverses the first and second abdominal segments, inserting on the posteroventral portion of the second segment (fig.

64). By the contractions of the second anterior oblique muscle the ventral part of the second abdominal segment is brought rostrad. Homologs of this muscle have been described in *Astacus* and *Pandalus*.

THIRD ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

Beginning in the second abdominal segment (fig. 61), the third anterior oblique muscle sweeps posteroventrad and inserts on the posteroventral rim of the third abdominal segment. It pulls the ventral part of the third segment forward. The same muscle is found in the abdomen of *Pandalus* and *Astacus*.

FOURTH ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

From its origin in the third abdominal segment, the fourth anterior oblique runs directly ventrad, then turns sharply caudad and passes along the ventral surface of the abdomen to its insertion on the fourth segment (fig. 61). The muscle in *Penaeus* is represented in *Astacus* and *Pandalus*.

FIFTH ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

This muscle runs in a gentle curve from its dorsal origin in the fourth abdominal segment to its insertion in the rear of the fifth segment (fig. 61). The medial manifestation of the fifth anterior oblique is much slenderer than is that of the preceding anterior oblique muscles. Contractions of the muscle flex the fifth segment ventrad with respect to the fourth segment. Schmidt and Berkeley describe the muscle in *Astacus* and *Pandalus*.

SIXTH ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

Like the fifth anterior oblique, the sixth curves gently from the dorsal part of the fifth abdominal segment to its special insertions at the caudal end of the long sixth segment (fig. 61). The ventral part of the sixth anterior oblique muscle lying in the sixth segment may be seen to be very thick. Actually, its cross-sectional area is no greater than the caudal portions of the other anterior oblique muscles, but by virtue of its concentration along the midline, a median sagittal view reveals a large portion of the muscle. The muscle is functionally similar to the preceding anterior oblique muscles. However, its insertions upon the sixth segment and the parts of the tail fan are

substantially modified from the typical plan of the preceding anterior oblique muscles.

At the posterior end of the last segment, the sixth anterior oblique muscle has three areas of insertion. Moving caudad from the anterior end of the sixth segment, certain fibers of the sixth anterior oblique muscle insert in tough connective tissue over a large area on the ventral surface of the sixth somite (figs. 71, 72). These fibers are obviously associated with the flexion of the sixth abdominal segment on the fifth. The major portion of the ventral fibers of the sixth anterior oblique continue caudad beyond the sternal insertion area to insert on a strong tendon, or apodeme of anterior oblique muscle (figs. 71, 72). This apodeme is firmly connected to the uropod protopodite, the base element from which the uropods arise. Thus the sixth anterior oblique flexes the uropods as well as the sixth segment.

The third area of sixth anterior oblique insertion is slightly dorsad of the ventral anterior oblique apodeme. It, too, is apodemal, and, while lighter than the ventral apodeme, is a strong tendon. This dorsal anterior oblique apodeme is best seen in median sagittal aspect (figs. 61; 74, *B*). In contrast to the ventral anterior oblique apodeme, the dorsal tendon bifurcates anteriorly into two parts. Two large and distinct portions of the sixth anterior oblique muscle insert on the tendinous bifurcations. The two parts of the dorsal apodeme fuse and run caudad to a point on the ventrolateral surface of the telson. The sixth anterior oblique muscle clearly flexes the telson in addition to its previously mentioned activities.

The sixth anterior oblique muscle of *Penaeus* is undoubtedly homologous with that of *Pandalus* and *Astacus*. Berkeley and Schmidt indicate the presence in the latter forms of the ventral apodeme of the anterior oblique muscle. They do not mention the dorsal apodeme of this muscle.

POSTERIOR LOOP OF ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

FIGURES 60, 61

Arising from the posterior portion of the dorso-medial area of the anterior oblique muscle, where the bilateral pairs of the latter are fused at the midline, is the posterior loop of the anterior oblique muscle. The muscle runs ventrocaudally, laterad of the central muscle, and then passes mesad to join the transverse muscle. At the mid-

line, the fibers of the posterior loop connect with those of its partner on the other side of the segment. Each anterior oblique muscle of *Penaeus* appears to have a posterior loop.

EXTERNAL ARM OF ANTERIOR OBLIQUE MUSCLE OF ABDOMEN

FIGURES 60, 63, 64

The external arm of anterior oblique muscle is much larger and more important than the posterior loop. The external arm connects with the broad dorsal end of the anterior oblique muscle at the midline (fig. 61). The muscle sweeps over the central muscle dorsally and then runs antero-ventrad along the outside of the abdominal muscles to a ventrolateral point just inside the next anterior abdominal somite (fig. 60). Daniel (1931c) considers this point the origin of the external arm, and he notes that the area is the common insertion of the anterior and posterior obliques of an anterior segment. To summarize, each external arm originates on the posteroventral edge of an abdominal segment and traverses very nearly the whole of a segment to its insertion on the main body of the anterior oblique muscle. Taking the anterior oblique muscle as a whole, including its external arm, the muscle functionally traverses three segments from the ventral surface of one, over the central muscle of the second, dorsally, and hence to the ventral surface of the third segment.

As one would expect, the abdomen of *Penaeus* contains 6 external arms accompanying 6 anterior oblique muscles (fig. 60). Its counterparts in the abdomen of *Pandalus* and *Astacus* are not clear.

POSTERIOR OBLIQUE MUSCLES OF ABDOMEN

FIGURES 61, 64

The abdomen of *Penaeus* has five posterior oblique muscles, beginning with the first abdominal segment. The posterior obliques are associated with the anterior obliques and function with the latter in the flexion of the abdomen. Except for the first posterior oblique, the muscles arise dorsal to the transverse muscles in close association with the head or anterior end of the central muscles. The posterior oblique then runs ventrocaudally through two segments to insert ventrad in common with its accompanying anterior oblique muscle (fig. 61). The first posterior oblique originates in tendinous tissue dorsal to the anterior

thoracic muscles. No posterior oblique muscle has been identified in the highly modified sixth abdominal segment of *Penaeus*. The posterior oblique muscles of *Penaeus* are represented in the abdomen of *Astacus* and *Pandalus*. The latter forms also have five posterior obliques.

OBLIQUE TRANSVERSE MUSCLES OF ABDOMEN

FIGURE 61

The oblique transverse muscle splits off the anterior oblique muscle dorsomedially at the junction of the main anterior oblique muscle and the external arm. The fibers turn ventrad between the anterior oblique and the central muscle and there join the central muscle. The affinities of the oblique transverse muscles of *Penaeus* in *Astacus* and *Pandalus* are not known.

1. Pleopods

The pleopods or swimming legs in many natant Crustacea are well-developed swimming appendages that enable the animals to propel themselves forward rapidly for great distances. In *Penaeus setiferus* the pleopod is not only heavily muscled but so constructed that the muscles move the pleopod, and in particular the propellers, the exopodite and endopodite, through a long power stroke. Each pair of pleopods beats in unison. The beat of the pleopods is synchronized by volleys of nerve impulses passing along the ventral abdominal nerve cord so that a beat wave reminiscent of peristaltic action passes down the abdomen from anterior to posterior. *P. setiferus* has five pairs of pleopods, typical of the Crustacea Natantia. The third set has been chosen for study here because of their unspecialized structure.

When feeding, the white shrimp creeps along the bottom on its long, slender walking legs, testing the substrate for food particles. The cephalic region of the shrimp is held high while the tail fan and telson rest on the bottom. As the animal moves about, the pleopods beat gently from time to time. The resulting flow of water aids in the animal's progress by slightly lifting the tail fan from the bottom. Close examination of the white shrimp on the bottom reveals that the main propulsive elements, the exopodites, beat in a lateral position, not brushing the substrate. The present writer assumed that when the shrimp, encountering an obstruction, rises free of the substrate, the

pleopod exopodites would beat in a vertical plane, in a straight line with the proximal elements of the pleopods. Such is not the case, however. In *P. setiferus* the pleopod exopodites always beat in a horizontal plane at right angles to the proximal pleopod elements during the power stroke, no matter whether the animal is creeping along the bottom or swimming freely in the water above. Dr. Edward Peebles, Tulane Medical School, has suggested that the lateral position of the pleopod exopodites places these organs outside of the stream of turbulence created by the walking legs.

The return or recovery stroke, during which the pleopod exopodite must be feathered, is made in the vertical plane, or nearly so. This action might be thought to cause the long exopodite to drag on the bottom, but during the return stroke the organ is relaxed and bends before the pressure of the water passing over it. The distal tip of the exopodite describes an oval. The power stroke draws the tip in a flat arc lateral to the ventral plane of the abdomen. At the end of the power stroke, water pressure from the anterior causes the exopodite to bend caudally and, together with the contraction of rotator and flexor muscles, to rotate one-quarter of a turn about its longitudinal axis. Upon rotation of the exopodite, the inertial drift of the animal through the water helps extensor muscles bring the exopodite from the lateral position of the power stroke to a ventral position. Rotation of the exopodite also enables the organ to present to the flow of water its cross section of least resistance as it is brought forward in recovery.

SKELETAL ELEMENTS

The skeletal and muscular elements of the third pleopod of *Penaeus setiferus* are arranged to perform the functions described above. The skeletal parts are roughly similar to those of the third pleopod of the European crawfish *Astacus astacus*, to those of the first pleopod (the pleopod of the second abdominal segment) in the female of *Calinectes sapidus*, to those of the third pleopod of *Cambarus longulus longulus*, and to the parts of the third pleopod of *Pandalus danae*.

Where the third pleopod of *Penaeus setiferus* is attached to the abdominal venter, the region of articulation of the ventral skeleton is reinforced by two V-shaped structures comprised of sclerotized bars (fig. 59). The lateral V (fig. 65, ventral skeletal support), traditionally said to be a contribution of the pleuron, has its apex directed

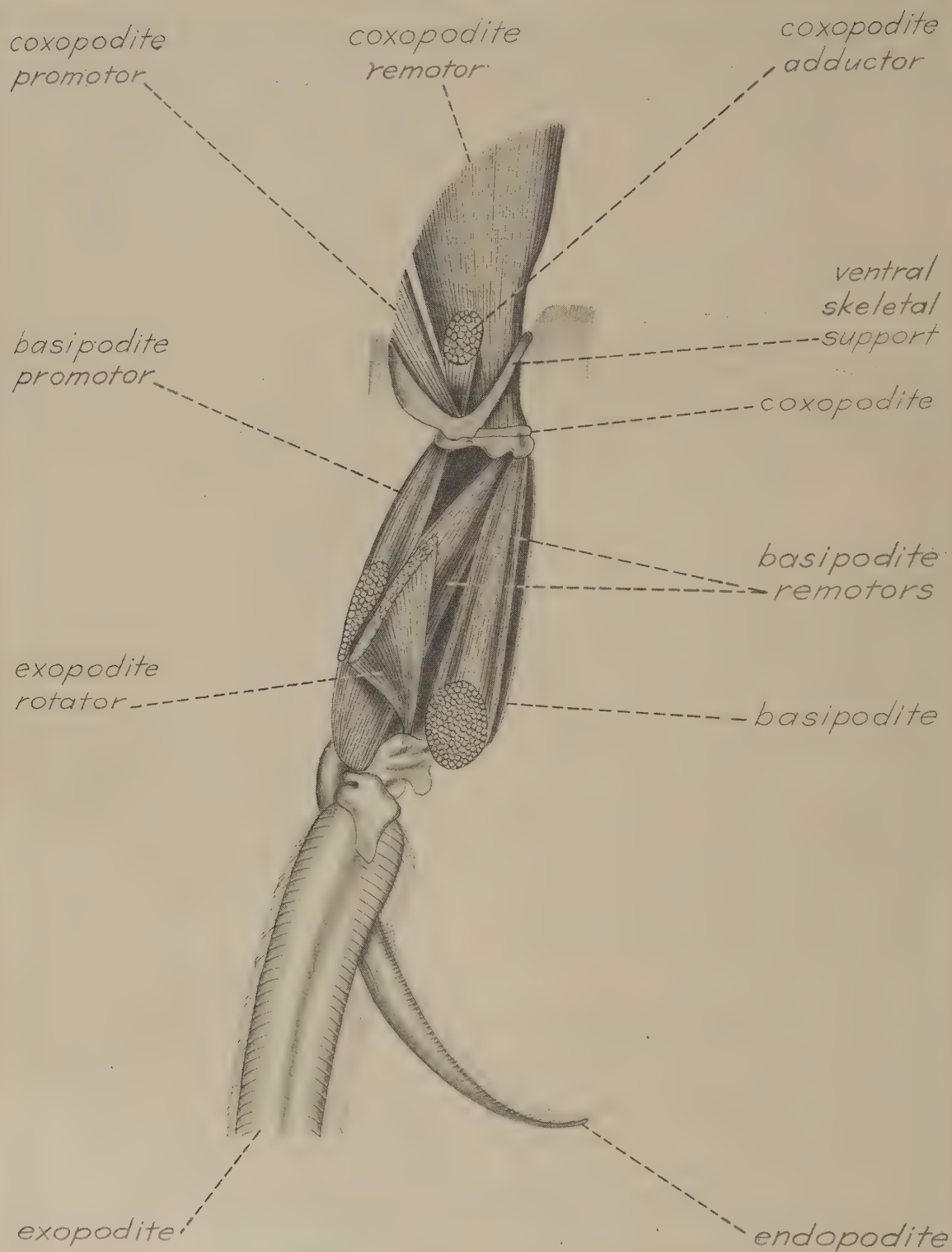


FIGURE 65.—Lateral view of left third pleopod showing basipodite and coxopodite musculature.

ventrad. At this point the structure articulates with the pleopod coxopodite. The mesial V, which also points ventrad, constitutes the support for the inner articulation of the pleopod coxopodite. The mesial V is a sternal element. The dorsal or free ends of the lateral and mesial arms of the V's are joined. The resulting ventral skeletal structure represents a set of combined triangles, if the coxopodite is included as a structural member, which can be described as a kind of Warren truss. The structural triangles provide support for the articular foramen to which the pleopod is attached. The ventral skeletal structure pertaining to the support of the pleopod is lighter in construction in *Penaeus* than in *Astacus* or *Cambarus*, although the pleopods of *Penaeus* are comparatively much larger and more powerful.

A further structural difference of note between the ventral skeletal pleopod support in *Penaeus* and that of the *Astacura* and the *Brachyura* is the position of the appendage relative to the ventral abdomen. In the *Astacura* particularly, the articular foramen of the pleopod is fully ventral, the ventral sternum and the so-called ventrolateral pleural plates being flat and lying horizontally. In *Penaeus* by contrast the sternum is convex and the lateral plates heretofore considered pleural are nearly vertical, placing the pleopod in a ventrolateral position exposed to the water in which the organ functions. Although Berkeley does not consider the subject, her illustrations indicate that the situation in *Pandalus* is similar to that in *Penaeus*.

The coxopodite (fig. 65) of the pleopod is a narrow, incomplete band or ring which articulates with the ventral skeletal elements dorsally and the basipodite ventrally. To the coxopodite are inserted muscles having their origins on the tergal plates of the abdominal skeleton and in the basipodite. The basipodite (fig. 65) is a broad and elongate structure, shaped like an inverted heart. The form of its cross section through the broad, distal portion is streamlined to reduce its resistance to the water during the recovery stroke. The basipodite articulates with the coxopodite proximally and with the endopodite and exopodite distally. By virtue of the arrangement of points of articulation and of its extensive musculature, the basipodite substantially reinforces the length and power of the propulsive stroke of the pleopod.

Distal to the basipodite, and visible only on the mesial side of the pleopod, lies a small, heavily sclerotized structure (fig. 68), the exopodite articular element, which articulates with the basipodite and the exopodite. Until more information is available, this structure will be considered an exopodite component. In *Penaeus*, several muscles insert in the structure. No trace of this appendage segment appears in the modified pleopods of *Astacus*, *Cambarus*, or *Callinectes*. Berkeley makes no reference to the structure in *Pandalus*.

The endopodite (fig. 68) articulates with the basipodite by a somewhat constricted area on the mesial side of the pleopod. Functionally the endopodite is simple, accompanying the exopodite in its movements. The exopodite (figs. 65 to 69) is a large, broad, oar-shaped organ, convex anteriorly and concave posteriorly. Like the endopodite, it is comprised of a long series of lightly sclerotized rings alternating with bands of thin cuticle. The skeleton of the exopodite is arranged thereby to bend when a stream of water strikes it from the anterior and to remain straight when water strikes it from the posterior. The exopodite functions like a curved, steel carpenter's rule which may be rolled on a spool in one direction, but which presents resistance to bending in the opposite direction. The functional surface of the endopodite and exopodite oar blades is greatly increased by long, plumose setae embedded in the margins. The pleopod endopodite and exopodite have been called flagella in *Astacus* and other reptant decapods in which the organs are extremely reduced by comparison with those of *Penaeus* and *Pandalus*. The term will not be used here, since the endopodites and exopodites of *Penaeus setiferus* are not flagella.

MUSCLE ELEMENTS

The third pleopod of *Penaeus setiferus* is operated by 17 or more muscles. The number of discrete muscles intrinsic to the pleopod endopodite in *Penaeus* is not clear, although the organ may contain two or more muscles. Schmidt in his account of *Astacus* describes 9 muscles for the third pleopod, Cochran lists 6 for the first pleopod of *Callinectes*, Hart (1952) describes 9 for the third pleopod of *Cambarus longulus longulus*, and Berkeley finds 11 in *Pandalus*.

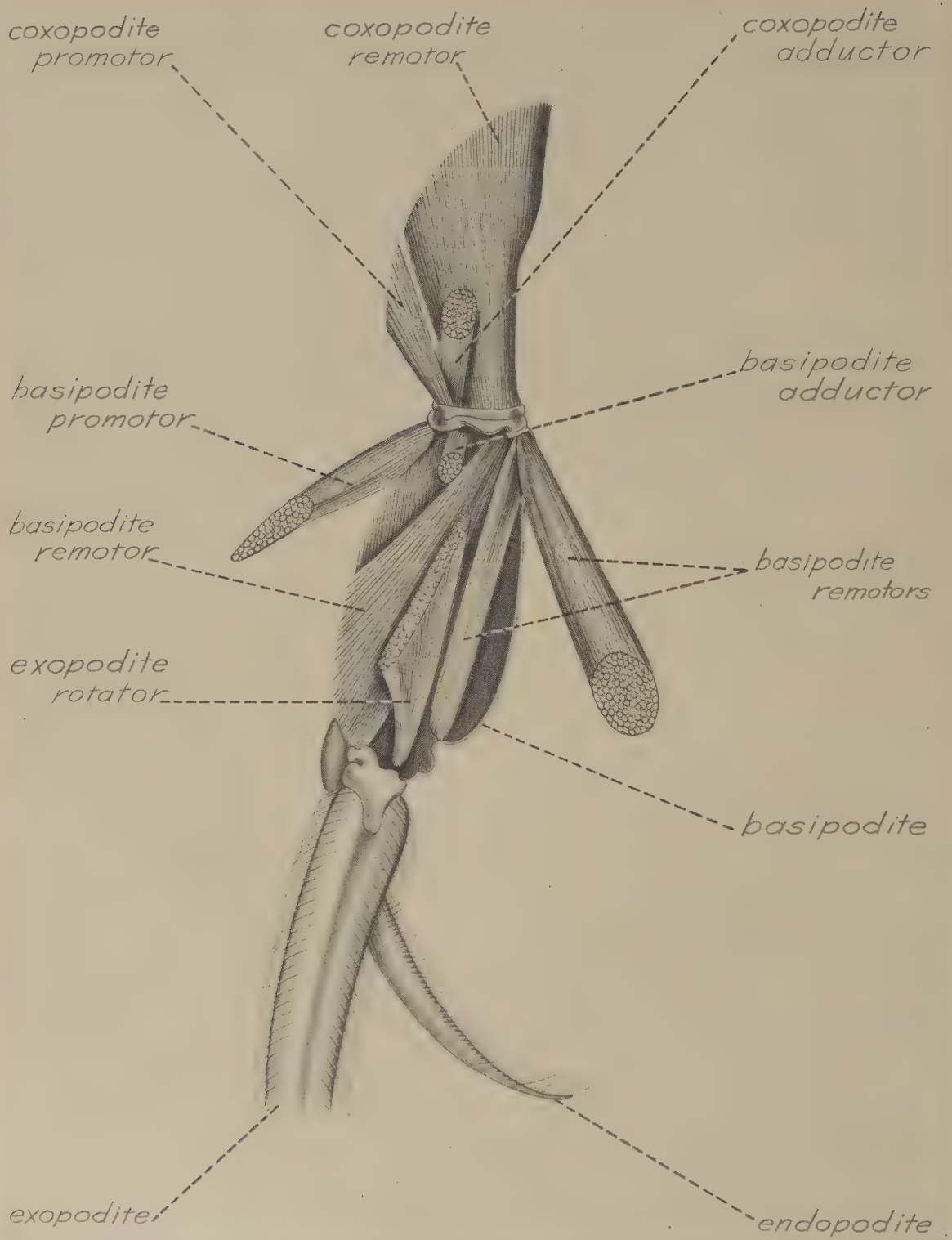


FIGURE 66.—Lateral view of left third pleopod showing basipodite and coxopodite musculature.

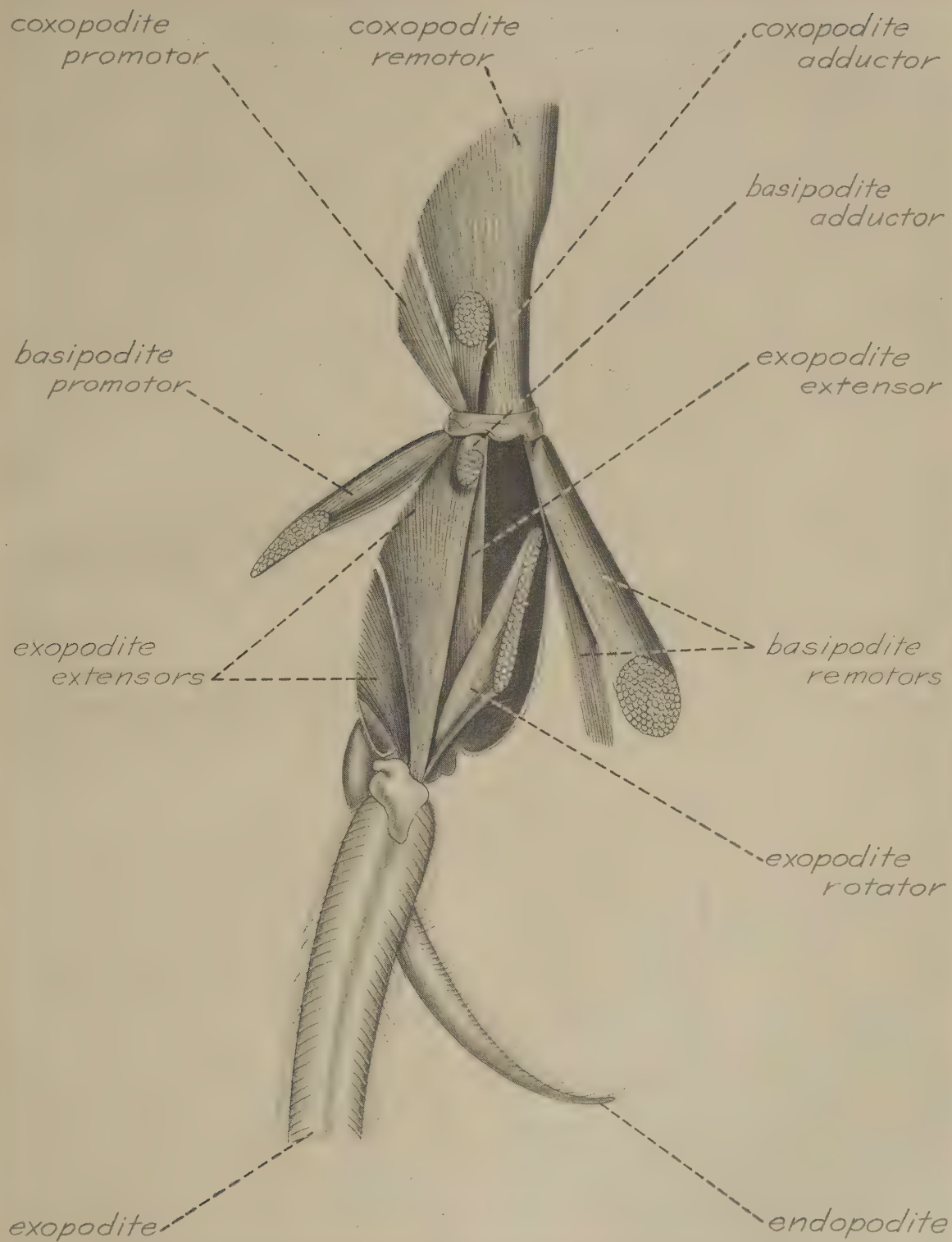


FIGURE 67.—Lateral view of left third pleopod showing basipodite and coxopodite musculature.

COXOPODITE REMOTOR MUSCLE

FIGURES 65 TO 69

The coxopodite remotor muscle, a broad up-right fan, originates in fibrous connective tissue attached to the tergum, the arched dorsal plate of the abdominal skeleton, and inserts about an arc which represents approximately the posterior third of the coxopodite. The area of insertion of this muscle is located on the proximal margin of the coxopodite. Upon contraction the muscle draws the posterior edge of the coxopodite dorsad on the points of coxopodite articulation. The pull is an efficient one, since the coxopodite remotor contracts in an almost straight line. The contraction of the muscle thereby draws the distal pleopod elements through an anterior-to-posterior arc.

Berkeley illustrates the muscle in her work on *Pandalus*. Schmidt in *Astacus* and Hart (1952) in *Cambarus* describe a coxopodite remotor muscle (musculus remotor III pedis spurii) for each of these species of crawfishes. On the basis of information presently available, the coxopodite remotor muscle of *Penaeus* is homologous with the musculus remotor III pedis spurii of *Astacus*, *Cambarus*, and *Pandalus*, although proof must wait upon a comparative study of the nerves. Cochran does not find a coxopodite remotor in *Callinectes*.

COXOPODITE PROMOTOR MUSCLE

FIGURES 65 TO 69

The coxopodite promotor muscle originates on the abdominal tergum anteroventrally to the coxopodite remotor muscle and inserts on the proximal margin of the coxopodite anterior to a line through the lateral and mesial articulation points at which the coxopodite articulates with the ventral abdominal skeleton. Mechanically, the action of the coxopodite promotor is much weaker than are the contractions of the coxopodite remotor, since the coxopodite promotor is smaller than the remotor and its area of insertion is much closer to the proximal fulcral line of the coxopodite than is that of the remotor muscle.

The coxopodite promotor pulls the preaxial margin of the coxopodite dorsad. The distal pleopod elements thus are drawn through an arc anteriorly, in opposition to the action of the coxopodite remotor. The coxopodite promotor muscle

has no exact functional counterpart in *Astacus* and *Cambarus*. The only muscle in the latter animals which could be homologous with the coxopodite promotor in *Penaeus* is the musculus rotator dorsalis basipoditis II pedis spurii described by Schmidt in *Astacus* and by Hart (1952) in *Cambarus*. The fact that the dorsal rotator muscle inserts on the basipodite in *Astacus* and *Cambarus* argues against this conclusion.

Berkeley describes a musculus rotator dorsalis basipoditis III pedis spurii in *Pandalus* that is very likely the homolog of the coxopodite promotor muscle in *Penaeus*, despite the difference in origin. However, her adoption of Schmidt's name for this muscle is unfortunate, since the musculus rotator dorsalis basipoditis in *Pandalus* actually promotes the basipodite, rather than rotates the pleopod. An appendage rotates about its long axis, not about an axis transverse to the appendage.

Cochran describes a coxopodite promotor muscle (musculus promotor coxopoditis I pedis spurii) in the pleopod of the second abdominal segment of *Callinectes*, and possibly the muscle is homologous with the coxopodite promotor in *Penaeus*. To remove from the realm of speculation a discussion of muscles having similar functions in two forms as distantly related as *Penaeus* and *Callinectes* will require careful study of the nerves.

COXOPODITE ADDUCTOR MUSCLES

FIGURES 65 TO 67, 69

Two coxopodite adductor muscles are found in the pleopod of the white shrimp. The lateral coxopodite adductor muscle (figs. 65 to 67) is a small, short muscle originating on the lateral tergal surface and inserting on the dorsomedial rim of the coxopodite. The muscle pulls the mesial side of the coxopodite dorsally, and in so doing brings the distal pleopod elements mesad. The action of the lateral adductor muscle is reinforced by a second coxopodite adductor muscle (fig. 69) located on the mesial side of the ventral skeletal support. The mesial adductor originates on a phragmal fold of the postcoxal sternum and inserts on the dorsomedial margin of the coxopodite.

The lateral coxopodite adductor muscle has been lost in the Astacura to which reference has been made, but appears in *Callinectes* as the largest muscle in the blue crab pleopod. The

mesial coxopodite adductor muscle, which has disappeared in *Callinectes*, possibly exists in *Astacus*, *Cambarus*, and *Pandalus* as the ventral basipodite rotator muscle (musculus rotator ventralis basipoditis III pedis spurii). From Berkeley's account of the origin, insertion, and action of the muscle in *Pandalus*, the musculus rotator ventralis basipoditis is really an adductor muscle. Again, a study of the nerves is called for.

BASIPODITE ABDUCTOR MUSCLE

FIGURE 68

Opposing the action of the coxopodite adductor muscles which tend to draw the basipodite and other distal parts of the pleopod towards the abdomen is the basipodite abductor muscle. This fan-shaped muscle originates on the mesial surface of the basipodite. The area of origin is elongate in the long axis of the basipodite (fig. 68). The muscle becomes narrow as it courses dorso-laterally to insert on the ventrolateral margin of the coxopodite. When the basipodite abductor muscle contracts, the basipodite and the distal pleopod elements are drawn away from the abdomen. The muscle has been lost in *Callinectes* and in the *Astacura* referred to above. The basipodite abductor muscle of *Penaeus* may have a homolog in *Pandalus* as the musculus adductor basipodite II pedis spurii, a muscle that, according to the description of Berkeley, is evidently an abductor.

BASIPODITE PROMOTOR MUSCLE

FIGURES 65, 66, 67, 69

The basipodite promotor muscle originates on an area of the anterior margin of the basipodite about one-third of the distance from the distal end of the basipodite. The muscle inserts on the anteroventral margin of the coxopodite. Muscle contractions draw the basipodite, and the distal parts of the pleopod, cephalad with respect to the coxopodite. Together with the coxopodite promotor, the basipodite promotor muscle moves the pleopod in its recovery stroke. The basipodite promotor extends the length of the stroke. No homolog of the basipodite promotor muscle in *Penaeus* is evident in the pleopod of *Astacus*, *Cambarus*, or *Callinectes*.

The third pleopod of *Pandalus*, however, has a muscle, the musculus productor basipoditis III

pedis spurii, that is similar in function and arrangement to the basipodite promotor muscle of *Penaeus*. The muscles in the two animals are almost certainly homologous, although proof must wait upon a study of nerve-muscle connections. Berkeley's use of the term "productor" for this muscle is questionable. She says, in part, that the musculus productor basipoditis ". . . moves the basipodite forward. . . ." In the opinion of the present writer, the action described by Berkeley for the muscle is not production, but promotion. Production, in the present context, is synonymous with the extension in length of a limb, and the musculus productor basipoditis of the third pleopod of *Pandalus* does not appear to be an extensor muscle.

BASIPODITE REMOTOR MUSCLES

FIGURES 65 to 69

The pleopod of the white shrimp contains 3 basipodite remotor muscles. Their contractions serve to increase greatly the length and power of the propulsive stroke initiated by the coxopodite remotor muscle. The anterior basipodite remotor muscle originates over a broad area in the anteroventral lobe of the basipodite (figs. 65, 66). From its origin the muscle runs diagonally across the interior of the basipodite to insert on the ventral edge of the coxopodite, posterior to a line through the points of articulation between the basipodite and the coxopodite. The posterior basipodite remotor muscles, of which there are two (figs. 65, 68), originate in the posteroventral lobe of the basipodite and extend dorsad to insert on the ventral margin of the coxopodite postaxially.

On the basis of arrangement and size, the anterior basipodite remotor muscle of *Penaeus* is evidently a homolog of the musculus reductor basipoditis III pedis spurii of *Pandalus*. The muscles are also functionally similar. Here again a question is raised over the use of terms. Berkeley describes the action of the musculus productor basipoditis as antagonistic to that of the musculus reductor basipoditis. If, as has been established above, the name "productor" for muscle function is synonymous with the term "extensor," then it follows that a reductor muscle is the same as a flexor muscle. Unfortunately, reduction has not in the anatomical sense the opposite meaning of production, but implies additional functions. For this reason the name "reductor" for muscle

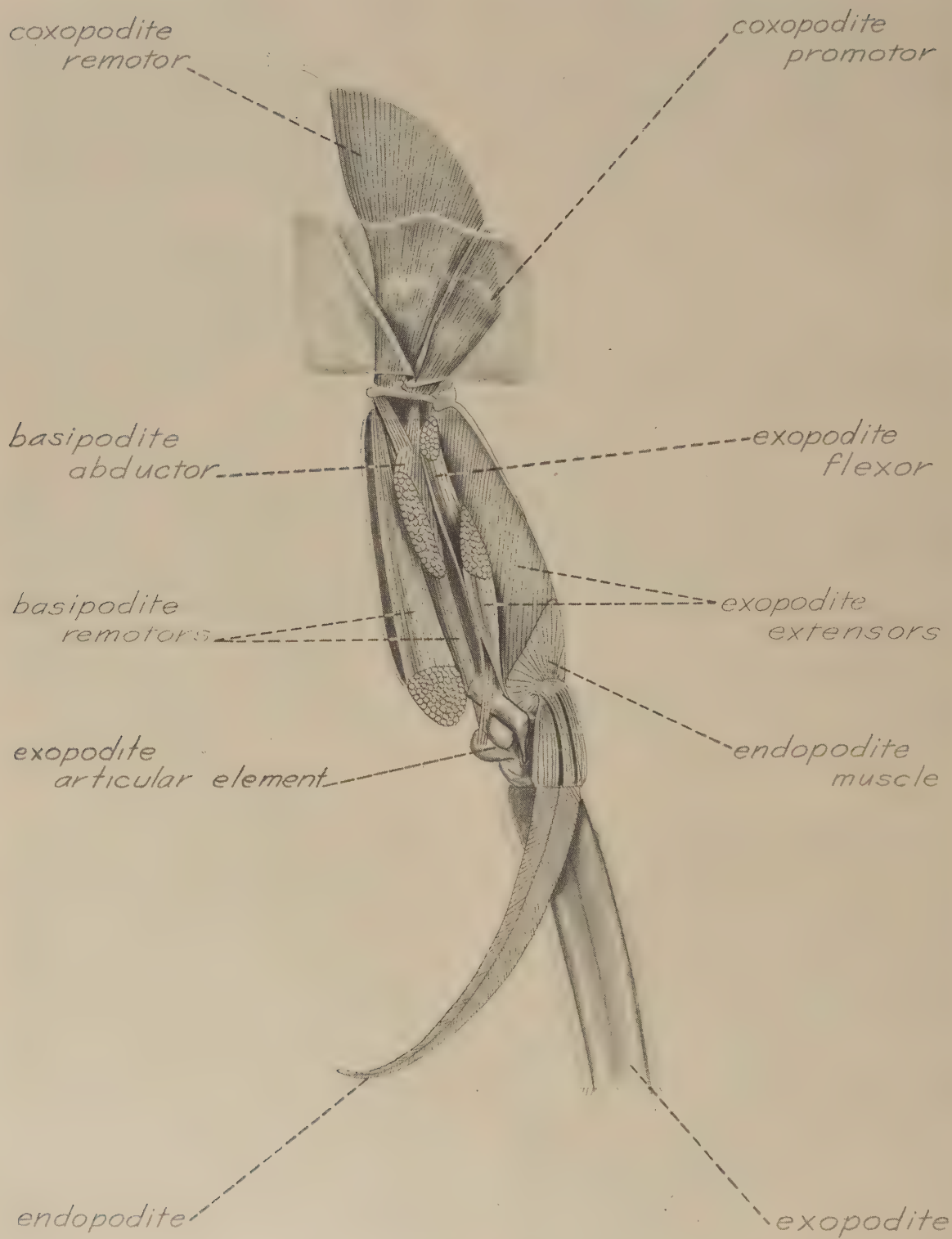


FIGURE 68.—Mesial view of left third pleopod showing coxopodite, basipodite, and endopodite musculature.

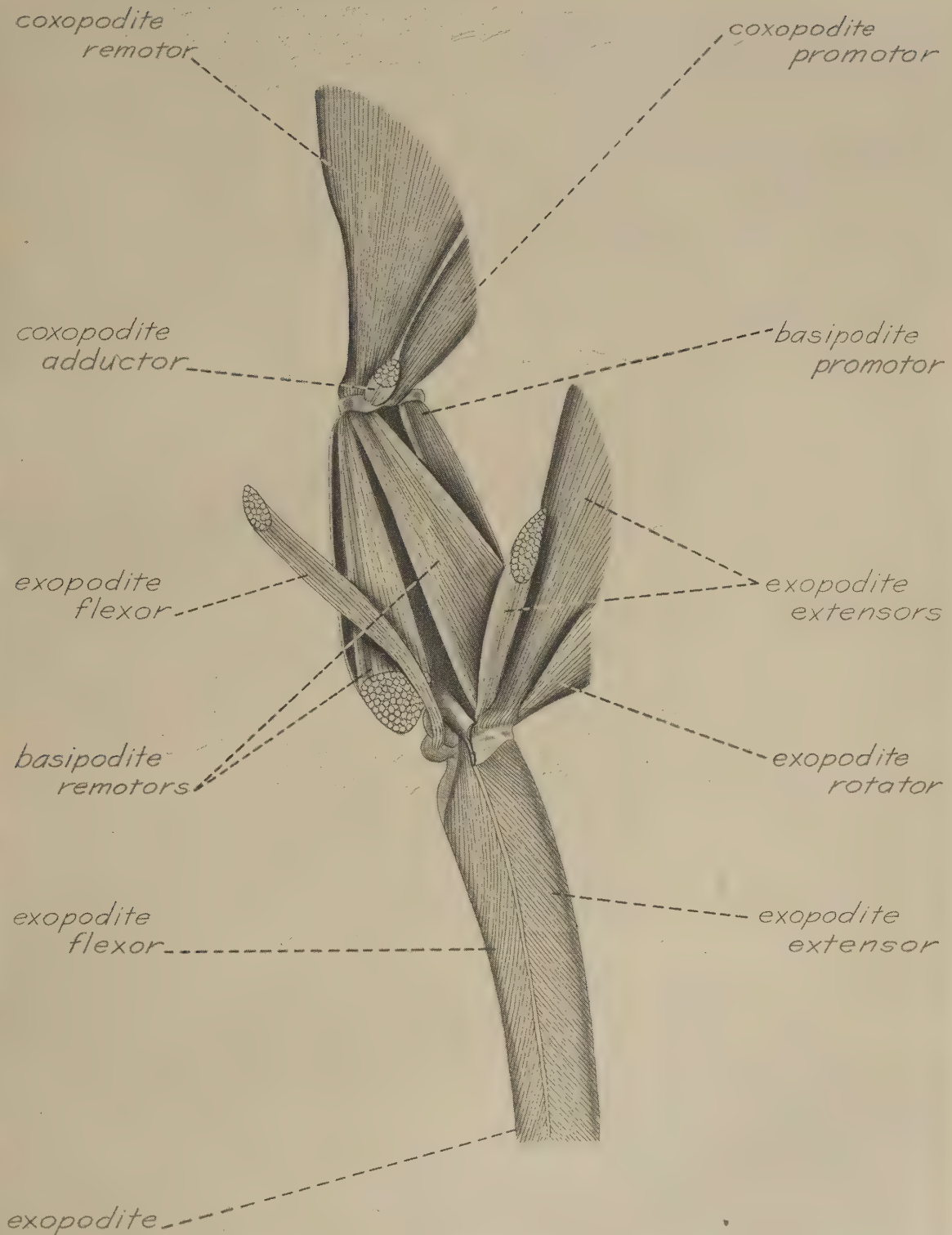


FIGURE 69.—Mesial view of left third pleopod showing muscles of coxopodite, basipodite, and exopodite. Endopodite removed.

action should be avoided. Moreover, the application of the term "reductor" to the musculus reductor basipoditis in *Pandalus* is questionable, since the muscle does not function as a flexor of the basipodite.

The basipodite remotor muscles apparently are not represented in the pleopod of *Astacus*, *Cambarus*, or *Callinectes*, although a trace of these important muscles in *Penaeus* may have remained during the evolution of the former animals as the basipodite reductor muscle (musculus reductor basipoditis).

EXPODITE ROTATOR MUSCLES

FIGURES 65, 66, 67, 69

The pleopod basipodite of *Penaeus setiferus* contains two muscles which rotate the exopodite. The elongate origin of the fan-shaped lateral exopodite rotator muscle is on the lateral surface of the basipodite and may be identified easily through the cuticle in preserved material (fig. 65). The muscle becomes narrow as it runs distomesially to insert on the mesial surface of the complex articular element joining the basipodite to the exopodite. The muscle functions to square the plane of the oar blade of the exopodite at the end of the recovery stroke for the catch of the following propulsive stroke. The mesial exopodite rotator muscle (fig. 69) is a small muscle located in the anterodistal lobe of the basipodite. Its contractions apparently aid in feathering the exopodite oar blade. These muscles do not appear in any of the crustaceans to which reference has been made, although the mesial exopodite rotator muscle may exist in *Pandalus* as the musculus adductor endopoditis.

EXPODITE EXTENSOR MUSCLES

FIGURES 67 TO 69

The third pleopod of *Penaeus setiferus* is provided with 3 exopodite extensor muscles, 2 of which originate in the basipodite and 1 in the exopodite. The largest of these has its origins along the entire anterior margin of the basipodite (figs. 67, 68). It is one of the largest muscles in the pleopod. The muscle inserts on an apodeme of the exopodite articular element. Inserting on the same apodeme is a slender exopodite extensor muscle which is located just posterior to the anterior exopodite extensor muscle described above (figs.

68, 69). The posterior exopodite extensor has its origin on the mesial surface of the basipodite (fig. 68). When the two extensor muscles contract they draw the articular element of the exopodite dorsad. The plane of articulation of this element is such that the exopodite is swung down from its lateral position in the propulsive stroke to a fully extended position. At the same time the articular element rotates the exopodite on its longitudinal axis, with the aid of the mesial exopodite rotator muscle, to feather the exopodite oar blade.

That the two exopodite extensor muscles located in the pleopod basipodite of *Penaeus* are missing in *Astacus*, *Cambarus*, and *Callinectes* is not surprising in view of the extensive rearrangements in the swimming appendages of the latter form. The pleopod basipodite of *Pandalus* has retained at least the larger of the exopodite extensor muscles of *Penaeus*, as the musculus adductor exopoditis III pedis spurii.

The third exopodite extensor (fig. 69) lies along the mesial edge of the exopodite, attaching on the margin of the exopodite articular element. Its contractions serve to extend the exopodite and to straighten the leading edge of the exopodite during the recovery stroke. The muscle is homologous with the exopodite flagellum muscle described by Schmidt and Hart (1952) in *Astacura*, and by Berkeley in *Pandalus*. It is missing in *Callinectes*.

EXPODITE FLEXOR MUSCLES

FIGURES 68, 69

Two exopodite flexor muscles appear in the third pleopod of *Penaeus setiferus*. One originates proximally on the medial surface of the basipodite (fig. 68), runs the length of the basipodite, and inserts on the posterior surface of the exopodite articular element (fig. 69). The muscle functions to flex the exopodite laterally to place the exopodite oar blade in position for the propulsive stroke. The proximal exopodite extensor muscle of *Penaeus* is in all likelihood the homolog of the musculus abductor exopoditis of the third pleopod of *Pandalus*.

The distal exopodite flexor muscle is intrinsic to the exopodite. The muscle is arranged along the lateral margin of the exopodite (fig. 69) and attaches to the posterior part of the exopodite articular element. The contractions of the muscle reinforce the flexing action of the proximal flexor

muscle. Like the distal exopodite extensor muscle, the distal exopodite flexor muscle is a homolog of the exopodite flagellum muscle (musculus flagellaris exopoditis III pedii spurii) in *Astacus*, *Cambarus*, and *Pandalus*.

ENDOPODITE MUSCLE

FIGURE 68

The endopodite muscle, or muscles, appears to be a multipart structure in *Penaeus setiferus*. A portion of the muscle lies mesially in the antero-distal lobe of the basipodite (fig. 68). Fibers of the muscle run through the constricted article connecting the basipodite with the endopodite and continue distally into the endopodite. Additional fibers that arise in the proximal end of the endopodite suggest that the endopodite contains more than one muscle body. The function of the endopodite musculature consists of stiffening the endopodite during the power stroke and, by their relaxation in the recovery stroke, of enabling the endopodite to bend before the flow of water.

The endopodite flagellum muscle of *Astacus*, *Cambarus*, and *Pandalus* is very likely the homolog of the muscle in the pleopod endopodite of *Penaeus*. The muscle has been lost in the blue crab.

2. Tail Fan

The tail fan is made up of the telson and uropods projecting from the posterior end of the sixth abdominal segment and intimately associated with it. Classically, the uropods have been treated as serially homologous appendages having the typical limb parts, however modified. The telson, on the other hand, has usually been considered an unsegmented posterior element bearing the anus, on grounds of the embryological addition of abdominal segments before the telson (Calman 1909). The whole tail fan is well adapted for the purpose of drawing the white shrimp backwards through the water as the great ventral abdominal muscles flex the abdomen.

SKELETAL ELEMENTS

The telson (figs. 70 to 74) is an apparently unpaired structure. Its broad, anterior portion articulates with the caudal end of the postero-dorsal part of the sixth abdominal somite. Lateral condyles allow free movements in the vertical plane, but limit horizontal motion. The telson

becomes narrow posteriorly, tapering to a sharp point. In section, the telson is roughly triangular. The thin sternum varies in shape from flat to slightly concave and the heavily sclerotized laterotergal plates are convex. Instead of having a mid-dorsal carina or ridge, the telson has a pair of ridges produced by a shallow groove in the dorsal midline (fig. 70). The structure lies above parts of the uropods and affords them some protection dorsally.

The uropods (figs. 70 to 74) arise from the posteroventral area of the sixth abdominal somite. Each is comprised of a strongly sclerotized basal element, the protopodite, from which the broad, flat uropods project. The large lateral uropod is the exopodite while the inner is the endopodite. The protopodite is supposed to be made up of the coxopodite and basipodite, but no skeletal trace of these articles is evident. A faint relic of segmentation in the exopodite remains on the dorsal surface (fig. 70) in the form of a transversely oriented groove. A portion of this groove sets off the tip of the lateral exopodite adductor muscle.

Due to the hinging of the uropodal elements, the uropods are capable of free motion. The articulation between the protopodite and the sixth abdominal segment, while strong, is relatively loose. The protopodite can move in the vertical and horizontal planes and also may rotate about its long axis. The points of articulation between the protopodite and the uropods are, on the other hand, condylic, and limit the exopodite and endopodite to the horizontal movements of spreading the uropods. Certain marginal areas of the telson and uropods are fringed with natatory hairs, much like the pleopods.

MUSCLE ELEMENTS

The muscles of the tail fan are all disposed to the function of these organs in the rapid backward swimming of the white shrimp. In plan the muscles are widely different from that of the typical limb. The most notable difference is the presence inside the sixth abdominal segment of several muscle groups which operate parts of the tail fan. This does not occur in the preceding abdominal segments. In comparing the muscles of the area in *Penaeus* with the tail fan muscles of *Astacus* and *Pandalus*, two kinds of differences are encountered. The first has to do with the number of functional muscle types, *Penaeus* having 16 compared with 18 muscle types in the crawfish.

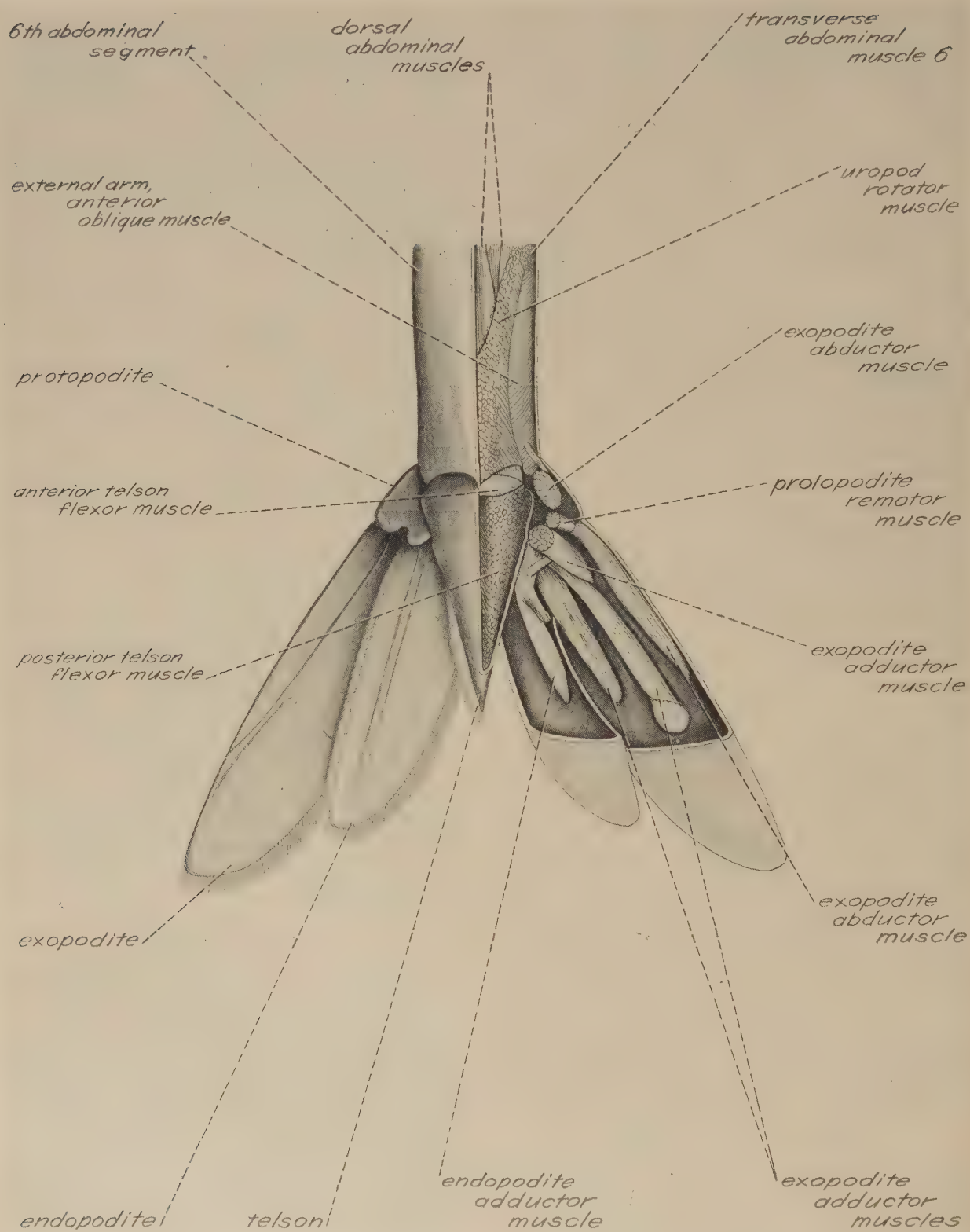


FIGURE 70.—Dorsal view of telson, uropods, and part of sixth abdominal segment. Left side intact. Dorsal cuticle of right side cut away to show muscles.

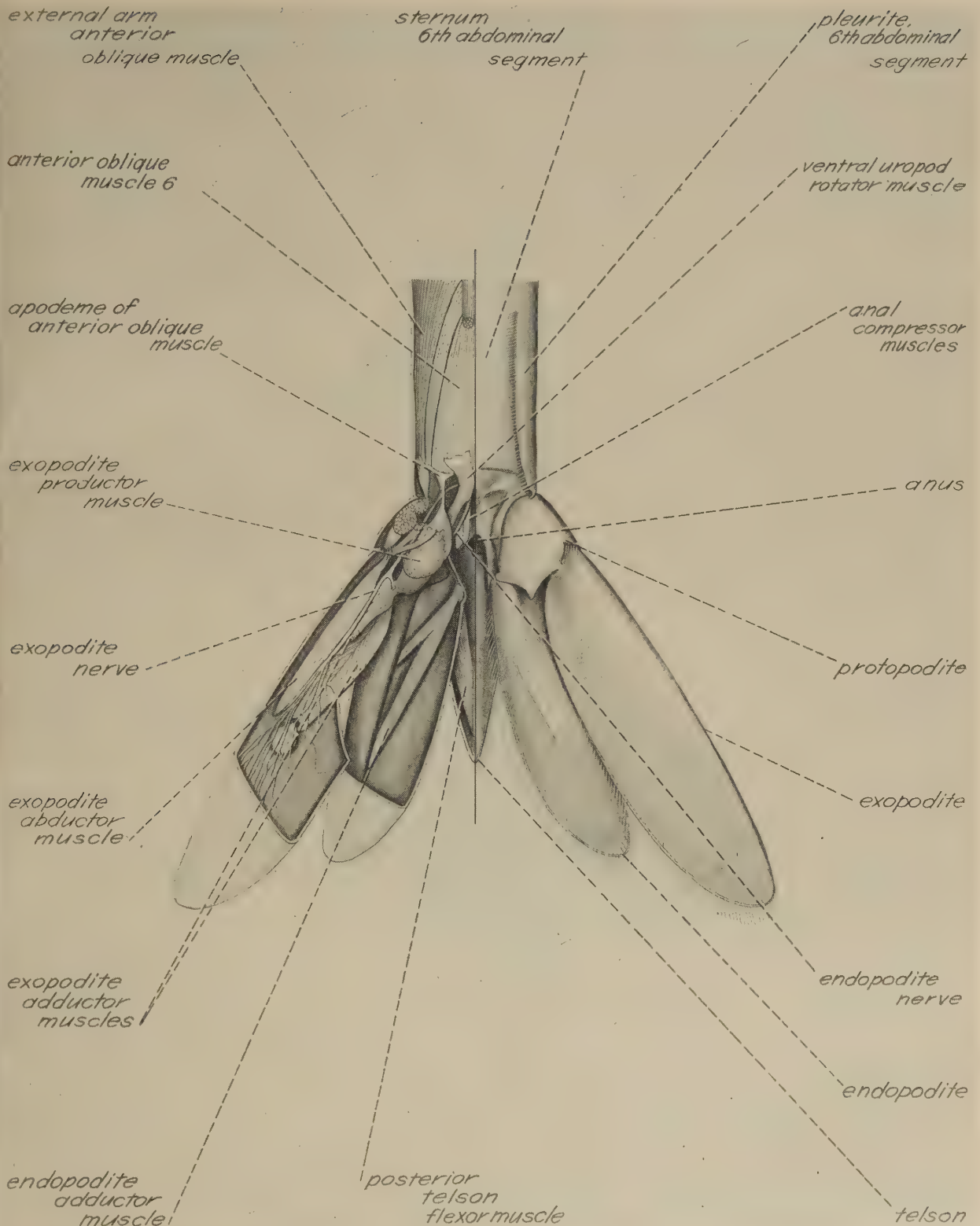


FIGURE 71.—Ventral view of telson, uropods, and part of sixth abdominal segment. Right side intact. Ventral cuticle of left side cut away to show muscles.

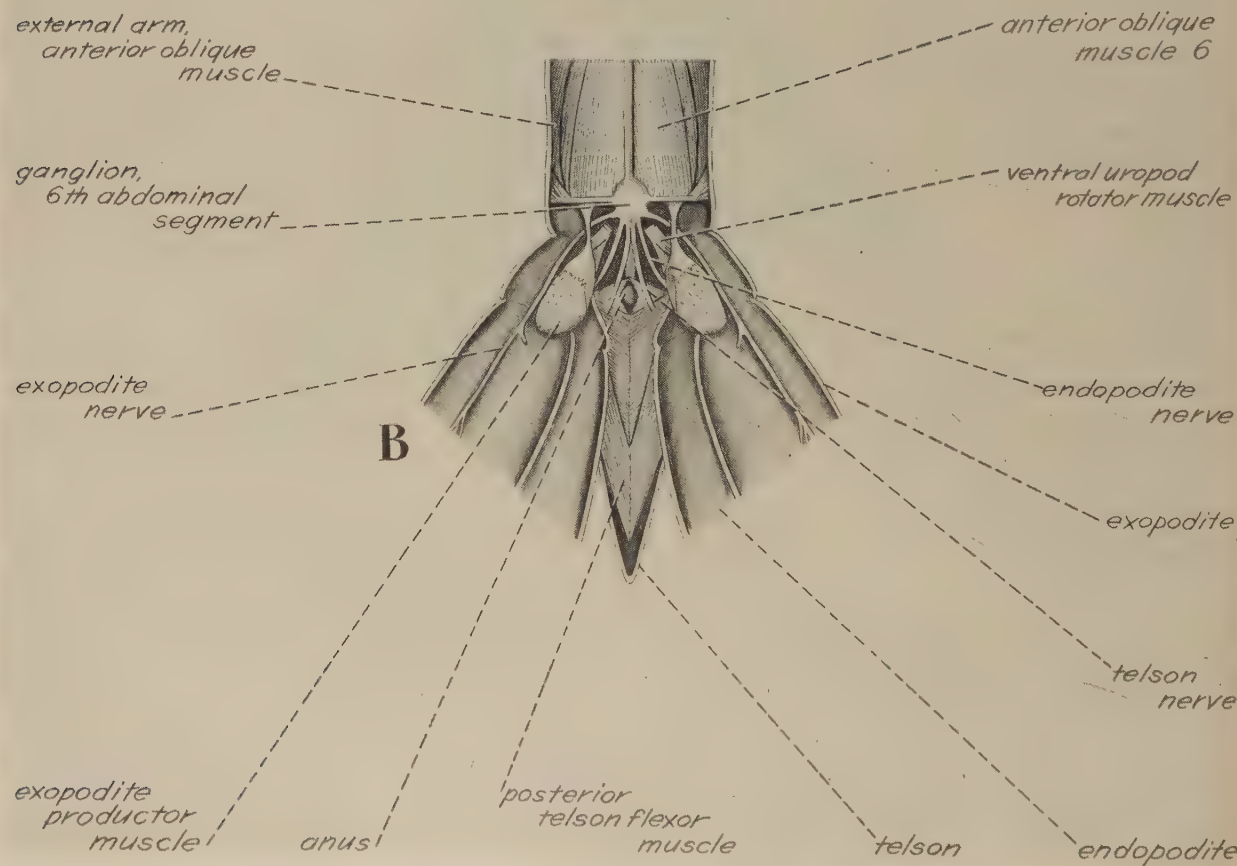
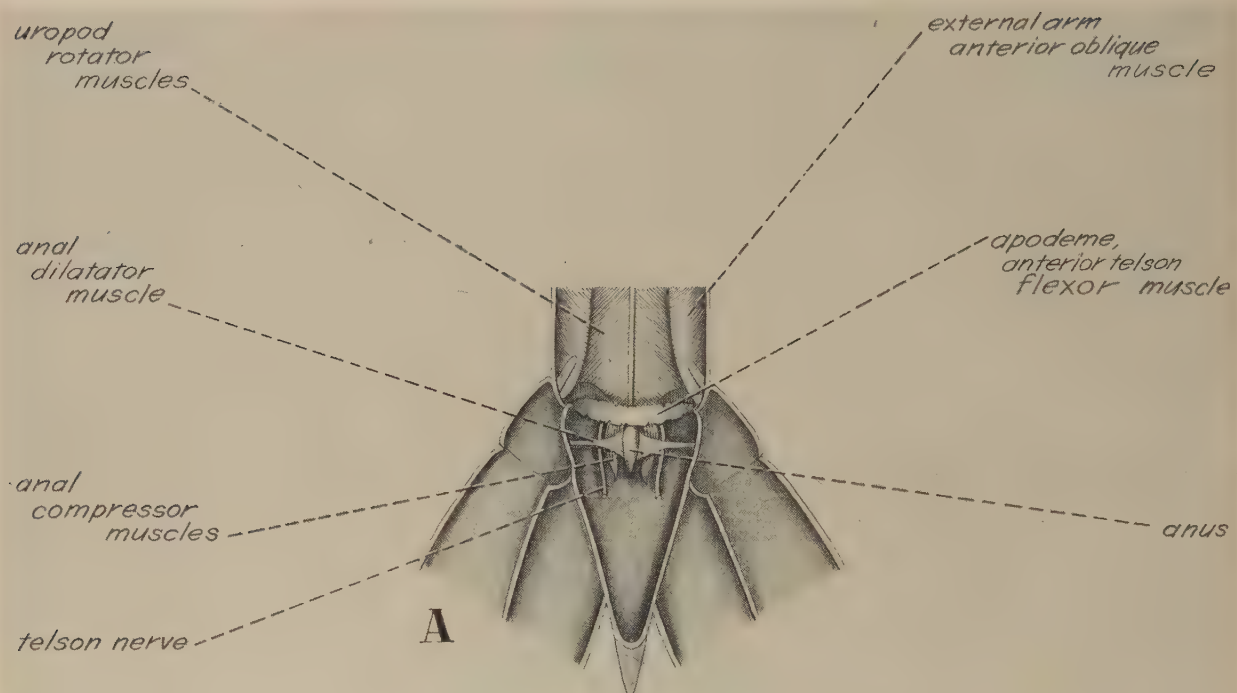


FIGURE 72.—Views of dissections of telson and uropods. A. Dorsal view. Muscles removed to show anus and posterior muscles of sixth abdominal segment. B. Ventral view. Muscles removed to show nerves.

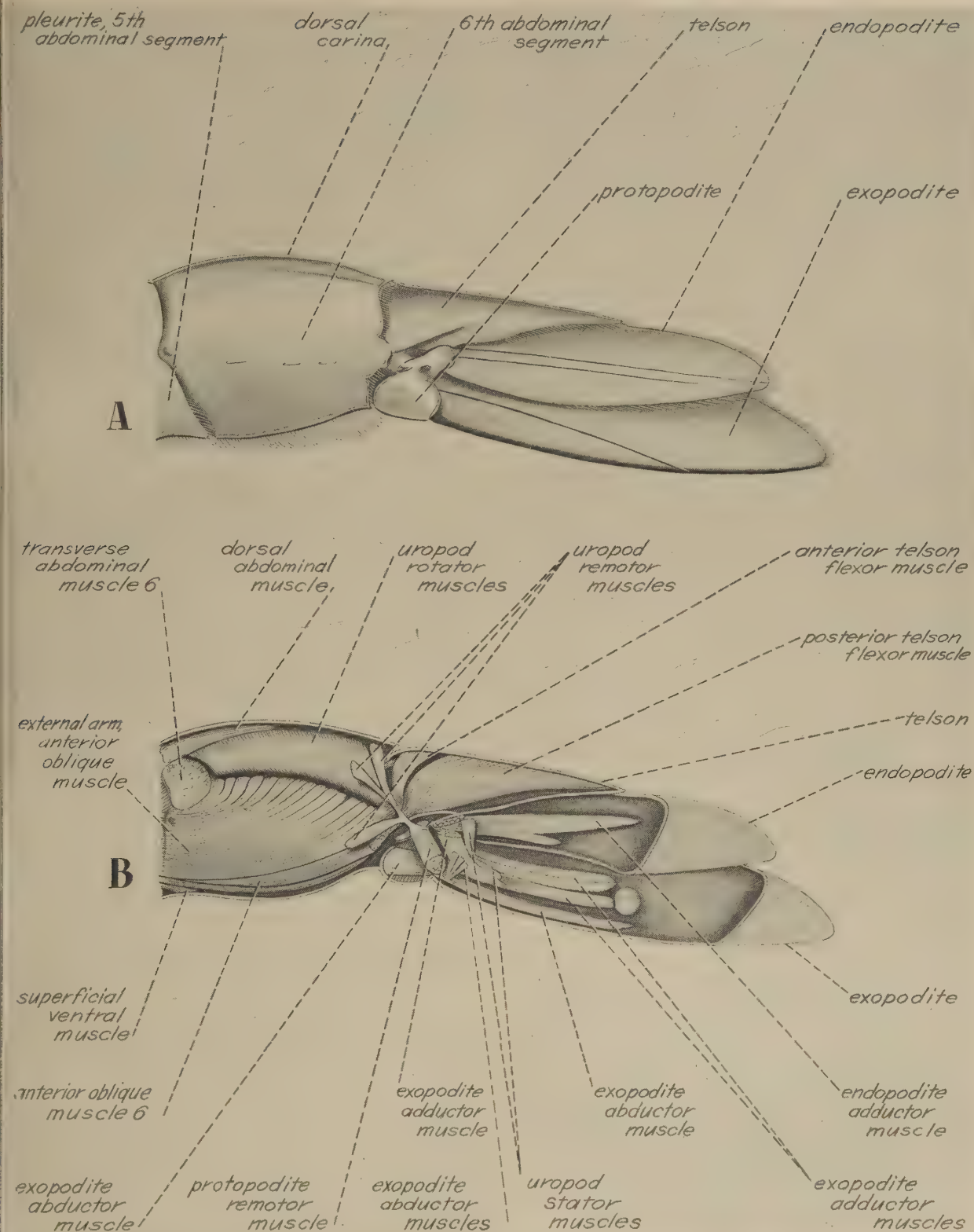


FIGURE 73.—Lateral view of left side of sixth abdominal segment, telson, and uropods. A. Intact structures. B. Lateral cuticle removed to show superficial lateral muscles.

The second difference follows from the first. We find muscles in the tail fan of *Penaeus*, similar to those in *Pandalus* and *Astacus*, that have quite dissimilar functions. These have been renamed in accordance with their presumed functions in *Penaeus*. The tail fan of the white shrimp appears to be operated by 27 muscles, including certain rectal muscles not found in *Astacus* or *Pandalus*. This compares with 18 muscles in *Astacus* and 13 in *Pandalus*.

UROPOD REMOTOR MUSCLES

FIGURE 73

Three uropod remotor muscles are found in the white shrimp. One originates dorsomedially in the posterodorsal corner of the sixth abdominal segment. The second arises slightly anterior to the first and runs posteroventrad to join the first on a common apodeme. The third originates lateroventrad of the first two and joins the common remotor apodeme at a point caudad of the first two remotor muscles. The common remotor apodeme now runs a short distance ventrocaudally to insert in heavy connective tissue beneath the dorsal rim of the uropod protopodite. Contractions of the muscles bring the uropods dorsad, in opposition to the action of the large abdominal muscles attached to the uropods. The two dorsal uropod remotor muscles are perhaps homologous to the medial uropod protopodite remotor muscle of *Pandalus* and *Astacus*, while the ventrolateral remotor is considered to be the homolog of the medial protopodite remotor muscle in the coon stripe shrimp and the crawfish.

PROTOPODITE REMOTOR MUSCLE OF UROPOD

FIGURES 70, 73

The protopodite remotor muscle is a short, thick structure originating in the same tendinous connective tissue upon which the uropod remotors insert. The protopodite remotor thus functionally prolongs the remotor muscle series to the caudal edge of the protopodite. The only muscle that looks similar to the protopodite remotor muscle of *Penaeus* is the exopodite reductor muscle of *Pandalus*, a muscle not found in *Penaeus*.

DORSAL UROPOD ROTATOR MUSCLES

FIGURES 70, 72, 73, 74

Due to the presence of transverse fasciae dividing them, three dorsal uropod rotator muscles are

found in each half of the sixth abdominal segment of *Penaeus* (fig. 74, A). These large muscles originate over much of the dorsomedial and lateral areas of the sixth abdominal tergum to the margin of the external arm of the anterior oblique muscle (fig. 73, B). The dorsal uropod rotators pass ventrocaudally to insert upon three branches of a large apodemal tendon shared with the telson flexor muscles. The tendon arises from the ventral rim of the protopodite at its junction with the sixth abdominal segment. On contraction, the uropod rotators turn the lateral side of the protopodite downward, thereby maintaining the uropods in the most advantageous position for drawing the animal backwards through the water. The dorsal uropod rotator muscles of *Penaeus* appear to be homologous with those of *Astacus* and *Pandalus*.

VENTRAL UROPOD ROTATOR MUSCLE

FIGURES 71, 72

The small ventral uropod rotator muscle originates by a broad apodeme attached to the posteroventral sternum of the sixth abdominal segment (fig. 71). The muscle runs caudally and laterally to insert by a strong tendon into the medial edge of the protopodite. Contractions of the muscle rotate the protopodite about its long axis, thus bringing the lateral edge of the protopodite and exopodite downwards. The ventral uropod rotator muscle of *Penaeus* is not shown in the studies of *Pandalus*. A ventral uropod rotator is indicated by Schmidt in *Astacus*. The muscle in the crawfish is much larger than its counterpart in *Penaeus*.

PROTOPODITE ROTATOR MUSCLE

FIGURE 74

The protopodite rotator muscle is revealed by the removal of the thick exopodite abductor muscle located in the lateral part of the protopodite (fig. 73, B). The muscle takes origin in the same connective tissue providing insertions for the uropod remotors and the origin of the protopodite remotor muscle. The protopodite rotator passes laterally and ventrally to an area of insertion on the ventrolateral surface of the protopodite. The counterpart of the muscle in *Astacus* and *Pandalus* is not clear.

UROPOD STATOR MUSCLES

FIGURES 73, 74

The uropod stator muscles are 2 or 3 small muscles whose function appears to be the retention of the exopodite and endopodite in position with respect to one another. The affinities of these muscles in other crustaceans are not known.

EXOPODITE ABDUCTOR MUSCLES

FIGURES 70, 71, 73

The uropod of *Penaeus* contains 4 exopodite abductor muscles, all varying widely in shape and size. The anteriormost is a short, strong muscle occupying the lateral part of the protopodite (fig. 73, B). The muscle originates in the anterior curvature of the protopodite and inserts on the lateral margin of the exopodite. The muscle turns the exopodite laterally about its dorsoventral condyles. Also intrinsic to the protopodite are two smaller exopodite abductor muscles inserting in common with the first abductor. The small abductors lie beneath the protopodite remotor muscle and originate at two points on the ventromedial surface of the protopodite. The fourth and caudal exopodite abductor muscle, best seen in dorsal view (fig. 70), runs along the lateral margin of the exopodite. The muscle originates distally and inserts on a lateral apodeme of the protopodite. The function of the exopodite abductors is to spread the tail fan.

The first and fourth exopodite abductor muscles of *Penaeus* are homologous with the dorsal and lateral exopodite abductors of *Astacus* and with the lateral abductor muscle of *Pandalus*. The relationships of the two small abductors in *Penaeus* are not clear. The small ventral uropod exopodite abductor muscle shown by Schmidt in *Astacus* does not appear in *Penaeus*.

EXOPODITE ADDUCTOR MUSCLES

FIGURES 70, 71, 73, 74

The uropod exopodite of *Penaeus* is operated by three exopodite adductor muscles. The first exopodite adductor is rostral to the other two. It is a short, strong, twisted muscle originating on the posterodorsal surface of the protopodite (fig. 70). The first exopodite adductor passes ventrocaudally through other muscles to an area of insertion on the anteroventral surface of the

exopodite (fig. 71). The second exopodite adductor muscle is the longest uropod muscle. The structure lies in the midline of the exopodite, inserting on the posterodorsal margin of the protopodite beneath the first adductor. The muscle takes origin in the distal region of the exopodite and on a little fascia dividing the main body of the muscle from a small, lenticular muscle (fig. 70). This little distal muscle is thought to be the vestige of a muscle operating the distal exopodite joint.

The third exopodite adductor lies along the medial margin of the exopodite. The muscle is made up of short fibers attaching to a long apodeme running through the middle of the structure. The apodeme arises from the posterodorsal margin of the protopodite. Contractions of the exopodite adductors turn the exopodite towards the midline of the animal, opposing the abductors.

The first exopodite adductor muscle of *Penaeus* appears in *Pandalus* as the anterior exopodite adductor. The second exopodite adductor found in the white shrimp is not represented in either *Pandalus* or *Astacus*. The third adductor of *Penaeus* is fully homologous with the muscle known as the posterior exopodite adductor in *Pandalus* and the uropod exopodite adductor muscle in the European crawfish.

EXOPODITE PRODUCTOR MUSCLE

FIGURES 71, 72, 74

The exopodite productor muscle of *Penaeus* is a broad, flat structure occupying the ventral part of the protopodite (figs. 71, 72). The productor originates in the heavy connective tissue of the large apodeme common to the dorsal uropod rotator muscles and the telson flexor muscles. The productor muscle passes caudad to a broad insertion on the ventral rim of the exopodite. The exopodite productor muscle is represented in the tail fan of *Pandalus*, but its relationships in the crawfish are not known. No reductor of the exopodite has been found in the tail fan of the white shrimp.

ENDOPODITE ADDUCTOR MUSCLE

FIGURES 70, 71, 73, 74

Originating in the distomedial part of the endopodite, the endopodite adductor muscle runs proximally in the endopodite to its point of in-

sersion on the ventromedial edge of the protopodite. The muscle turns the endopodite toward the midline, with the result that the endopodites are closed. The endopodite adductor of *Penaeus* is probably homologous with the same muscle in the tail fan of *Pandalus* and *Astacus*.

ANTERIOR TELSON FLEXOR MUSCLE

FIGURES 70, 72, 73, 74

The anterior telson flexor muscle originates over most of the anterodorsal portion of the telson (fig. 70). The muscle passes directly ventrad to insert on the apodeme shared by the dorsal rotators and the telson flexors. The muscle has almost exact counterparts in the telson of *Pandalus* and *Astacus*. The anterior telson flexor muscle pulls the telson downward upon the uropods in the vertical plane.

POSTERIOR TELSON FLEXOR MUSCLE

FIGURES 70, 71, 72, 73, 74

The posterior telson flexor muscle is much larger than the anterior telson flexor. The posterior telson flexor originates over all of the dorsal and lateral tergum of the telson not taken up by the small anterior flexor. The muscle fibers run anteroventrally, narrowing to their insertion on the common apodemal material to which the dorsal rotators and anterior telson flexor are attached (fig. 74, A). The posterior flexor reinforces the action of the anterior telson flexor in bringing the telson ventrad. The posterior telson flexor muscle of *Penaeus* is represented in similar form in *Astacus* and *Pandalus*.

VENTRAL TELSON FLEXOR MUSCLE

FIGURE 74, B

The ventral telson flexor muscle is located within the sixth abdominal segment dorsal to the anterior oblique muscle apodemes. The muscle occupies a scoop-shaped depression in the postero-dorsal part of anterior oblique muscle 6 (fig. 74, B) where it takes its origin. The muscle passes directly caudad to insert on an apodeme arising from the anterior margin of the telson sternum. The muscle turns the telson ventrad in the vertical plane. A counterpart of the ventral telson flexor muscle of *Penaeus* is found in *Pandalus*. The muscle is missing in *Astacus*.

ANAL COMPRESSOR MUSCLES

FIGURES 71, 72

The anal compressor muscles lie ventral to the rectum and alongside of it and insert into the lateral side of the anal opening. Contractions of the muscles flatten the sides of the anal opening in the long axis and thereby tend to close it. A similar muscle is found in *Astacus*, but not, according to Berkeley, in *Pandalus*.

ANAL DILATATOR MUSCLE

FIGURE 72

The anal dilatator muscle runs at right angles to the anal compressor and slightly above it (fig. 72, A), connecting into the lateral tergum of the telson by a tendon. Contractions of the dilatator open the anal aperture. The same muscle is found in the other crustaceans to which we have made reference.

RECTAL ATTRACTOR MUSCLES

FIGURE 74, B

The rectal attractor muscles are two small muscles of doubtful affinity not described by Schmidt in *Astacus* or Berkeley in *Pandalus*. The muscles lie in the midline of the sixth abdominal segment, between the uropod rotators. Both are attached to the median dorsal edge of the telson. The small, dorsal attractor passes rostrad to a tendinous fascia associated with the uropod rotators. If this muscle were larger it might better be named a telson extensor, but its size argues against this interpretation. The larger, ventral attractor muscle inserts in connective tissue on the dorsal surface of the rectum and undoubtedly draws the rectum caudad in movements associated with defecation.

II. THE NERVOUS SYSTEM

To the comparative morphologist, the nervous system is of fundamental importance. Of all the systems—skeletal, muscle, alimentary, reproductive, etc.—the annulate nervous system has most nearly retained its generalized form during the evolution of the group. That is, the nervous systems of any two annulates are remarkably similar. Nervous systems of Annulata, then, as has been amply shown by Snodgrass (1938), Ferris (1953), and others, are singularly conservative of form in

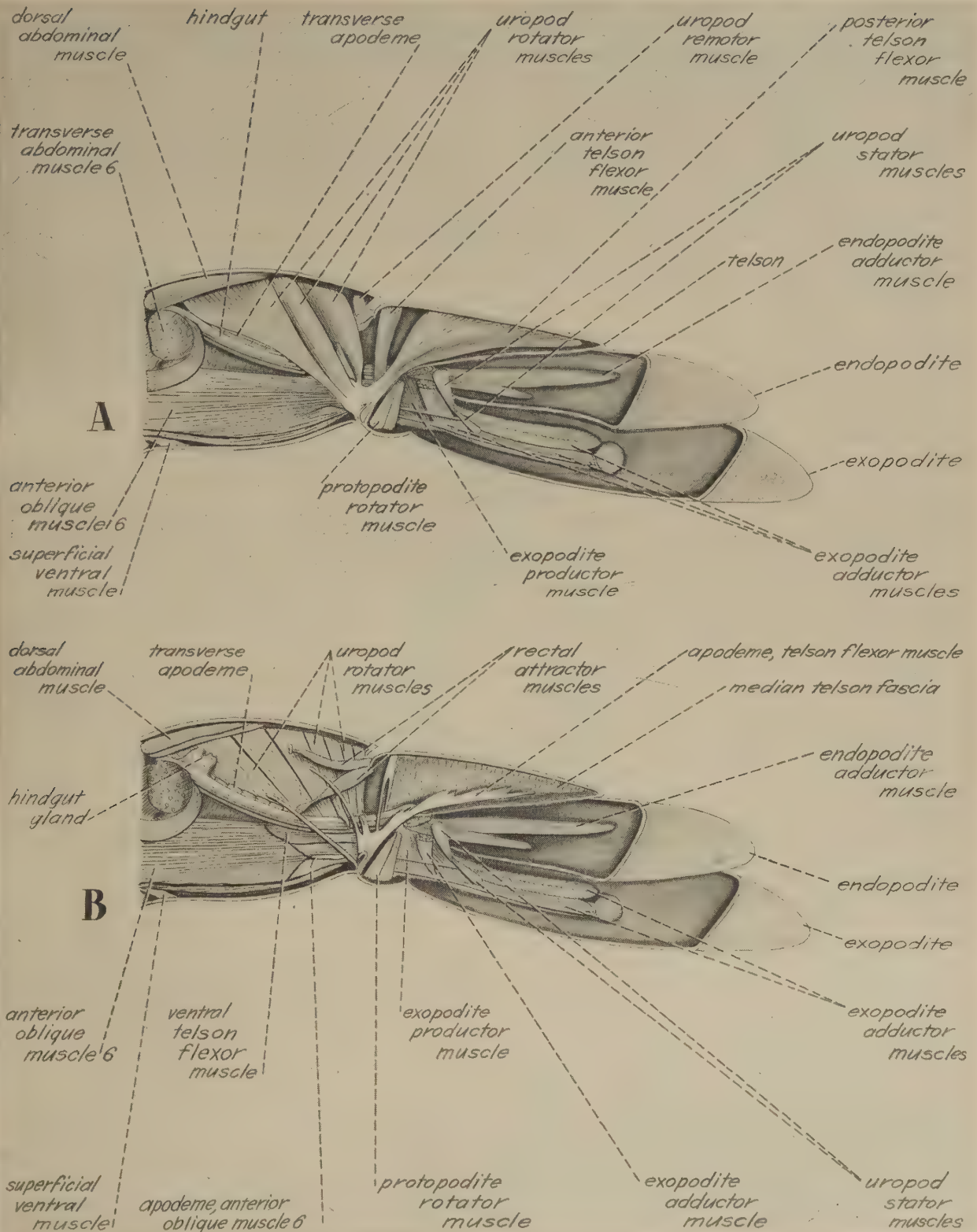


FIGURE 74.—Lateral view of left side of sixth abdominal segment, telson, and uropods. A. Lateral muscles removed. B. Median sagittal view.

time, and for this reason, may be used by the morphologist to bring order out of apparent phylogenetic chaos in the other organic systems. Gaps exist in our knowledge of decapod neuroanatomy, and frequent reference has been made to the lack of detailed information about innervations of muscles and other structures in morphological work.

The basis for an understanding of the history of muscles and other structures is the exact and full understanding of the details of the nervous system. The literature of arthropod morphology does not provide the facts, except in rare instances, nor are all the anatomical facts about the nerves of *Penaus* made available in the present work. The reason for this is the high technical skill and great periods of time required to work out in detail the gross anatomy and histology of a nervous system. The 2 years devoted to the present research on white shrimp have not been sufficient for this purpose.

Despite present shortcomings, a number of important details of the central nervous system of *Penaus* have been worked out. Typical of Annelata, the shrimp nervous system is comprised of a dorsal brain connected to the ganglionated ventral longitudinal nerve cord below the gut by two large tracts. The gut passes between these tracts. In general, the brain or supraesophageal ganglion, receives nerves from the special sense organs of the head and supplies nerves to the muscles operating them. The first ganglion of the ventral nerve cord, usually called the subesophageal ganglion, together with the following metameric ventral ganglia, receive impulses from sensory end organs of the body and appendages and send motor impulses to the muscles moving these structures.

The dorsal brain of annulates is variously composed. The arthropod brain is usually said to consist of an anterior protocerebrum containing the nerve centers of the eyes and other preantennal appendages supposed to have existed in primitive forms. The protocerebrum is joined to a second brain part, the deutocerebrum, an area associated with the antennules, or first antennae. In all insects and most Crustacea, a third brain region, the tritocerebrum, is added to the other parts. The tritocerebrum has traditionally been said to be the nerve center for the antennae (second antennae), although Ferris (1953) presents evidence opposing this view. Classically, the tritocerebrum has been considered the first ganglion of the ventral

nerve cord due to the presence of a large postoral commissure between the lobes of the tritocerebrum. The tritocerebral lobes have thus moved around the mouth to join the dorsal brain in many arthropods.

In some crustaceans, however, including *Penaus setiferus*, the tritocerebral lobes have not become part of the dorsal brain and instead remain ventrally located. Although clearly tritocerebral, the ganglia do not send nerves to the second antennae.

The ventral nerve cord is the fusion product of a "ladder" nervous system, wherein the paired ganglia of each segment have come together at the midline. Longitudinal segmental coalescence has frequently been followed by ganglionic coalescence in the ventral nerve cord with attendant obscuring of primitive metamerism.

In the following treatment, the nerves of the dorsal brain and tritocerebrum will be considered in the first section and those of the ventral nerve cord in the second.

A. Nerves of Supraesophageal Ganglion and Tritocerebrum

The supraesophageal ganglion, or dorsal brain (figs. 75, 76), lies within the head lobe in the dorsal part of the protocephalon. The head lobe is protected dorsally by the broadening base of the rostrum. The dorsal brain is made up of nerve cell bodies and tracts associated with the nerves running out of it.

TEGUMENTAL NERVES

FIGURES 75, 76

The tegumental nerves arise from slightly different points on the anterior face of the supraesophageal ganglion and run directly rostrad to the epidermis of the head lobe. Keim (1915) does not show similar structures in *Astacus*.

OPTIC TRACT

FIGURES 6 TO 10, 75, 76

The optic tract, a part of the brain, rises from the anterolateral region of the supraesophageal ganglion, runs distally in the eyestalk, increasing in diameter, and enters the calathus. Within the calathus, the optic tract enlarges to incorporate the various distal optic ganglia and makes contact with the nerves from the ommatidia (figs. 9, 10).

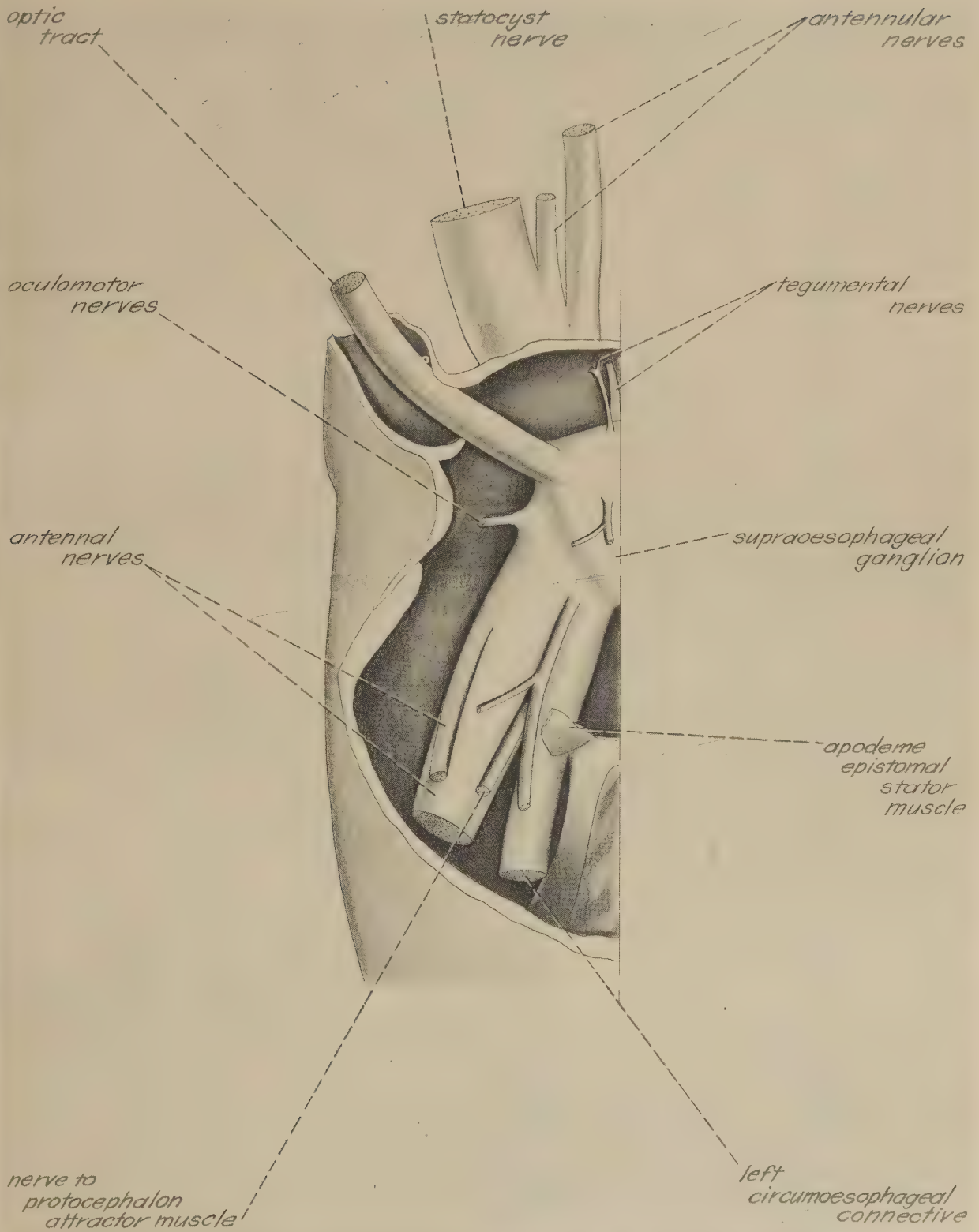


FIGURE 75.—Dorsal view of supraesophageal ganglion. Carapace and dorsal muscles removed.

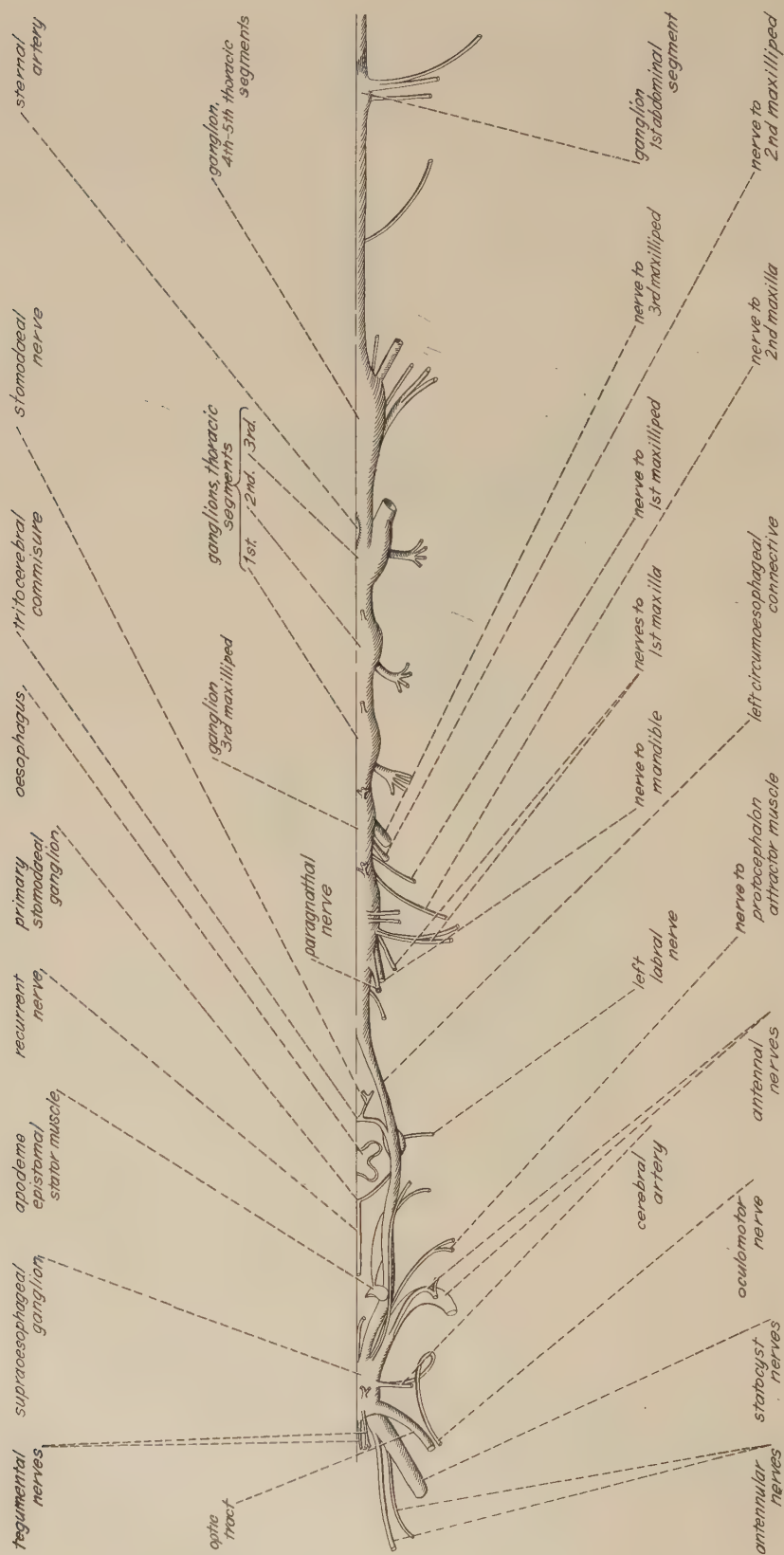


FIGURE 76.—Dorsal view of left half of brain and ventral nerve cord to ganglion of first abdominal segment.

These distal optic ganglia do not show superficially; however, longitudinal sections indicate very clearly the presence of a proximal, medial, and distal ganglion. If the distal optic ganglionic mass is pulled away from the dioptric elements of the eye, the tearing is confined to natural lines of weakness representing a deep concavity. Lining the concavity so produced is the capillary arbor (fig. 9), a structure which will be treated more fully in the section on the circulatory system.

NEUROHORMONAL ELEMENTS

FIGURE 10

Along the lateral side of the optic tract, and embedded in the perineurium in the proximal region of the optic tract, is a small nerve which branches out of the perineurium distal to the basal segment of the eyestalk. This nerve puts out several tiny branches to muscles and then enters a glandlike structure previously identified by Young (1956) as the X-Organ (fig. 10) of Hanström (1948), and which should be referred to as the pars ganglionaris X organi (Carlisle and Passano, 1953) rather than the X-Organ of Hanström.

From the pars ganglionaris X organi, a nerve continues along the optic tract distally to enter another, and larger, glandlike organ termed the "sinus gland" (fig. 10). The sinus gland lies against and sends branches into the optic ganglionic mass at the distal end of the optic tract. A second part of the X-Organ, that described by Hanström, is associated with the anterior eyestalk pore, or sensory pore (fig. 10). Knowles and Carlisle (1956) have proposed the term sensory pore X-Organ for the structure, to distinguish it from the ganglionic part. The identification of the parts of the X-Organ and of the sinus gland was made on doubtful grounds, since no supporting histological or experimental evidence was presented (Young 1956).

Confusion surrounding the identification of the X-Organ may be found in the literature of neurosecretory experiments (Knowles and Carlisle, 1956). Evidently the European and American workers have used the term "X-Organ" for different structures. The reason may lie in a weakness in communications, for the illustrations in some works of this literature are, to say the least, circumscribed (Passano 1953), however impor-

tant the textual material may be to the experimental biologist. Welsh (1941), on the other hand, has taken pains to illustrate clearly his experiments on retinal pigment migrations in *Cambarus bartoni* (Fabricius 1798); unfortunately his identification of the X-Organ appears to be in error.

Keim (1915) in his account of the nerves in *Astacus* does not illustrate the sinus gland or the parts of the X-Organ.

OCULOMOTOR NERVE

FIGURES 8, 9, 10

The oculomotor nerve originates on the lateral side of the dorsal brain, slightly posterior to the optic tract, and, beginning ventrally, describes an almost complete loop around the protocephalon attractor muscles. It proceeds to the dorsolateral region of the protocephalon attractor, between the muscle and the outer epidermis. From the latter position the nerve turns sharply anterior and runs into the eyestalk, giving off branches to various of the eyestalk muscles. The nerve in *Penaeus* is the same as the eye muscle nerve described by Keim (1915) in *Astacus*.

Regrettably, very little can be said of homologies between the nerves serving the eyestalk of the various Crustacea, since so little information exists on the subject. Certainly, optic tracts, oculomotor nerves, and eyestalk neurohormonal elements in *Penaeus*, *Astacus*, and *Cambarus* are likely to be homologous structures. Further anatomical information on the nerves will have to be provided before the comparative morphology of this region of the brain and eyestalk will be in any way a satisfactory story.

ANTENNULAR NERVES

FIGURES 14, 15, 75, 76

The nerves of the antennule pass rostrad from the anteroventral region of the supraesophageal ganglion within a single perineurium. Inside the antennule the single tract divides into three nerves. The largest is the short, flat statocyst nerve which runs anterolaterally and spreads widely on the ventral surface of the first antennular segment beneath the statocyst. This nerve is presumably the sensory nerve of the statocyst. The smaller antennular nerves parallel the statocyst nerve in the proximal region of the anten-

nule, then turn anteriorly and run the length of the antennule including the flagella. This nerve is probably mixed, since it sends off branches to the muscles as well as to the sensory flagella.

Except for the work of Keim (1915) on the nerves of *Astacus*, little information exists on the innervation of the crustacean antennule. The situation in *Penaeus* appears to be roughly comparable to the arrangement of the antennular nerves in *Astacus*. The most striking difference is the size of the nerve innervating the statocyst. That of *Penaeus* is very much larger than the statocyst nerve of *Astacus*.

ANTENNAL NERVES

FIGURES 24, 75, 76

The antennal nerves pass out of the supraesophageal ganglion posteroventrally, from a point laterad of the circumesophageal connective. The antennal nerves proceed ventrocaudally for a short distance and then turn anteriorly to run into the antenna through its large foramen, or into the mass of the antennal muscles. Within the body of the antenna, the nerves give off branches to various muscles and sensory endings in the antenna and scale. One branch enters the antennal flagellum (fig. 24, *B*) and is extensively subdivided. Small groups of neurons are thus split off to each of the many sensory elements in the flagellum (fig. 25).

The antennal nerves of *Penaeus* appear to be similar to those described by Keim in *Astacus*.

NERVE TO PROTOCEPHALON ATTRACTOR MUSCLE

FIGURES 75, 76

This structure comes out of the dorsal brain between the antennal nerves and the circumesophageal connective and passes into the substance of the protocephalon attractor muscle. I have previously offered objections to assigning the protocephalon attractors to the "antennal" segment, on grounds of multiple insertions of the muscle. However, the position of the protocephalon attractor nerve origin on the brain tends to suggest that at least part of the muscle has affinities with the antenna.

LABRAL NERVE

FIGURES 27, 76

Each lateral half of the superficially unpaired labrum is supplied with a labral nerve descending

from the tritocerebral ganglia (illustrated as the labral ganglion in fig. 27). Upon entering the labral sac, the nerve divides into parts which go to many labral muscles, to glands, and to sensory endings in the epidermis. From the work of Schmidt and Keim on the muscles and nerves of *Astacus* one receives the impression that this European crawfish does not possess a labrum, since no muscles or nerves are listed, nor in fact is the labrum mentioned. Henry (1948a) finds that the labral nerve of *Palaemon paucidens* de Haan and other Crustacea arises from the region of the primary stomodaeal ganglion, a stomatogastric ganglion connected to the tritocerebrum.

TRITOCEREBRAL GANGLIA AND NERVES

The tritocerebral ganglia or lobes represent the ganglia of the first ventral segment. As has been previously mentioned, the tritocerebral lobes are reflected into the dorsal brain in many arthropods, but not in *Penaeus*. In the latter the ganglia remain in the primitive, divided condition, connected by a transverse commissure, the tritocerebral (or stomodaeal) commissure (fig. 76), passing below the gut. The tritocerebral commissure sends a small nerve, the stomodaeal nerve, posteriorly from each side of the midline. The experimental workers evidently prefer the somewhat meaningless term, postcommissural nerve, for the structure (Knowles 1953). Keim makes no mention of the postcommissural, stomodaeal nerves in *Astacus*.

Associated with the tritocerebrum are the nerves of the stomodaeal system. So far as this system has been worked out in *Penaeus*, it appears to be simple. A primary stomodaeal ganglion lying on the anterior surface (primitive dorsal surface) of the esophagus is joined to each tritocerebral lobe by a nerve passing round the gut. From the primary stomodaeal ganglion another nerve, the recurrent nerve, passes dorsally along the dorsal surface of the esophagus to the gastric mill where it becomes subdivided. Keim finds a similar situation in *Astacus*. Henry (1948a) describes a secondary stomodaeal ganglion connected independently to the tritocerebrum, and from her work on lower Crustacea we may conclude that the presence of two stomodaeal ganglia is the more typical situation.

Posterior to the tritocerebral ganglia the circumesophageal connectives passing around the gut anastomose to form the ventral nerve cord.

Ventral Nerve Cord

FIGURES 36, 61, 63, 76

The ventral nerve cord of *Penaeus* is a longitudinal series of ganglia interconnected by fused pairs of intersegmental nerve tracts. A certain amount of longitudinal coalescence has taken place in the ventral cord, notably in the anterior, thoracic regions and less so in the abdomen. The ganglia of the gnathal appendages are conspicuously fused.

PARAGNATHAL NERVE

FIGURE 76

The element termed the nerve to paragnath arises from the ventral cord anterodorsad of the mandibular nerve. In *Penaeus* the paragnathal nerve is in no way posterior to the mandibular nerve as shown by Henry (1948a) for various crustaceans. However, whether the nerve is histologically anterior to the nerve to the mandible, as Chaudonneret (1956) holds, is not known. If Chaudonneret is correct in his claim that the paragnatha are anterior to the mandibles, then we are faced with the necessity of explaining the segmental interrelationships suggested by this view. Chaudonneret escapes from the dilemma by homologizing the paragnatha with the insectan superlinguae, a theory that has been advanced in the past (Snodgrass 1935). At the same time, Chaudonneret stoutly defends the morphological independence of the paragnathal nerve from that of the mandible. He thus arrives at a paradox, since the superlinguae arise from the hypopharynx and appear to belong to the mandibular segment (Snodgrass 1935). If, on the other hand, the superlinguae are not mandibular, and their homology with the paragnatha holds good, we must assign the paragnatha to some other segment, perhaps pre-mandibular and almost certainly postoral.

MANDIBULAR NERVE

FIGURES 36, 76

The large nerve to mandible is given off the ventrolateral surface of the anterior ventral nerve ganglion and runs into the heavy muscles of the mandibular segment. A branch of the nerve enters the mandibular palp.

MAXILLARY NERVES

FIGURES 36, 76

The nerves to the first and second maxillae depart from the ventral nerve cord and pass into the maxillae. The nerves to the first maxilla are much smaller than the one to the more active second maxilla. These nerves are very likely mixed since they separate into branches which run to muscles and to sensory end cells in the appendage extremities.

MAXILLIPEDAL NERVES

FIGURES 36, 43 TO 48, 76

The nerves to the first and second maxillipeds leave the anterior ventral nerve ganglion ventrolaterally from its caudal portion and run into the maxillipeds. The nerve to the third maxilliped arises from its own ganglion.

NERVES TO PEREIOPODS

FIGURES 36, 49, 51 TO 53, 56 TO 58

Beginning with the ganglion of the third maxillipeds, the thoracic ganglia are indicated for the most part by independent swellings in each segment. From the ventral surface of each ganglion a large nerve departs into each appendage. Between the ganglion of the third thoracic segment and the anterior portion of the coalesced fourth and fifth ganglia, the two longitudinal nerve tracts have been retained in the primitive divided condition to permit the sternal artery to pass through them. The thick ventral nerve cord of the thorax narrows posterior to the fused ganglia of the fourth and fifth thoracic segments and runs into the abdomen.

ABDOMINAL NERVES

FIGURES 61, 63, 71, 72, 76

The abdominal ganglia of the ventral nerve cord are all substantially similar, except for the terminal ganglion of the sixth abdominal segment. Two nerves arise from each of the first five abdominal ganglia. The anterior one enters the pleopod on that side and the posterior nerve sweeps laterally and dorsally to branch among the abdominal muscles of the segment.

The sixth abdominal ganglion lies in the posteroventral portion of the sixth segment (fig. 72, B). Four pairs of major nerves arise from it.

The first nerve is dorsolateral and runs beneath the apodeme of anterior oblique muscle 6 on its way to the muscles of the sixth abdominal segment. The second nerve serves the uropod exopodite, the third the endopodite, and the fourth, which arises posterodorsally, runs near the anal canal and enters the telson. These nerves are apparently mixed. The exopodite nerve (fig. 72, *B*) is easily observed. After giving off branches to the exopodite muscles, this nerve undergoes extensive subdivision to form a kind of sensory "cauda equina" in the distal regions of the exopodite.

III. CIRCULATORY SYSTEM

The typical blood circulatory system of Crustacea Decapoda is a closed arterial-open venous system. It consists of a heart and arteries by which blood is transported to arterial capillary beds throughout the body. Venous blood returns to the heart through conjoint blood sinuses in the appendages and body into which the capillaries empty. In general, the gills are served by the venous system. Although the circulatory systems of all decapods have marked similarities, the arteries display greater differences in detail than the nervous system or the muscle system. Furthermore, the circulatory elements are variable within the individual animal. The blood vascular system will be treated in two parts, the heart and pericardium and the blood vessels.

A. Heart and Pericardium

FIGURES 79, 80

The pericardium is a thin, contractile membrane surrounding the heart. The membrane is penetrated by passageways by which venous blood may enter the pericardial chamber. Slender muscle bundles inserted on the surface of the pericardium aid in its rhythmic contractions.

The heart of *Penaeus setiferus* is a many-sided structure lying within the muscular pericardium in the posterodorsal region of the carapace. Three pairs of valvular afferent ostia may be seen on the surface of the heart, two pairs dorsally and one pair placed laterally. By means of the valves in these pores, pericardial blood enters the heart during diastole and is prevented from escaping in the systolic beat. Structurally, the heart is more than a tubular expansion, and is, instead, comprised of a system of connected sinusoids.

Microscopical sections indicate that the outer and intersinusoidal walls are made up of muscle fibers interspersed with connective tissue of a type strikingly reminiscent of the wavy elastic fibers in the tunica media of a vertebrate artery.

Although the details of cardiac innervation have not been worked out in the present study of *Penaeus*, the work of Alexandrowicz (1932) shows that considerable uniformity in the heart nerves of decapod crustaceans exists. Alexandrowicz has found that the decapod heart is well supplied with nerves. Restricted to the internal heart structures is a local system of heart neurons interspersed among the muscles of the sinusoidal septa. Alexandrowicz considers the local system inherently automatic.

The heart neurons are connected to the central nervous system by a pair of dorsal cardiac nerves which arise from the subesophageal ganglion. The dorsal nerves Alexandrowicz believes to be regulatory and he subdivides them functionally into two groups of inhibitory and one of accelerator fibers. Distinct from the heart neurons and dorsal nerves is a third system serving the muscles of the pericardium and heart valves. The pericardial nerves are ventral and the ostial valve nerves are situated dorsally, and the two are interconnected with one another. Inexplicably, the pericardium-ostial valve system is not tied to either the heart neurons or the dorsal cardiac nerves. Consequently, Alexandrowicz states, the pericardium has its own rhythm.

B. Blood Vessels of the Body

LATERAL ANTERIOR ARTERY

FIGURES 79, 80

The narrow, apical end of the heart points rostrally. From this point in the midline a very small vessel extends a short distance anteriorly (fig. 80). This little artery may be the vestige of the ophthalmic artery (Huxley 1906), or the median aorta (Baumann 1919) found in *Astacus* and other crustaceans. In the opinion of Professor Mayrat (letter, May 3, 1957), ophthalmic (or median aortic) vascular function has apparently been taken over by a pair of lateral anterior arteries (figs. 79, 80) arising from the anterior end of the heart, laterad of the midline.

As the lateral anterior arteries run forward, they give off a number of branches, the first of which leaves just anterior to the heart. The

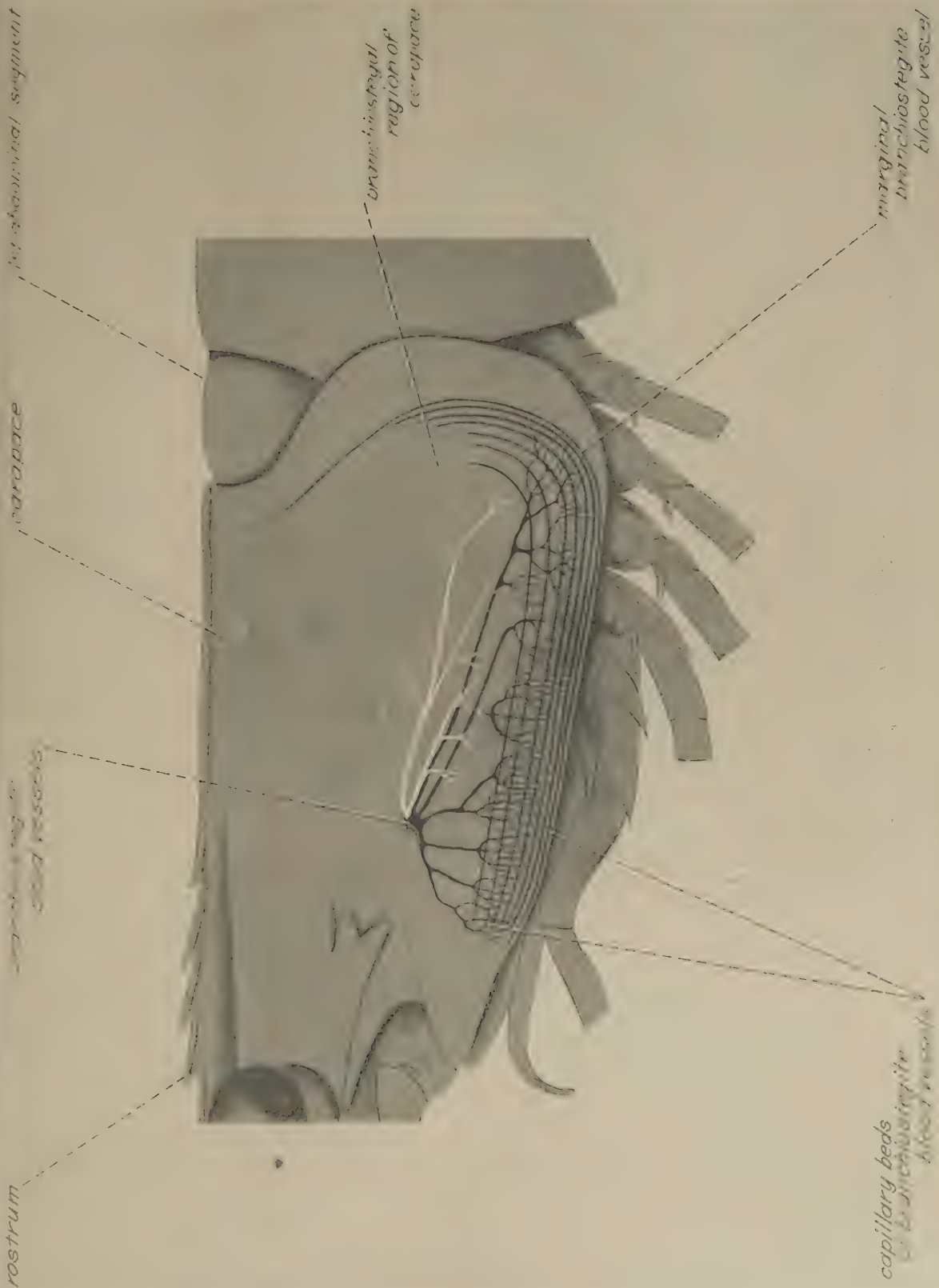


FIGURE 77.—Lateral view of gnathothorax showing blood vessels of branchiostegite epithelium.

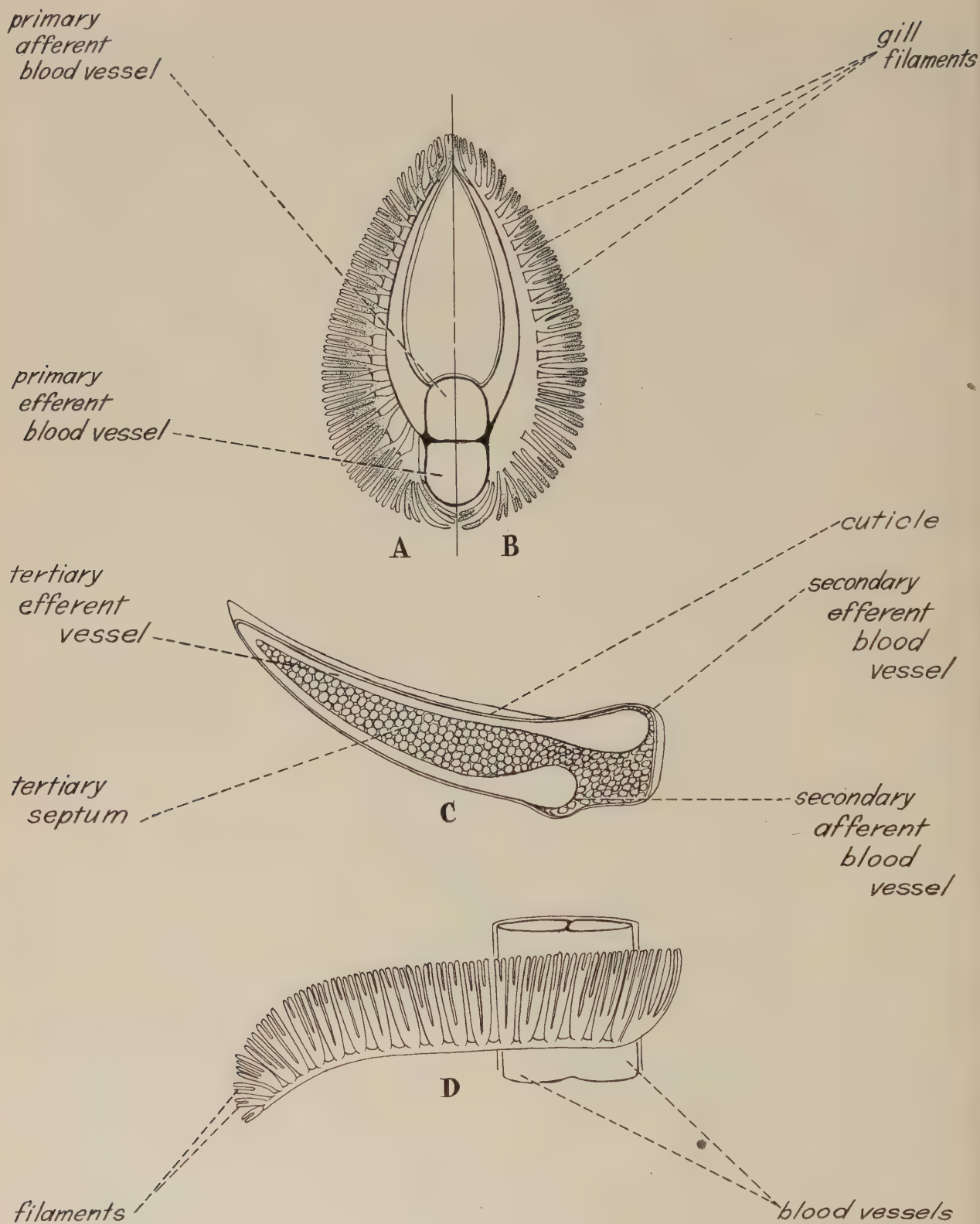


FIGURE 78.—Blood circulation of gills. A. Dorsal, and B. ventral views of single gill lamina. C. Lateral view of single gill filament showing circulation. D. Lateral view of single gill lamina.

branch is the subgastric artery (fig. 79) and it runs around the posterior end of the gastric mill to a curious glandlike organ lying beneath the gastric mill. The subgastric artery is similar to the internal ramus of the lateral anterior artery described by Baumann (1919) in *Astacus*. Rostrad of the subgastric branch, the lateral anterior artery puts out the dorsal gastric artery (figs. 79, 80) which serves the dorsal regions of the gastric mill. The gastric artery, too, divides into at least three major divisions (fig. 80), one running some distance rostrally, another caudad into a glandlike structure of unknown function (see fig. 79, dorsal gland, and a third passing around the gastric mill ventrally.

The lateral anterior artery now turns to the midline to join its counterpart from the opposite side. The optic artery continues anteriorly to the eyestalk from this turn, giving off a branch, the cerebral artery, to the supraesophageal ganglion. At the midline, the two lateral anterior arteries form a median longitudinal vessel. The anterior portion extends into the rostrum while the posterior part, the recurrent artery, runs caudad to the dorsal surface of the gastric mill. At the junction of the optic artery with the oculomotor, a small branch is given off which runs beneath the optic tract and thence to the midline to meet its opposite number from the other side (fig. 80) slightly rostrad of the dorsal brain. No evidence of the frontal heart so prominent in this region in other decapods (Baumann 1917) has been found in *Penaeus*.

The next major branch of the lateral anterior artery anterior to the gastric branch is the mandibular artery. Upon leaving the lateral anterior artery this branch passes between the lateral mandibular condyle and the origin of the lateral anterior thoracic muscle 1 (fig. 80) into the substance of the mandibular muscles where the vessel undergoes further subdivision.

Rostrad of the mandibular artery, the lateral anterior artery gives off the large antennal artery whose subdivisions include a branch to the antennule. The major portion of the antennal artery leads to the muscles and other organs of the antenna.

HEPATIC ARTERY

Running from the heart is another pair of arteries, the hepatic arteries (fig. 79). These project from the anteroventral surface of the

heart and run into the hepatopancreas. They are difficult to follow within the gland in preserved material because of postmortem effects.

DORSAL ABDOMINAL ARTERY

The largest artery passing from the broad, posterior end of the heart is the unpaired dorsal abdominal artery (figs. 61, 62, 79, 80). This vessel runs the length of the abdomen between the dorsal abdominal muscles and the gut. As it proceeds caudad, the dorsal abdominal artery gives off pairs of segmental arteries at each segmental junction (fig. 62). In addition, numerous small vessels from the dorsal abdominal artery irrigate the gut throughout its length. Posteriorly, the dorsal abdominal artery bifurcates around the hindgut gland (fig. 82) and runs alongside the rectum as a paired vessel. In the neighborhood of the rectum the paired vessels turn ventrad, after giving several branches to various structures of the region, including the telson, and anastomose beneath the posterior part of the rectum. Having fused, the vessel passes caudad of the ganglion of the sixth abdominal segment and then turns anteriorly as the posterior subneural artery (figs. 81, 82). So far as can be determined, the posterior subneural artery extends no further rostrad than the anteroventral part of the sixth abdominal segment.

On the dorsal surface, the segmental arteries give off branches adjacent to the dorsal abdominal artery (fig. 62). These branches enter the large ventral abdominal muscles. The remainder of the segmental arteries now pass ventrally along the lateral surface of the abdomen at the intersegmental lines. A substantial branch splits off internally and runs across the external arm of the anterior oblique muscle in each segment (fig. 60). The segmental artery (see lateral artery, fig. 81) continues ventrad between the abdominal muscles and the promoter-remotor muscle mass of the pleopod and sends a branch, the pleopod artery, into the pleopod (fig. 81). A second branch proceeds into the substance of the abdominal ganglion. This branch also divides again to form the postganglionic loop (fig. 81) which provides a vascular connection between the segmental arteries of both sides.

STERNAL ARTERY

In general, the ventral vascular supply is less distinctly laid out than are the dorsal circulatory

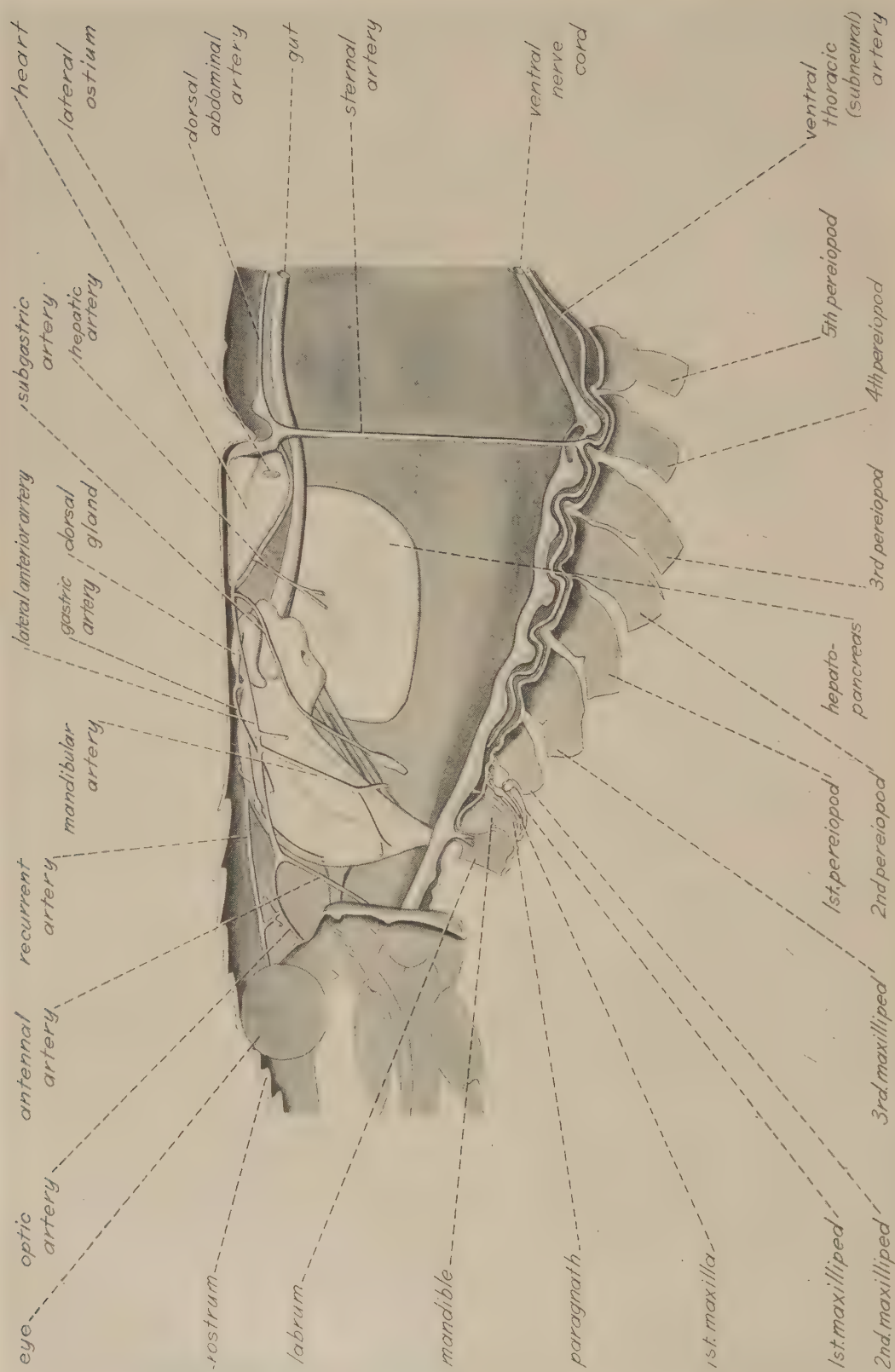


FIGURE 79.—Lateral view of gnathothorax showing circulatory elements.

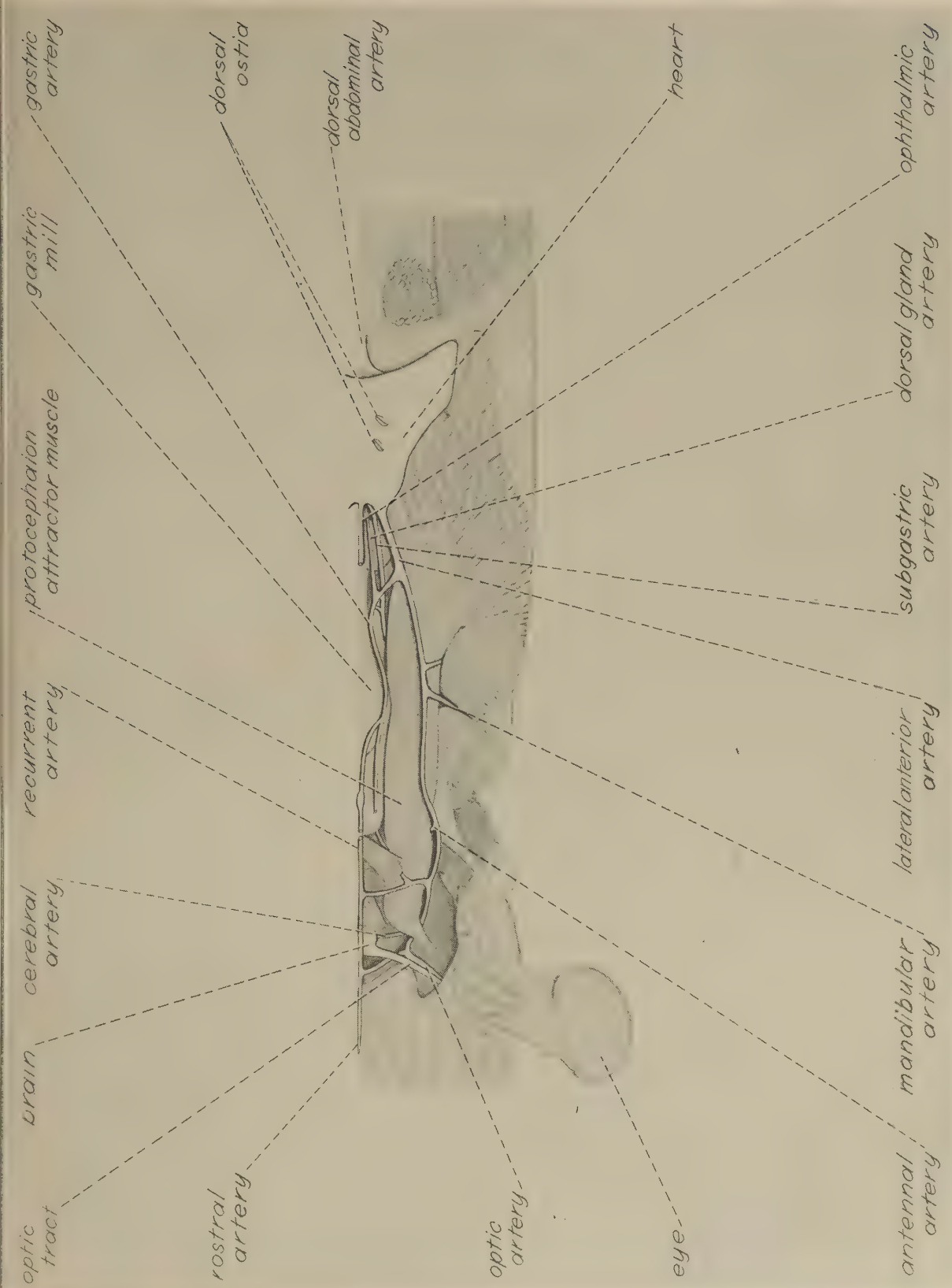


FIGURE 80.—Dorsal view of gnathothorax showing circulatory elements.

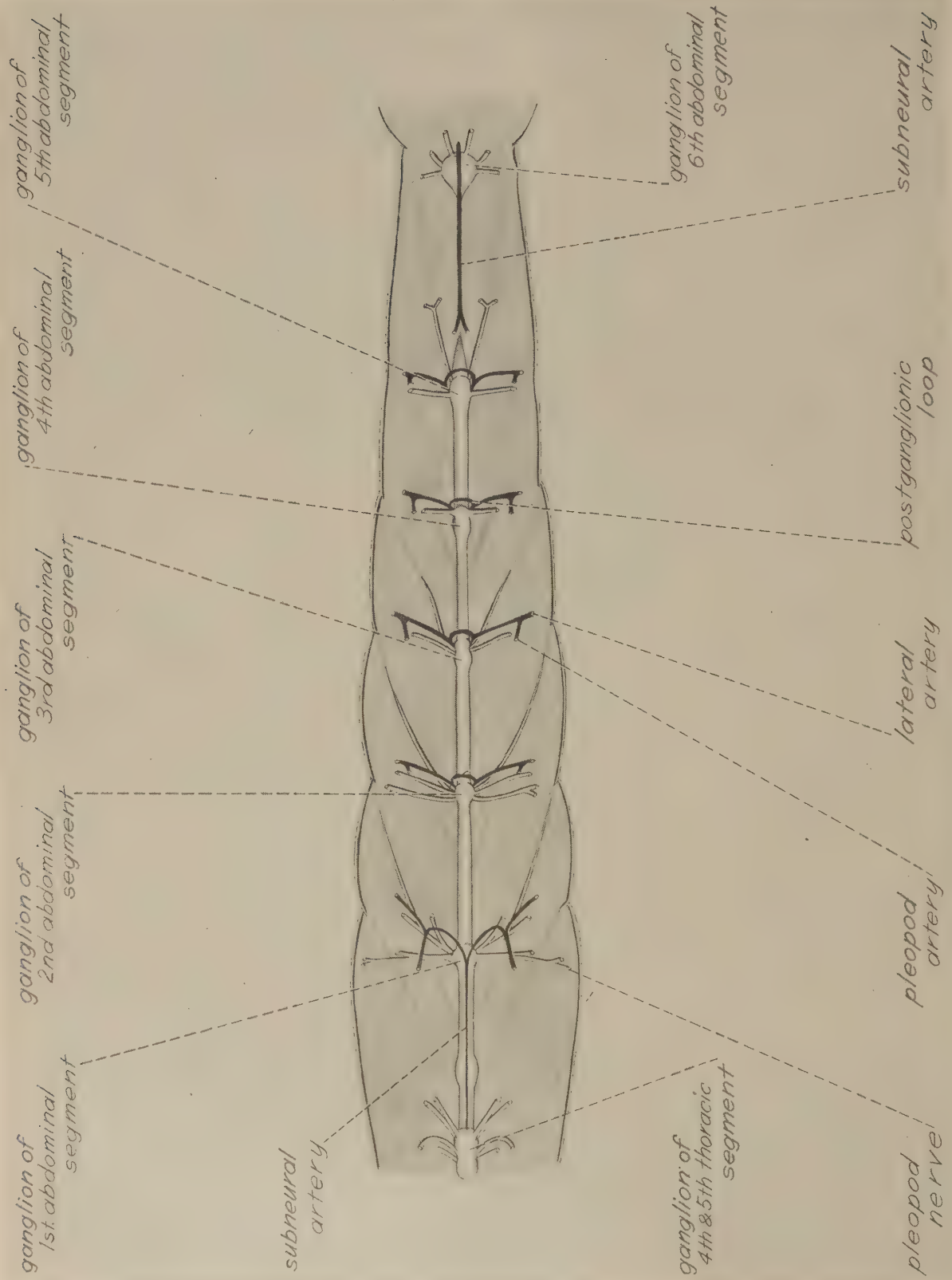


FIGURE 81.—Ventral view of abdomen showing neural and circulatory elements.

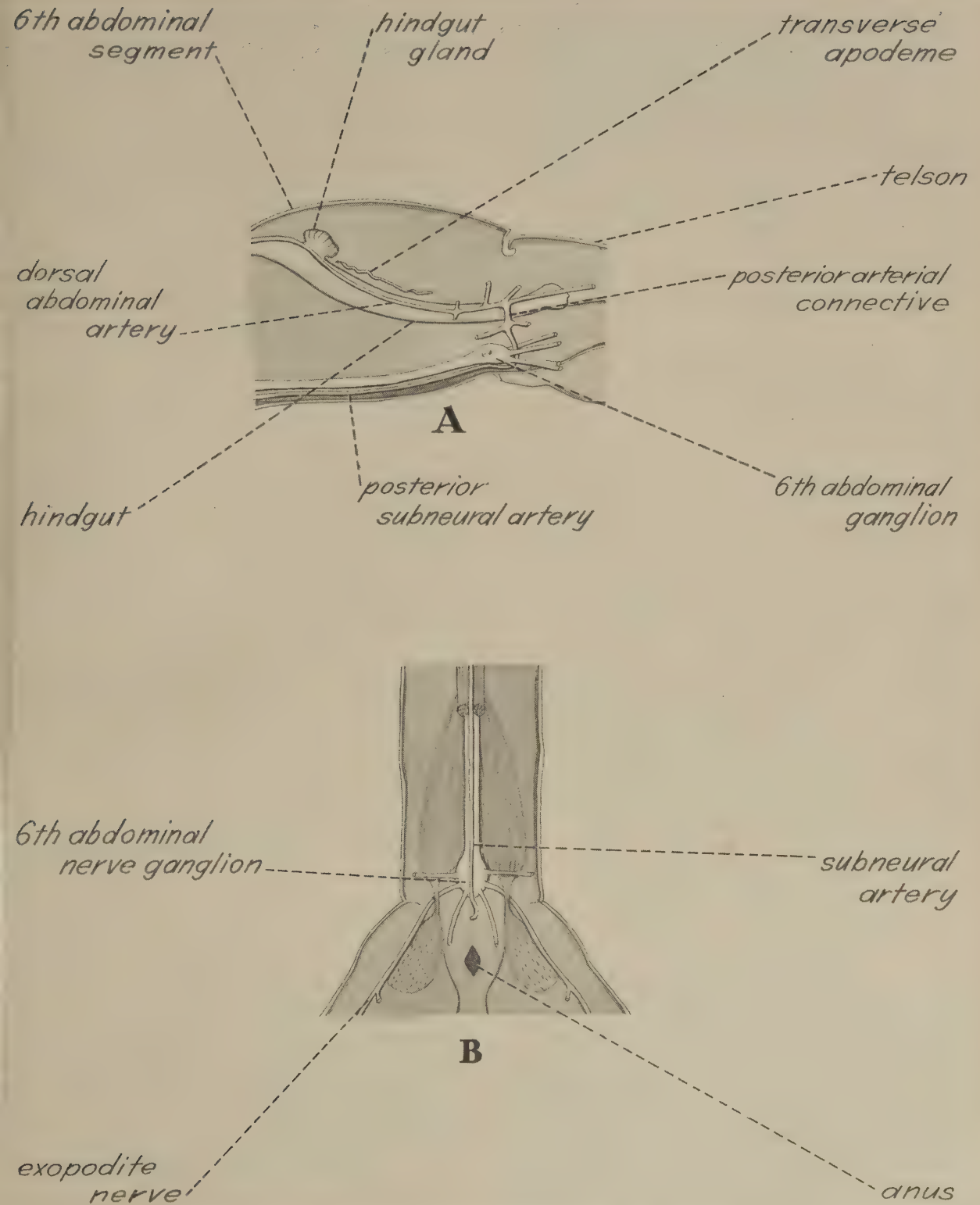


FIGURE 82.—Neural, circulatory, and digestive structures in posterior abdominal segments. A. Lateral view of sixth abdominal segment and part of telson. B. Ventral view of sixth abdominal segment, and parts of uropods and telson.

elements. The principal vessel supplying blood to the ventral regions is the sternal artery (fig. 79) which in some Malacostraca arises from the posteroventral surface of the heart, but in *Penaeus* is a branch of the very rostral portion of the dorsal abdominal artery. The sternal artery descends directly ventrad past the gut and through the ventral nerve cord between the ganglia of the third and fourth ambulatory legs (figs. 76, 79). Here, the artery divides into an anterior portion running forward beneath the nerve cord in the gnathothorax and a posterior piece which eventually enters the abdomen (figs. 36, 79, 81).

VENTRAL THORACIC ARTERY

The ventral thoracic (subneural) artery (figs. 36, 79) supplies blood to the gnathothoracic appendages (fig. 36) as it passes anteriorly. Slightly rostrad of the nerve to the second maxilla, the ventral thoracic artery bifurcates. The two branches turn sharply laterad, sending branches into the nerve cord in the region of the mandibular nerve and also into the anterior structures of the ventral thorax. No connection between the anterior branches of the ventral thoracic artery and the lateral anterior artery has been found in *Penaeus*.

The portion of the ventral thoracic artery posterior to the sternal junction gives off large branches to the fourth and fifth walking legs and the anterior ends of the anterior oblique muscles (fig. 36). A small branch continues subneurally to join other vessels beneath the first abdominal ganglion.

B. Appendicular Blood Vessels

EYESTALK VASCULAR SUPPLY

FIGURES 7 TO 10

Blood is pumped to the eyestalk through the optic branch of the lateral anterior artery. Once in the eyestalk the vessel, now known as the oculomotor artery, runs medially along the optic tract and divides into several branches at the distal end of the eyestalk. The most proximal branch bifurcates on the dorsal surface of the optic tract (figs. 7, 8), sending a short vessel to and apparently through a small gland on the optic tract here designated as the X-Organ of Hanström (1948) and about which something has been said in the section on the nervous system. A small part of the arterial branch to the gland continues

proximally along the dorsal surface of the optic tract and has not been traced beyond the connective tissue of the basal segment of the eyestalk. The larger part of the proximal oculomotor branch runs distally into the distal optic ganglionic mass (figs. 7, 8).

Distally, the oculomotor artery divides into two large branches, one of which (figs. 8, 9) carries blood into a highly branched, dendritic structure embedded deeply among the optic ganglion cells (fig. 9). The organ has been named the capillary arbor (Young 1956), since it appears to distribute blood to ganglionic cells. Nothing similar has been found in the literature of the arthropod eye. However, Professor Mayrat reminds me that he has described (Mayrat 1956) a similar structure in *Praunus flexuosus* (O. F. Müller).

The other, and most-distal oculomotor branch, repeatedly divides to form a vascular plexus on the medial surface of the eyestalk, just beneath a pore to the exterior (figs. 9, 10). The pore is designated as the anterior eyestalk pore. This structure was first described by Hanström (1948) as the eye papilla or sensory papilla and is shown by him as having some kind of structural and spatial relationships with the X-Organ in several crustaceans. Mayrat (1956) also finds the X-Organ near the eyestalk pore in *Praunus*. The function of the anterior eyestalk pore is unknown.

ANTENNULAR BLOOD SUPPLY

FIGURE 14

The blood vascular supply to the antennule is comprised of a large branch of the antennal artery, which is in turn a branch of the lateral anterior artery. The antennal branch enters the antennule in the mesial region near the point of entry of the antennular nerves. The antennular artery courses anteriorly along the mesial side of the antennule giving off small branches to muscles and other structures and finally dividing to supply the two flagella. The arterial branches are seen to subdivide to form capillary beds about the antennule.

ANTENNAL BLOOD SUPPLY

FIGURES 21, 79, 80

The antenna receives its blood from the antennal branch of the lateral anterior artery. The vessel gives off branches in the region of the large

antennal muscles and enters the antenna proper with the antennal nerves. Here it divides to supply the antennal scale and endopodite segments.

VENTRAL APPENDAGE BLOOD SUPPLY

The ventrally located appendages of the head and gnathothorax all appear to receive blood from branches of the ventral thoracic artery. Those of the abdomen, as has already been mentioned, receive fluid from the segmental branches of the dorsal abdominal artery.

D. Venous System

Arterial circulation consists of a closed afferent system subdivided into capillary beds in muscles and other organs. Venous return to the heart appears to be carried out in an open system, through sinuses in the appendages which empty into larger sinuses in the hemocoel. In general, the appendicular sinusoids are found on either one margin or the other of the appendage, rather than in the middle. The sinuses of the body which drain the venous blood from the appendages are located in particular places in the body. The sinus into which the eyestalks empty occupies the anterodorsal regions of the protocephalon. This sinus runs into a larger one anterior to the gastric mill into which blood from the other protocephalon appendages drain. The largest thoracic sinus lies along the sternum. It receives blood from the dorsal sinuses and the gnathothoracic appendages and supplies blood to the respiratory organs. From the sternal sinus the venous blood passes dorso-laterally to the pericardium.

In the abdomen, the sinus system is relatively simple. The dorsal regions are drained by a sinus or set of sinuses between the dorsal and ventral abdominal arteries. These appear to run directly rostrad into the heart. On the ventral surface segmentally arranged, conjoint sinuses pass venous blood rostrally to the sternal sinus of the thorax and thence to the heart. The sternal sinuses of the abdomen extend some distance dorsolaterally in conjunction with the sinuses of the pleonic appendages.

E. Respiratory System

The subject of gill formulae and arrangement in *Penaeus* and other crustaceans has been dis-

cussed previously in the general section on the gnathothorax. The details of the respiratory system will be taken up at this point, since the organs concerned with gas exchange are intimately associated with the circulation of body fluids.

The gills of *Penaeus* are of the dendrobranchiate type (Calman 1909). This type of gill is comprised of a primary axis or rachis from which pairs of secondary structures bearing gill filaments arise at right angles. The secondary structures and gill filaments thus appear to be laminar units. Proceeding distally, each succeeding layer of secondary structures and filaments nests within the preceding layer.

For the details of gill circulation we are indebted to Prof. Jerome E. Stein, Texas Agriculture and Mechanical College, Galveston, Tex. He finds that blood enters the primary rachis of the gill by means of a primary afferent blood vessel (fig. 78, *A, B*) which is separated from the primary efferent blood vessel by a longitudinal septum. Primary afferent blood is directed into the secondary afferent blood vessels (fig. 78, *C*) by a secondary septum. Blood now passes out into the secondary structures to the gill filaments, into which it runs due to the arrangement of a tertiary septum dividing the tertiary afferent and efferent blood vessels in each filament. Blood thus flows around the tip of the filament and returns to the sternal sinus through the efferent vessels of the gill. The gill filament cuticle and epidermis appear to be very thin to allow for gas exchange.

An important accessory organ of respiration is the branchiostegite. The inner cuticle adjacent to the gills is thin and the underlying epidermal layers are heavily vascularized. Specimens preserved in Zenker's fluid display with great clarity the branchiostegal vessels (fig. 77) and fine nerves. The vessels appear to be of two types. One set represents the subdivisions of a large vessel seen at the dorsal margin of the branchiostegite a short distance caudad of the hepatic spine. The capillaries of the vessel approach the periphery of the branchiostegite at right angles to its margin. A second set of vessels run parallel to the margin of the branchiostegite (fig. 77), and are reminiscent of growth rings, since the larger shrimps in general have more rows of these vessels than do the younger animals. Whether the marginal vessels reflect the age of the white shrimp in terms of the number of molts is not known.

IV. DIGESTIVE SYSTEM

The digestive system is made up of those structures concerned with the enzymatic and mechanical reduction of food particles, the absorption of their products, and the evacuation of particulate wastes. The digestive systems of Crustacea Decapoda are all relatively uniform, being comprised of a stomodaeum, or foregut, which includes the esophagus and gastric mill, and probably the hepatopancreas; the midgut, or mesenteron, which in *Penaeus* appears to run from the gastric mill to the hindgut gland on the anterior end of the sixth abdominal segment; and the hindgut, or proctodaeum, which includes the rectal gland, rectum, and anal canal. The alimentary structures will be treated according to these three regions. The subdivisions of the gut are based upon the presence of a cuticular lining of the fore- and hindgut. That section not lined with cuticle is supposed to be the midgut. This criterion is here adopted for whatever it is worth.

A. Foregut

ESOPHAGUS

FIGURES 83 TO 85

The esophagus is a vertical tube which receives food particles thrust into the mouth by the labrum and postoral mouthparts. The tube is lined with cuticle, yet is so constructed that extensive contraction and expansion is possible. A cross-sectional view of the esophagus (fig. 85, *A*) indicates how these movements come about. The cuticular lining is folded inwards between the corners. Esophageal constrictor muscles (fig. 83), short fibers running transversely, draw the corners together and thereby close the lumen of the tube. Expansion of the lumen is accomplished by anterior and posterior esophageal dilatator muscles (fig. 83) which are inserted into the inward folds. Their contractions pull the folds outwards. The esophagus is constricted at its junction with the gastric mill preventing backflow of the food particles from the latter.

GASTRIC MILL

FIGURES 83 TO 85

The gastric mill is a bulbous, folded structure occupying the dorsomedial region of the thorax. As decapod gastric mill structure goes, that of

Penaeus is relatively simple. Its surface is invested with thin sheets of constrictor muscles. Numerous dilatator muscles attach about the outside of the structure. The gastric mill, like the esophagus, is lined with cuticle, but here the latter substance is reflected into deep folds and toothlike structures (fig. 84) for the grinding of food. Transverse sections through the mill (fig. 85) indicate that the theoretical tubular form of the gastric mill is much modified. In general, the walls have been thrown into three pairs of folds. The uppermost fold (fig. 85, *B*), which disappears caudally (figs. 85, *C*₁, et seq.), serves no other purpose than the expansion and contraction of the gastric mill in response to feeding and digestion.

The middle fold, in contrast, constitutes a movable ridge or shoulder upon which a row of increasingly large teeth, the lateral denticles (fig. 84; 85, *B*, *C*₁, *C*₂) are borne. These apparently play an important role in the reduction in size of the gastric mill contents. The lower, or ventral, fold (fig. 85) is separated from the dorsal folds by an extensive lateral fold. The inner margin of the ventral fold is setose. The setae tend to set apart a ventral channel, or channels, running the length of the gastric mill. This ventral channel broadens posteriorly (fig. 85, *D*) and in the posterior end of the gastric mill the lateral ends of the channel bend ventrad (fig. 85, *E*). Here, the channel enters a filter (fig. 84) which is made up of the closely appressed, setose walls of the ventral channel. The setae are arranged in regular rows. According to Calman (1909), the filter is very constant in the penaeids. At its posterior end, the filter has a pair of ducts through which the digestive fluids of the hepatopancreas enter the gastric mill (fig. 85, *E*). Presumably the filter prevents the entry of particulate matter into the main collecting ducts of the hepatopancreas.

The principal device for chewing in the gastric mill is the median tooth, situated in the midline at the dorsalmost point of the mill (figs. 84; 85, *C*₂, *D*). The median tooth is a heavily sclerotized structure, a proliferation of the gastric mill wall. The tooth rests on articular rods which enable its operating muscles to move it some distance into and out of the lumen of the mill and with considerable force.

Posterior to the regions of the median tooth and beyond the filter the terminal lappets of the gastric mill project into the lumen of the midgut

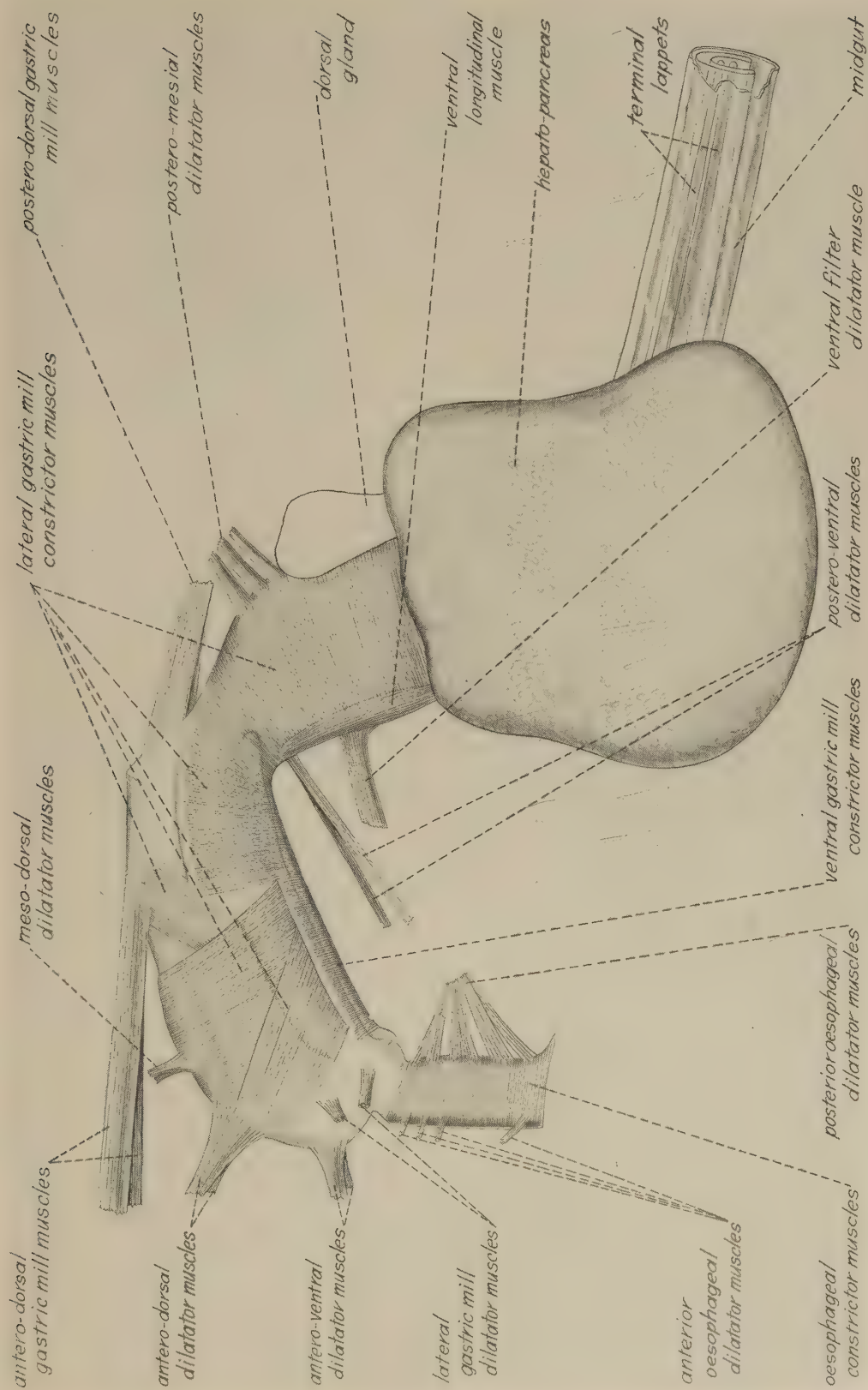


FIGURE 83.—Lateral view of left side of gastric mill and hepatopancreas.

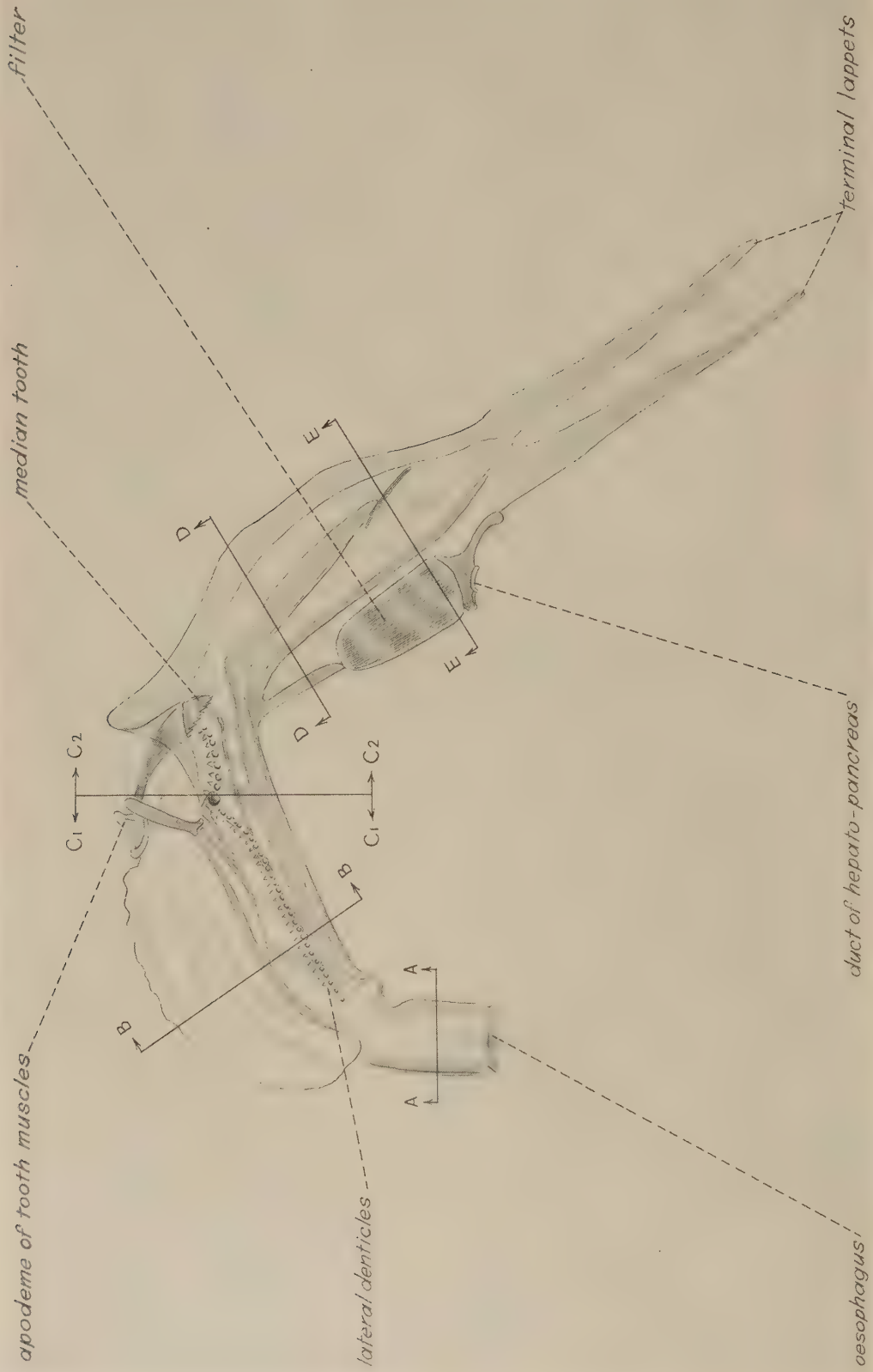


FIGURE 84.—Lateral view of cleared gastric mill showing sclerotized structures. The solid, lettered cross lines refer to the sections in figure 85. The arrows at the ends of the solid cross lines indicate the direction in which the viewer sees the sections in figure 85.

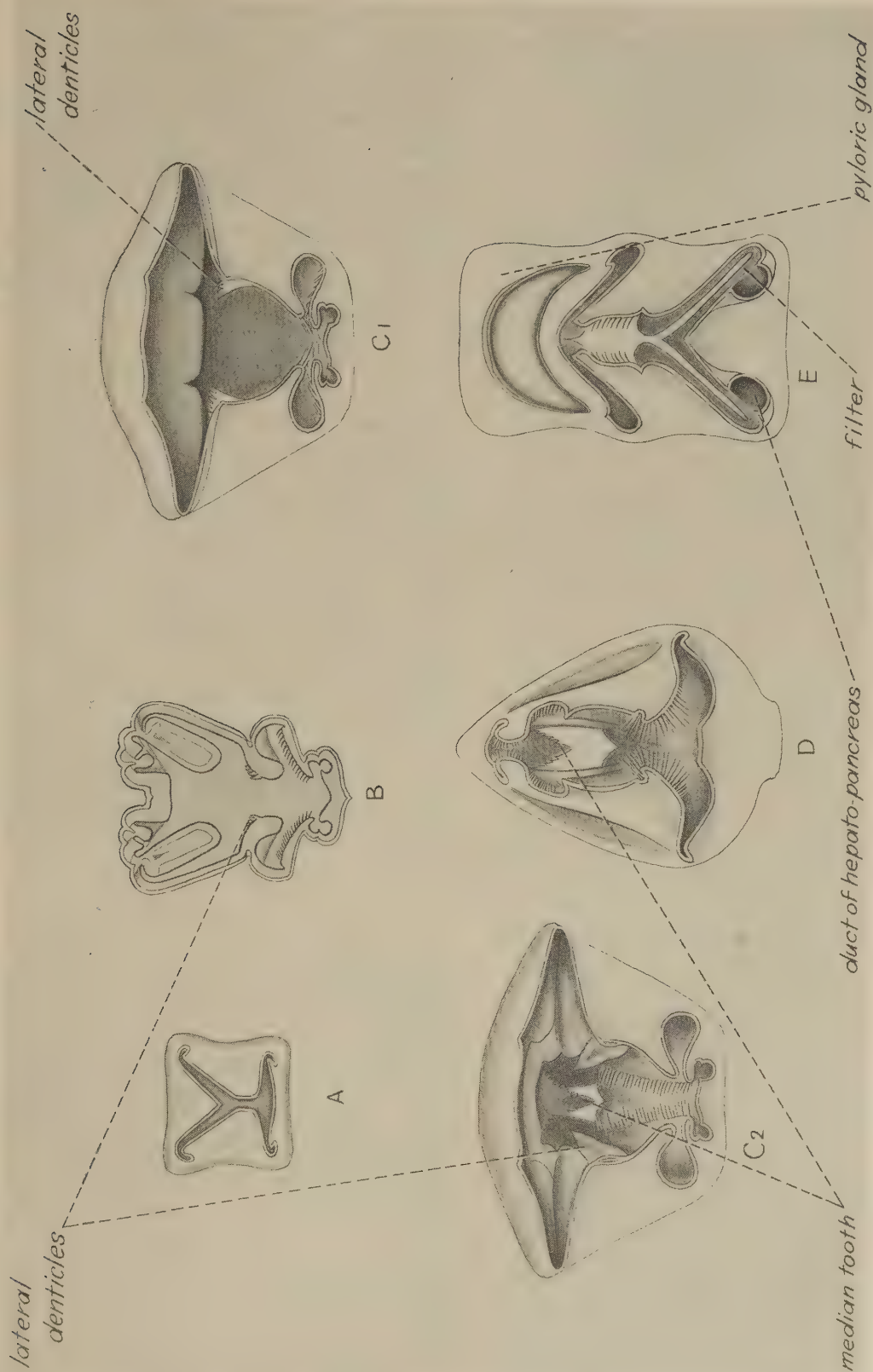


FIGURE 85.—Cross sections through the esophagus and gastric mill. The letters below the figures refer to the corresponding letters at the ends of the solid lines on figure 84.

(figs. 83, 84). In the posterior section of the gastric mill, dorsal to the filter, is a structure here designated as the dorsal (pyloric) gland (figs. 83; 85, *E*). The gland is mounted on the dorsal surface of the gastric mill. Its function is unknown.

The constrictor muscles of the gastric mill are arranged rather like those of the esophagus, in that the fibers are transverse to the long axis of the gut and are attached to lateral folds. The principal members of this group are the lateral gastric mill constrictor muscles (fig. 83). These attach between the long dorsolateral fold and the deep lateral fold dividing the inward ventral fold of the ventral channel. Contractions of the lateral constrictors reduce the diameter of the gut lumen and apply the lateral denticles to the gastric mill contents. Ventral gastric mill constrictor muscles (fig. 83) bring the setose margins of the ventral channel together. A third sheet of fibers, the ventral longitudinal muscle (fig. 83) runs between the lesser curvature of the gastric mill and the anterior end of the filter. Their function is not clear.

In general, the constrictor muscle systems is opposed by some 10 pairs of dilatator muscles. Just dorsal of the esophagus are two small lateral gastric mill dilatator muscles. Anterior to these is an anteroventral dilatator muscle, inserted hard by its mate on the other side. Next above is inserted the anterodorsal dilatator muscle, also close to its counterpart at the midline of the mill. On the anterodorsal surface of the gastric mill, caudad of the anterodorsal dilatators, is the mesodorsal dilatator muscle, inserting on the midline. Posteriorly, behind the curvature of the gastric mill, is inserted a group of perhaps three posteromedial dilatator muscles. Ventral to the gastric mill is found the long, thin posteroventral dilatator muscle which inserts with its counterpart into the ventral midline at the ventral curvature of the gastric mill. And lastly, a ventral filter dilatator muscle inserts into the ventral midline in the region of the filter.

Gross movements of the gastric mill, as well as operation of the median tooth, are accomplished by the large anterodorsal gastric mill muscle and the posterodorsal gastric mill muscle, both of which insert into the dorsal part of the mill, in the region of the median tooth. They are paired muscles. Some of the actions of these muscles may be observed in the living animal during feeding.

HEPATOPANCREAS

FIGURE 83

The main digestive gland of *Penaeus* appears to be the hepatopancreas. This large gland is situated in the posterior region of the thorax, anteroventrad of the heart. It surrounds the junction of the gastric mill with the midgut. In consistency, the hepatopancreas is a mass of closely packed secretory tubules whose products apparently are poured into the ducts of the hepatopancreas at the posterior end of the gastric mill filter.

B. Midgut

The extent of the midgut, or mesenteron, in Crustacea is evidently variable (Calman 1909). That part of the alimentary canal not sclerotized in *Penaeus* runs from some point near the posterior portion of the hepatopancreas to the hindgut gland in, the reader will recall, the anterior end of the sixth abdominal segment. The midgut is a simple, straight tube throughout its length. Its association with the dorsal abdominal artery is intimate. Very many small vessels connect the two all the way from the heart to the hindgut gland. Transverse sections of the midgut at various points show that the gut lumen is lined with low columnar epithelium resting on a basement membrane. The outer layers appears to be connective tissue in which muscle fibers are distributed.

C. Hindgut

The proctodaeum, or hindgut, begins at the hindgut gland and includes the rectum and anus. Specimens cleared in strong alkali retain the cuticular lining of this region of the gut. The hindgut (rectal) gland (figs. 61, 62, 74) projects dorsad from the rectum. The gland is composed of tubules lined with large secretory cells. The tubules are blind dorsally and open into the dorsal surface of the rectum. The function of the gland is unknown. The rectum (figs. 61, 74, 83), too, is lined with secretory epithelium reflected into deep folds. The gland cells of the hindgut gland and rectum may play a part in osmotic balance. The anus (fig. 82) consists of a bulb of cuticle at the caudal end of the rectum and a large tube passing ventrad to the anal opening. As has been shown, the opening is controlled by muscles.

V. EXCRETORY SYSTEM

The details of the excretory system and antennal gland of *Penaeus* have not been worked out completely. Compared to the compact and easily visible green gland of the crawfishes, the excretory organs of the white shrimp are much more diffused. Attempts have been made to clarify the extent of the excretory glands of *Penaeus* by injecting dyes into the excretory pores, and such information as is here presented is based on these procedures.

The hemocoelic excretory gland of the white shrimp seems to be made up of two major portions. The smaller, dorsal portion lies above the supraesophageal ganglion (fig. 86). The gland is continuous with the larger, ventral part of the system by means of its lateral arms. The ventral gland lies beneath the dorsal brain on the midline (fig. 86) and extends into the body of the antennae. The dorsal and ventral glands are composed of soft, glandular walls and trabeculae enclosing myriads of sinusoids. The portion of the gland entering the antenna makes contact with a compact, nodular, antennal excretory gland (figs. 20, 21) in the basal segments of the antenna. If the surface of the antennal gland is removed, gland concretions or nodules are found embedded in the tissues. A short duct from the antennal gland leads to the exterior through the excretory pore (figs. 21, 23, 24). The median coxal location of the pore is a constant character of Decapoda.

VI. REPRODUCTIVE SYSTEM

The external sex organs of Crustacea Decapoda have been widely used as systematic characters in the groups and those of the Penaeidae are no exception. In the males the petasma (fig. 87), a modified endopodite of the first pleopod, and in the females the thelycum (fig. 89), the modified internal plates of the fourth and fifth thoracic segments, have received attention. A system of descriptive nomenclature, largely meaningless, has grown up about the external sex characters, in particular the parts of the petasma. Of inferior taxonomic importance, but constant in the decapods, are the locations of the gonopores. Those of the male lie within folds of thin cuticle between the bases of the fifth walking legs (fig. 88), while the female gonoducts open to the exterior through biate structures between the third walking legs (fig. 89). A modification of the second pleopods,

the appendix masculina (fig. 3), varies in shape and size within the Tribe Penaeidae, that of some genera, as in *Penaeus*, being unspecialized compared to the complex appendix masculina in *Hymenopenaeus*.

The petasma of the male is a system of lightly sclerotized longitudinal rods connected to folds of thin cuticle which permit the petasma to inflate broadly. The structure is folded close to the basipodite of the first pleopod in young males. With the molt to sexual maturity, the medial margin of each half of the petasma becomes joined to its counterpart. Under the compound microscope one can see that the margins are invested with tiny, knobbed processes. Thrust together the knobs tend to form a lock, in the words of King (1948), "like a zipper." Although the copulatory act of penaeids has rarely been observed, apparently the spermatophore is extruded from the male gonopores and held on the posterior or ventral surface of the petasma and then thrust against the thelycum of the female where it is held in place by the setae and sculpturing of that region.

The thelycum of *Penaeus setiferus* (fig. 89) is made of a series of lobes and protuberances characteristic of the species (Burkenroad 1936). These presumably enable the animal to retain the spermatophore for a sufficient time to insure fertilization.

Enough research on the internal reproductive organs of *Penaeus setiferus* has been carried out in the present work to bear out the findings of King (1948), therefore no illustrations repeating his have been prepared on the subject.

The testes of the male are paired. They lie on the dorsal surface of the hepatopancreas ventral to the heart. Each testis is comprised of several lobes extending over the surface of the hepatopancreas. The vas deferens extends from the posterior end of the testis, makes a lateral loop and passes ventrad to the terminal ampoule, a vesicle situated above the male gonopore on the sternum. The ampoule is primarily a glandular structure that secretes the spermatophore.

The ovaries, like the testes, are paired. The ovary has a long, anterior projection which runs along the anterior portion of the esophagus and gastric mill and several lateral projections lying on the surface of the hepatopancreas. The heart is situated dorsal to the region with the lateral projections. Behind the heart a lobe of the ovary, the dorsal ovarian lobe (fig. 62), runs the length

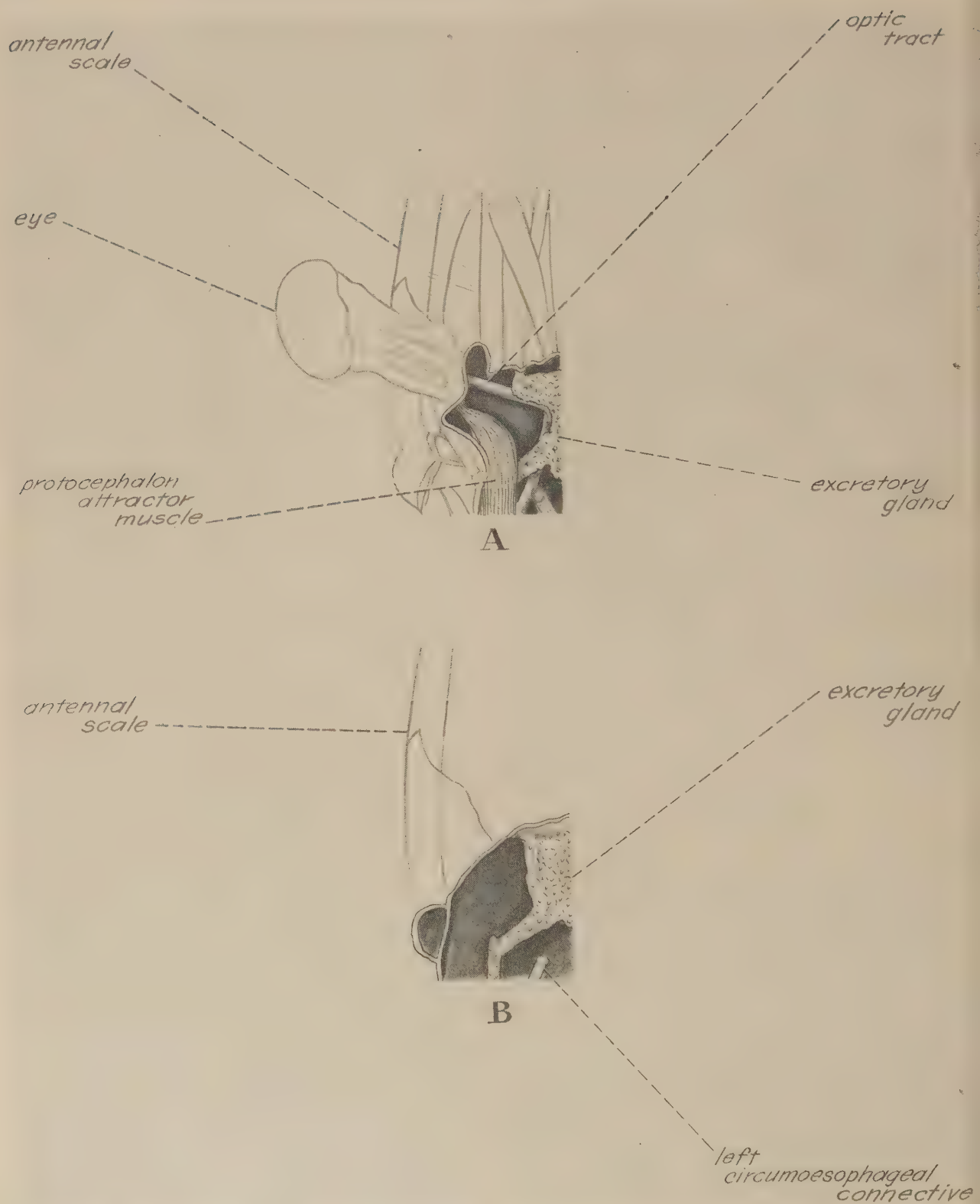


FIGURE 86.—Dorsal view of head region showing excretory gland. Carapace removed. A. Part of excretory gland dorsal to brain. B. Part of excretory gland ventral to brain.

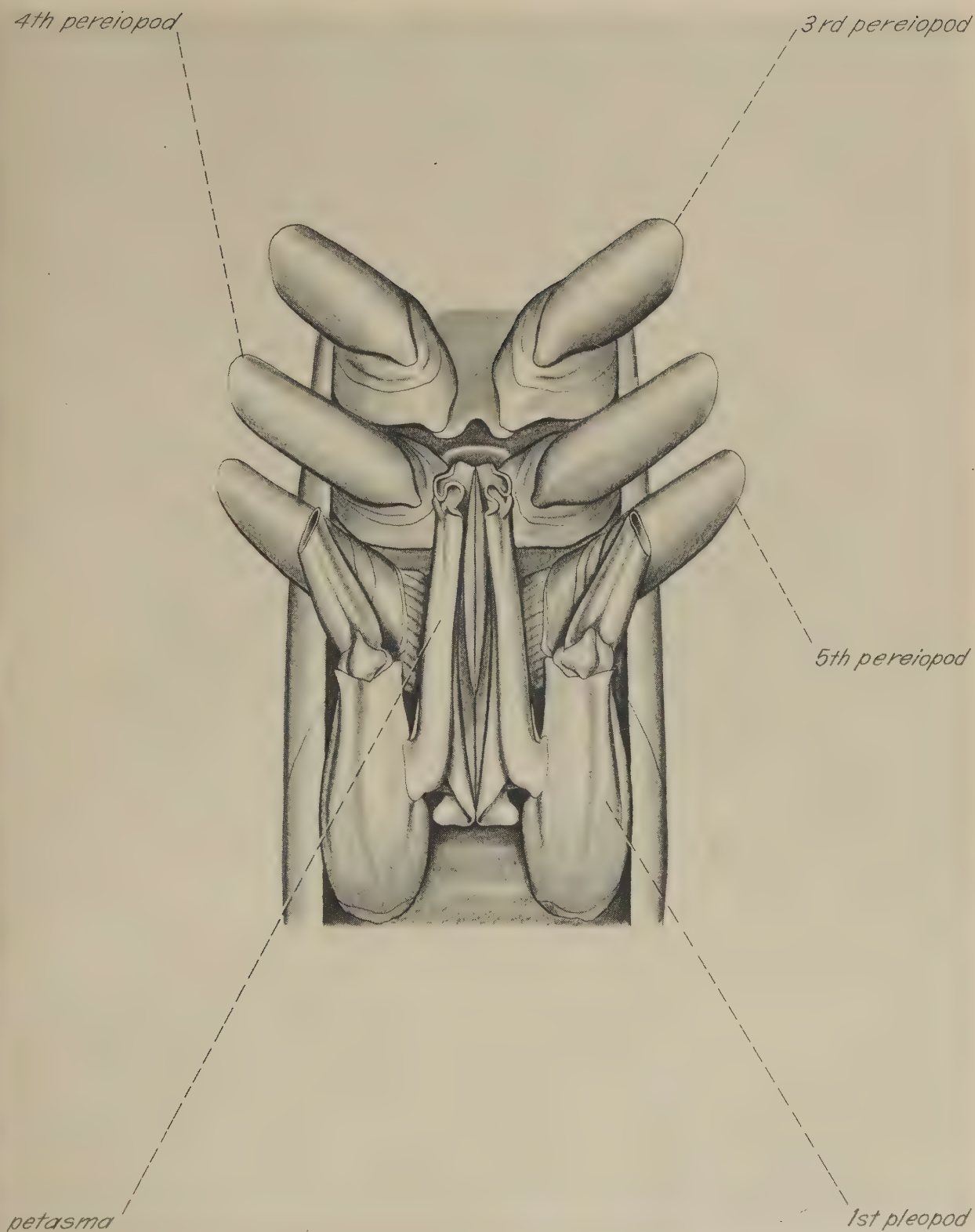


FIGURE 87.—Ventral view of parts of thorax and abdomen showing petasma of male.

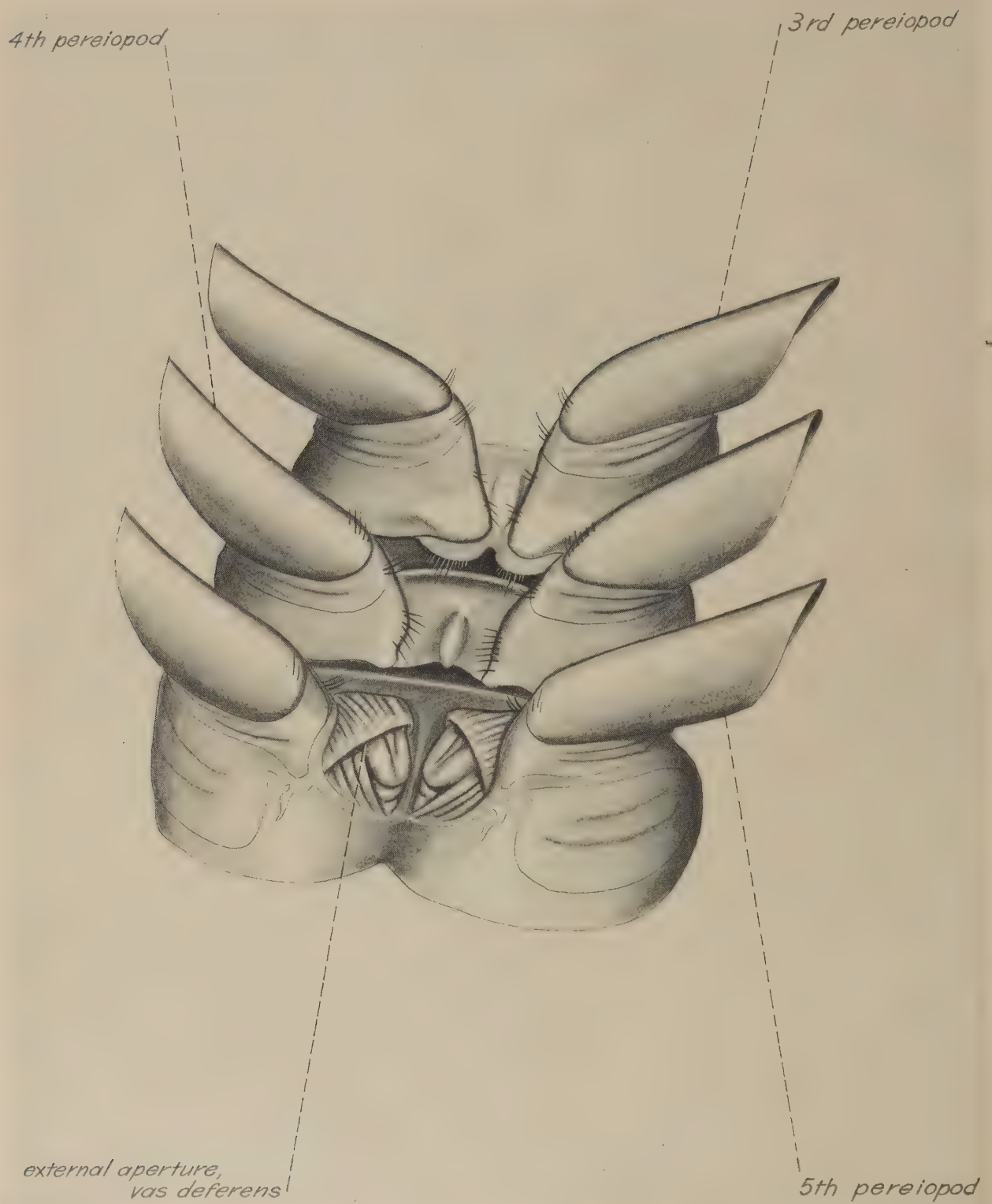


FIGURE 88.—Ventral view of parts of thorax in male. Petasma removed.

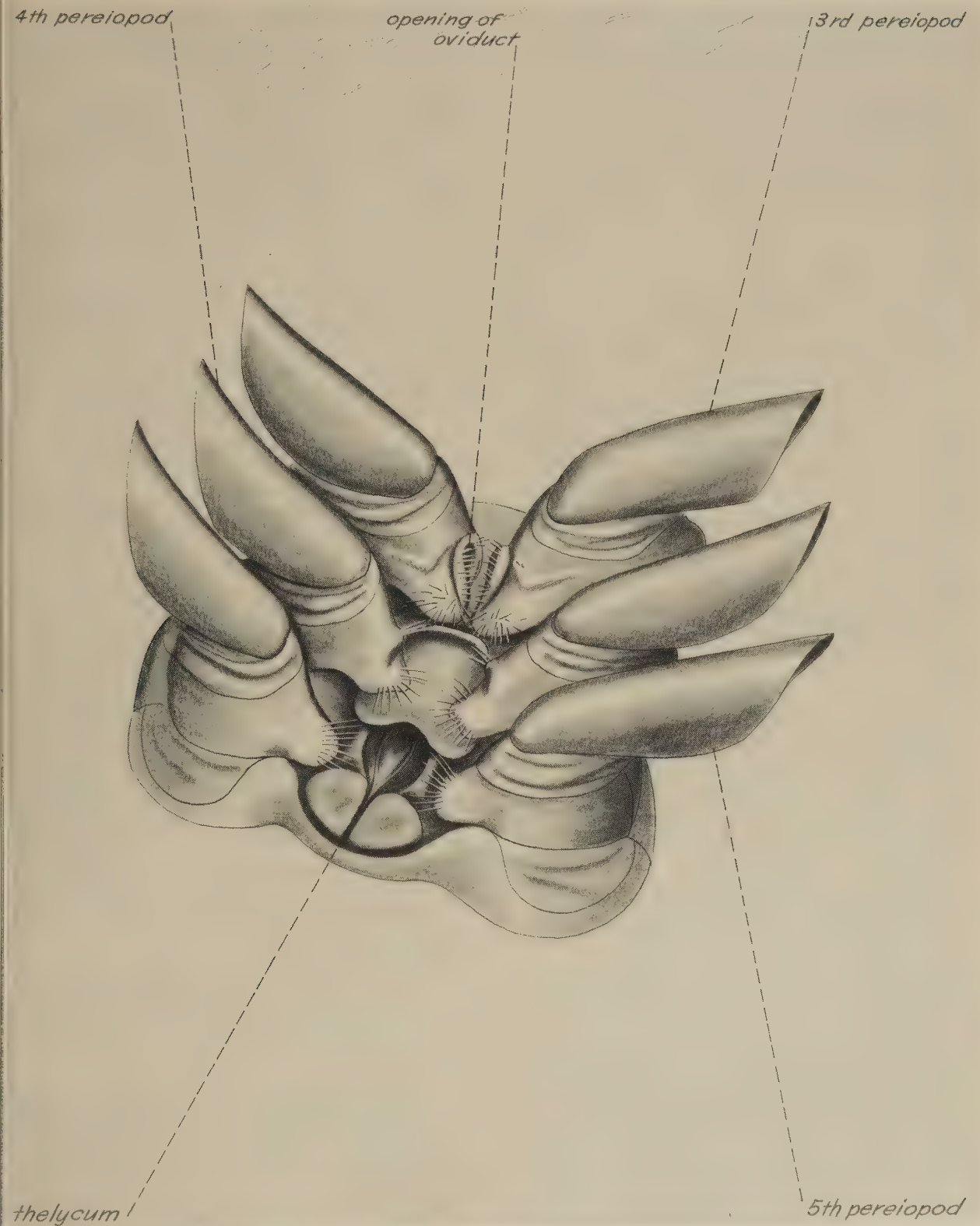


FIGURE 89.—Ventral view of parts of thorax in female.

of the abdomen dorsolateral to the midgut. The oviduct passes ventrad from the thoracic ovary to the female gonopore on the third thoracic leg.

LITERATURE CITED

- ALEXANDROWICZ, J. S.
1932. The innervation of the heart in the Crustacea. I. Decapoda. Quarterly Journal of Microscopical Science, London, 75: 181-249.
- BALSS, H.
1913. Ueber die Chemoreception bei Garneelen. Biologisches Zentralblatt, Leipzig, 33: 508-512.
- BAUMANN, H.
1917. Das Cor frontale bei dekapoden Krebsen. Zoologischer Anzeiger, 49: 137-144.
1919. Das Gefäß-system von *Astacus fluviatilis* (*Potamobius astacus* L.). Ein Beitrag zur Morphologie der Dekapoden. Zeitschrift für Wissenschaftliche Zoologie, Leipzig, 118: 246-312.
- BERKELEY, A. A.
1928. The musculature of *Pandalus danae* Stimpson. Transactions of the Royal Canadian Institute, Toronto, 16: 181-321.
- BERNHARDS, H.
1916. Der Bau des Komplexauges von *Astacus fluviatilis*. Zeitschrift für Wissenschaftliche Zoologie, 116: 649-707.
- BURKENROAD, MARTIN D.
1939. Further observations on Penaeidae of the northern Gulf of Mexico. Bulletin of the Bingham Oceanographic Collection, 6: 1-62.
- CALMAN, W. T.
1909. Part VII, Appendiculata, Third Fascicle, Crustacea. In: A treatise on zoology, ed. by Sir Ray Lankester. London, Adam and Charles Black. 346 pp.
- CARLISLE, D. B., and L. M. PASSANO.
1953. The X-organ of Crustacea. Nature, 171: 1070.
- CHAUDONNET, JEAN
1955. Le système nerveux de la région gnathale de l'écrevisse américaine (*Cambarus affinis* Say) et la question des paragnathes. (Note préliminaire) Travaux du Laboratoire de Zoologie et de la Station Aquicole Grimaldi de la Faculté des Sciences de Dijon, 12 1-6.
1956. Le système nerveux de la région gnathale de l'écrevisse *Cambarus affinis* (Say). Annales des Sciences Naturelles: Zoologie et Biologie Animale, 11^e Série, 18 (3): 33-61.
- COCHRAN, DORIS M.
1935. The skeletal musculature of the blue crab, *Callinectes sapidus* Rathbun. Smithsonian Miscellaneous Collection, 92 (9): 1-76.
- DANIEL, R. J.
1928. The abdominal muscular systems of *Praunus flexuosus* (Müller). Proceedings and Transactions of the Liverpool Biological Society (Appendix, 36 Lanc Sea Fish. Lab. Rept. for 1927), 42: 5-41.
1929. The abdominal muscular system of *Meganyctiphanes norvegica* (M. Sars). Proceedings and Transactions of the Liverpool Biological Society, 43: 149-180.
- DANIEL, R. J.—Continued
1931a. The abdominal muscular systems of *Homarus vulgaris* (L.) and *Palinurus vulgaris* (Latr.). Proceedings and Transactions of the Liverpool Biological Society, 45: 3-49.
1931b. The abdominal muscles of the shore crab (*Carcinus moenas*) and of the Zoëa and Megalope stages. Proceedings and Transactions of the Liverpool Biological Society, 45: 50-56.
1931c. Comparative study of the abdominal musculature in Malacostraca. Part I. The main ventral muscles of the typical abdominal segments. Proceedings and Transactions of the Liverpool Biological Society, 45: 57-71.
1931d. The abdominal muscular systems of *Paranapides lacustris* (Smith). Report of the Lancashire Sea-Fish. Laboratory for 1931, pages 26-45.
1933. The abdominal muscular systems of *Lophogaster typicus* M. Sars, and *Gnathophausia Zoëa* Suhm, and their relationships with the musculatures of other Malacostraca. Proceedings and Transactions of the Liverpool Biological Society, 47: 71-133.
- DEBAISIEUX, P.
1954. Histogénèse des muscles et charpentes chez les Crustacés. La Cellule, Liège, T. 56, 3: 265-305.
- DOFLEIN, FRANZ.
1911. Ueber den Geruchssinn bei Wassertieren. Biologisches Zentralblatt, Leipzig, 31: 706-707.
- FERRIS, G. F.
1947. The contradictions of the insect head. Microentomology, 12 (3): 60-64.
1953. On the comparative morphology of the Annulata. A summing up. Microentomology, 18: 2-15.
- GROBBEN, K.
1917. Der Schalenschliessmuskel der dekapoden Crustaceen, zugleich ein Beitrag zur Kenntnis ihrer Kopfmuskulatur. Sitzungsberichte der Kaiserliche Akademie der Wissenschaften, Wien, Abt. 1. 126: 473-494.
1919. Über die Muskulatur des Vorderkopfes der Stomatopoden. Sitzungsberichte der Kaiserliche Akademie der Wissenschaften, Wien, Abt. 1, 128: 185-214.
- HANSTRÖM, B.
1948. The brain, the sense organs, and the incretory organs of the head in the Crustacea Malacostraca. Bulletin Biologique de France et de Belgique, Supplément 33: 98-126.
- HART, C. W., JR.
1952. The exoskeleton and musculature of the appendages of the first three abdominal segments of *Cambarus longulus longulus* Girard (Decapoda, Astacidae). Virginia Journal of Science, 3 (1): 39-48.
- HENRY, L. M.
1948a. The nervous system and the segmentation of the head in the Annulata. Section IV, Arthropoda. Microentomology, 13 (1): 1-26.
1948b. The nervous system and the segmentation of the head in the Annulata. Section V, Onychophora. Microentomology, 13 (2): 27-48.

- HUDINAGA, M.
1935. Studies on the development of *Penaeus japonicus* Bate. Report of the Hayatomo Fishery Institute, Tobata, Vol. 1, No. 1. Abstracted in: Japanese Journal of Zoology, 6 (4) : 80, 1936.
1942. Reproduction, development and rearing of *Penaeus japonicus* Bate. Japanese Journal of Zoology, 10 (2) : 305-393.
- HUXLEY, T. H.
1906. The crayfish. 7th ed. Kegan Paul, Trench, Trübner & Co., Ltd., London. 371 pages.
- JOHANNSEN, O. A., and F. H. BUTT.
1941. Embryology of insects and myriapods. McGraw-Hill Book Co., New York.
- JOHNSON, M. C., and J. R. FIELDING.
1956. Propagation of the white shrimp, *Penaeus setiferus* (Linn.), in captivity. Tulane Studies in Zoology, 4 (6) : 175-190.
- KEIM, W.
1915. Das Nervensystem von *Astacus fluviatilis* (*Potamobius astacus* L.). Zeitschrift für Wissenschaftliche Zoologie, 113: 485-545.
- KING, J. E.
1948. A study of the reproductive organs of the common marine shrimp, *Penaeus setiferus* (Linnaeus). Biological Bulletin, 94: 244-262.
- KISHINOUE, K.
1900. The nauplius stage of *Penaeus*. Zoologischer Anzeiger, Leipzig, 23 (607) : 73-75.
- KNOWLES, F. G. W.
1953. Endocrine activity in the crustacean nervous system. Proceedings of the Royal Society of London, Series B, Biological Sciences, 141: 248-267.
- KNOWLES, F. G. W., and D. B. CARLISLE.
1956. Endocrine control in the Crustacea. Biological Reviews, 31: 396-473.
- LISSMAN, H. W., and A. WOLSKY.
1935. Antennulen-Aussenast als Chemorezeptor. Erwiderung an Herrn Luther. Zoologischer Anzeiger, Leipzig, 110: 92-96.
- MAYRAT, A.
1955. Mise en évidence de tendons chez les crustacés dans le muscle attracteur du synciput de *Praunus flexuosus* O. F. Müller. Bulletin de la Société Zoologique de France, 80 (2-3) : 81-85.
1956a. Le système artériel de *Praunus flexuosus* et le prétendu cœur frontale des Malacostracés. Bulletin, Station Océanographique de Salammbô, Tunisia, 53: 44-49.
1956b. Oeil, centres optiques et glandes endocrines de *Praunus flexuosus* (O. F. Müller). Archives de Zoologie expérimentale et Générale, 93 (Fasc. 4) : 319-363.
- MEREDITH, S. S.
1954. The appendages of *Crangon vulgaris*, the brown shrimp. Proceedings of the Leeds Philosophical and Literary Society, Scientific Section, 6 (4) : 228-242.
- MÜLLER, F.
1863. Die Verwandlung der Garneelen. Arch. Naturgesch., 29: 8-23. (Translated in Annals and Magazine of Natural History, 14 (3) : 104-115, 1864).
- PASSANO, L. M.
1953. Neurosecretory control of molting in crabs by the X-organ sinus gland complex. Physiologia Comparata et Oecologia, 3 (2-3) : 155-189.
- PEARSON, J. C.
1939. The early life histories of some American Penaeidae, chiefly the commercial shrimp, *Penaeus setiferus* (Linn.). Bulletin of the Bureau of Fisheries, 49 (30) : 1-73.
- PERKINS, E. B.
1928. Color changes in crustacea, especially in *Palaeomonetes*. Journal of Experimental Zoology, 50: 71-105.
- RAMADAN, M. M.
1952. Contribution to our knowledge of the structure of the compound eyes of Decapoda Crustacea. Acta Universitatis Lundensis, N. S. 2, 48 (3) : 1-20.
- RICHARDS, A. GLENN.
1951. The integument of arthropods. University of Minnesota Press. 411 pages.
- RIOJA, ENRIQUE
1939a. Estudios carcinológicos. I. Caracteres sexuales secundarios de los Penaeidae. El órgano antenular de los machos. Anales del Instituto de Biología, Mexico, 10 (3-4) : 313-319.
1939b. Estudios carcinológicos. II. Caracteres sexuales secundarios de los Penaeidae. Los caracteres sexuales en algunas especies de Aristeinae: *Aristeus antennatus* (Risso), *Aristeomorpha foliacea* (Risso) y *Plesiopenaeus edwardsianus* (J. G. Johnson). Anales del Instituto de Biología, Mexico, 10 (3-4) : 322-333.
1940a. La morfología de las cerdas de las piezas bucales de los Penaeidos (Crus. Decap.) y su valor diagnóstico. Ciencia Mexico, 1: 116-117.
1940b. Estudios carcinológicos. III. Descripción de un órgano setiforme en el tercer maxilipede de algunos Penaeidae. Anales del Instituto de Biología, Mexico, 11 (1) : 261-266.
1940c. Estudios carcinológicos. IV. Observaciones sobre las antenulas de algunas especies del género *Penaeus*. Anales del Instituto de Biología, Mexico, 11 (1) : 267-273.
1941a. Estudios carcinológicos. VIII. Contribución al la morfología e interpretación del petasma de los Penaeidae (Crust. Decapodos). Anales del Instituto de Biología, Mexico, 12 (1) : 199-221.
1941b. Estudios carcinológicos. IX. Descripción del macho maduro de *Penaeus vannamei* Boone, hallado en las costas Pacífico de Mexico. Anales del Instituto de Biología, Mexico, 12 (1) : 223-229.
1942a. Estudios carcinológicos. XI. Observaciones acerca de algunos caracteres sexuales secundarios en el camarón de rostro largo (*Artemesia longinaris* Bate) y en el langostin (*Hymenopenaeus mülleri* Bate). Anales del Instituto de Biología, Mexico, 13 (2) : 659-674.
1942b. Estudios carcinológicos. XII. Contribución al estudio de los caracteres sexuales secundarios de dos especies de los géneros *Trachypeneus* y *Xiphopenaeus* de las costas Mexicanas del Pacífico. Anales del Instituto de Biología, Mexico, 13 (2) : 675-684.

SCHMIDT, W.

1915. Die Muskulatur von *Astacus fluviatilis* (*Potamobius astacus* L.). Zeitschrift für Wissenschaftliche Zoologie, 113: 165-251.

SNODGRASS, R. E.

1935. Principles of insect morphology. McGraw-Hill Book Co., New York. 667 pages.
1938. Evolution of the Annelida, Onychophora, and Arthropoda. Smithsonian Miscellaneous Collections, 97 (6): 1-159.
- 1951a. Comparative studies on the head of mandibulate arthropods. Comstock Publishing Co., Ithaca, N. Y. 118 pages.
- 1951b. Comparative studies on the jaws of mandibulate arthropods. Smithsonian Miscellaneous Collections, 116: 1-85.
1952. A textbook of arthropod anatomy. Comstock Publishing Associates, Ithaca, N. Y. 363 pages.

SPIEGEL, A.

1927. Über die Chemorezeption von *Crangon vulgaris* Fabr. Zeitschrift für Wissenschaftliche Biologie, Abt. C. Zeitschrift für Vergleichende Physiologie, 6: 688-730.

STEINBERG, J. E., and E. C. DOUGHERTY.

1957. The skeleton shrimps (Crustacea: Caprellidae) of the Gulf of Mexico. Tulane Studies in Zoology, 5 (11): 267-288.

VOSS, G. L.

1955. A key to the commercial & potentially commercial shrimp of the Family Penaeidae of the Western North Atlantic and the Gulf of Mexico. Technical Series No. 14, The Marine Laboratory, University of Miami, pp. 1-23.

WELSH, J. H.

1941. The sinus gland and the 24-hour cycles of retinal pigment migration in the crayfish. Journal of Experimental Zoology, 86 (1): 35-49.

YOUNG, J. H.

1953. Embryology of the mouthparts of Anoplura. Microentomology, 18 (4): 85-133.
1956. Anatomy of the eyestalk of the white shrimp, *Penaeus setiferus* (Linn., 1758). Tulane Studies in Zoology, 10 (3): 169-190.

BIBLIOGRAPHY

ABRAHAM, A., and A. WOLSKY.

1930. Die Geruchsorgane der Landisopoden. Zeitschrift für Wissenschaftliche Biologie, Abt. für Anatomie, Morphologie, und Ökologie der Tiere, 17: 441-463.

ABRAHAMCZIK-SCANZONI, H.

1942. Beiträge zur Kenntnis der Muskulatur und des Innenskeletts der Krabben. Zoologische Jahrbücher, Abt. 2 für Anatomie und Ontogenie der Tiere, Jena, 67: 293-380.

AIYER, R. P.

1953. On the female reproductive system of *Palaeomonidae* Heller. Journal of the Zoological Society of India, 5 (2): 227-234.

ALEXANDROWICZ, J. S.

1933. Innervation des branchies de *Squilla mantis*. Archives de Zoologie Expérimentale et Générale, Paris, 75: 21-34.
1934. The innervation of the heart of Crustacea. II. Stomatopoda. Quarterly Journal of Microscopical Science, London, 76: 511-548.
1951. Muscle receptor organs in the abdomen of *Homarus vulgaris* and *Palinurus vulgaris*. Quarterly Journal of Microscopical Science, London, 92: 163-199.
- 1952a. Innervation of the heart of *Ligia oceanica*. Journal of the Marine Biological Association, U. K., 31: 85-95.
- 1952b. Receptor elements on the thoracic muscles of *Homarus vulgaris* and *Palinurus vulgaris*. Quarterly Journal of Microscopical Science, 93: 315-346.

ANDERSON, W. W., J. E. KING, and M. J. LINDNER.

1949. Early stages in the life history of the common marine shrimp. *Penaeus setiferus* (Linnaeus). Biological Bulletin, 96 (2): 168-172.

ANDERSON, W. W., and MILTON J. LINDNER.

1945. A provisional key to the shrimps of the family Penaeidae with especial reference to American forms. Transactions of the American Fisheries Society, 73 (for year of 1943): 284-319.

ANDREWS, E. A.

1911. Sperm transfer in certain decapods. Proceedings of the Smithsonian Institution, U. S. National Museum, 39: 419-434.

ARGILAS, A.

1929. Observations morphologiques sur les espèces des côtes algériennes de la famille des Penaeidae (Macroures nageurs). Bulletin des Travaux Publiés par la Station d'Aquiculture et de Pêche de Castiglione, Nouvelle Serie, Algiers, fasc. 1, pp. 29-101.

ARVY, L., M. GABE, and J. LHOSTE.

1947. Etude mécanique du sang de *Ligia oceanica* L. Bulletin du Laboratoire Maritime du Muséum National d'Histoire Naturelle, Dinard, Fasc. 29, pp. 5-10.

BARTH, G.

1934. Untersuchungen über Myochordotonalorgane bei Dekapoden Crustaceen. Zeitschrift für Wissenschaftliche Zoologie, Leipzig, 145: 576-624.

BELL, J. C.

1906. The reactions of crayfish to chemical stimuli. Journal of Comparative Neurology, Philadelphia, 16: 299-326.

BERNARD, F.

1937. Recherches sur la morphogénèse des yeux composés d'Arthropodes. Développement, croissance, réduction. Bulletin Biologique de la France et de la Belgique, Supplement, 23: 1-162.

BLOCH, F.

1935. Contribution à l'étude des gamètes et de la fécondation chez les Crustacés Décapodes. Travaux de l'Institut Zoologique de Lille et de la Station Maritime de Wimereux (Pas-de-Calais), 12: 181-279.

- BOAS, J. E. V.
1889. Kleinere carcinologische Mitteilungen. 2. Ueber den ungleichen Entwicklungsgang der Salzwasser und der Süßwasser-Form von *Palaemonetes varians*. Zoologische Jahrbücher, Abt. 3 für Systematik Ökologie und Geographie der Tiere, Jena, 4: 793-805.
- BOCK, F.
1923. Die Blutcirculation in den Kiemen von Flusskrebs. Zoologischer Anzeiger, 56: 198-205.
1925. Die Respirationsorgane von *Potamobius astacus* Leach. Zeitschrift für Wissenschaftliche Zoologie, 124: 51-117.
- BÖRNER, C.
1921. Die Gliedmassen der Arthropoden. In Arnold Lang's Handbuch der Morphologie der Wirbellosen Tiere, Jena, 4: 649-694.
- BORRADAILE, L. A.
1907. On the classification of decapod crustaceans. Annals and Magazine of Natural History, Series 7, Vol. 19, p. 147.
- BOUVIER, E. L.
1889. Le système nerveux des Crustacés Décapodes et ses rapports avec l'appareil circulatoire. Annales des Sciences Naturelles: Zoologie et Biologie Animale, 11^e Série, Paris, (7): 73-106.
1891. Recherches anatomiques sur le système artériel des Crustacés Décapodes. Annales des Sciences Naturelles: Zoologie et Biologie Animale, 11^e Série, Paris, (7), 9: 197-282.
- BOUVIER, L.
1936a. Observations relatives à l'Écrevisse sur la constitution des flancs chez les Crustacés. Comptes Rendus Hebdomadaires des Séances de L'Académie des Sciences, Paris, 203: 222-225.
1936b. Observations complémentaires sur les pattes des Crustacés décapodes appartenant à la section des Astacomorphes. Comptes Rendus Hebdomadaires des Séances de L'Académie des Sciences, Paris, 203: 1301-1305.
- BOYDEN, A.
1943. Serology and animal systematics. American Naturalist, 77: 234-255.
- BRIAN, A.
1938. L'armatura triturante dello stomaco di alcuni Peneidi meso-abissali a della *Pasiphaea multidentata* Esmark. Monitore Zoologica Italiano, Florence, 49: 153-165.
- BROAD, CARTER.
1950. Results of shrimp research in North Carolina. Proceedings of the Gulf and Caribbean Fisheries Institute, Third Annual Session, 9 pages.
- BROCK, F.
1948. Das Verhalten dekapoder Crustaceen beim Aufbau des Beutefeldes. Remane and others in: Verhandlungen der Deutsche Zoologischen Gesellschaft, Geest and Portig, Leipzig, 1949, pp. 416-426. (Published as Supplementband, Zoologischer Anzeiger.)
- BROOKS, W. K.
1882. Lucifer: A study in morphology. Philosophical Transactions of the Royal Society of London, Series B, Biological Sciences, 173: 57-137.
1883. The metamorphosis of *Penaeus*. Annals and Magazine of Natural History, 2 (19): 6.
BROOKS, W. K., and F. H. HERRICK.
1892. The embryology and metamorphosis of the Macroura. Memoirs of the National Academy of Science, 5: 323-463.
- BURKENROAD, MARTIN D.
1934a. The Penaeidae of Louisiana with a discussion of their world relationships. Bulletin of the American Museum of Natural History, 68 (2): 61-143.
1934b. Littoral Penaeidae chiefly from the Bingham Oceanographic Collection. With a revision of *Penaeopsis* and descriptions of two new genera of eleven American species. Bulletin of the Bingham Oceanographic Collection, 4 (7): 1-109.
1936. A new species of *Penaeus* from the American Atlantic. Ann. Acad. Brasileira Sci., T. 8, No. 4.
1937. The Templeton Crocker Expedition. XII. Sergestidae (Crustacea Decapoda) from the lower Californian region, with descriptions of two new species and some remarks on the Organs of Pesta in *Sergestes*. Zoologica, New York, 22: 315-329.
1938. The Templeton Crocker Expedition. XIII. Penaeidae from the region of Lower California and Clarion Island, with descriptions of four new species. Zoologica, New York, 23 (1): 55-91.
1947. Reproductive activities of decapod Crustacea. American Naturalist, 81: 392-398.
1949. Occurrence and life histories of commercial shrimp. Science, 110 (2869): 688-689.
1950. Measurement of the natural growth rates of decapod crustaceans. Proceedings of the Gulf and Caribbean Fisheries Institute, Third Annual Session, 2 pp.
- BRUNTZ, L.
1903. Contribution à l'étude de l'excrétion chez les Arthropodes. Archives de Biologie, Liège, 20: 217-422.
- BUDDENBROCK, W. VON.
1914. Orientierung der Kresbse. Zoologische Jahrbücher, Abt. für Allgemeine Zoologie und Physiologie, 34: 479.
- CALMAN, W. T.
1917. Notes on the morphology of *Bathynella* and some allied Crustacea. Quarterly Journal of Microscopical Science, 62: 489-514.
1920. The Cambridge natural history. Volume IV, ed. by S. F. Harmer and A. E. Shipley, p. 162.
- CARLISLE, D. B., and P. F. R. DOHRN.
1953. Studies on *Lysmata seticaudata* Risso. VI. Notes on the structure of the neurosecretory system of the eyestalk. Pubblicazioni della Stazione Zoologica di Napoli, 24: 69-83.
- CARLISLE, D. B., and C. G. BUTLER.
1956. The "queen-substance" of honeybees and the ovary-inhibiting hormone of crustaceans. Nature, 177: 276-277.

- CHACE, F. A., JR.
1951. The number of species of decapod and stomatopod Crustacea. *Journal of the Washington Academy of Sciences*, 41: 370-372.
- CHAPPUIS, P. A.
1915. *Bathynella natans* und ihre stellung in System. *Zoologische Jahrbücher, Abt. für Systematik, Geographie und Biologie der Tiere, Jena*, 40: 147-176.
- CHIDESTER, F. E.
1912. The biology of the crayfish. *American Naturalist*, 46: 279-293.
- CLARK, E., and F. M. BURNET.
1942. The application of serological methods to the study of Crustacea. *Australian Journal of Experimental Biology and Medical Science, Adelaide*, 20: 89-95.
- CLAUS, C.
1891. Das Medianauge der Crustaceen. *Arb. Zool. Inst. Univ. Wien*, 9: 225-266.
- CRAMPTON, G. C.
1928a. The evolution of insects, chilopods, diplopods, Crustacea, and other arthropods indicated by a study of the head capsule. *Canadian Entomologist*, 60: 284-301.
1928b. The evolution of the head region in lower arthropods and its bearing upon the origin and relationships of the arthropodan groups. *Canadian Entomologist*, 60: 284-301.
- CRONIN, L. E.
1947. Anatomy and histology of the male reproductive system of *Callinectes sapidus* Rathbun. *Journal of Morphology*, 81: 209-239.
- DAKIN, W. H.
1946. Life history of a species of *Metapenaeus* in Australian coastal lakes. *Nature*, 158 (4003): 99.
- DANIEL, R. J.
1930. The abdominal muscular systems of the Zoëa and Mysis stages of the shrimp (*Crangon vulgaris* Fabr.) and their bearing on phylogeny. *Proceedings and Transactions of the Liverpool Biological Society*, 44: 95-109.
1932. Comparative study of the abdominal musculature in Malacostraca. Part II. The superficial and main ventral muscles and lateral muscles and their continuation into the thorax. Report of the Lancashire Sea-Fish. Laboratory for 1931, pp. 46-104.
- DEBAISIEUX, P.
1948. Statocystes de *Praunus* et leurs poils sensoriels. *Annales de la Société Royale Zoologique de Belgique*, 78: 26-31.
- DELAGE, Y.
1183. Circulation et respiration chez les Crustacés Schizopodes. *Archives de Zoologie Expérimentale et Générale, Paris, S. 2, T. 1*, pp. 105-130.
- DEMAL, JEAN
1953. Genèse et différenciation d'hémocytes chez *Palaeomon varians* Leach. *La Cellule, Liège*, 56: 85-102.
- DESMARETS, A.
1929. Rapport entre la carapace des Crustacés et les organes internes. *Bulletin de la Société d'Etude des Sciences Naturelles d'Elbeuf*, 47: 65-74.
- DUCHATEAU, G., ET AL.
1940. L'équipement enzymatique de l'hépatopancreas des Crustacés en ce que concerne le catabolisme des purines. *Comptes Rendus des Séances de la Société de Biologie et de ses Filiales, Paris*, 133: 274-276.
- EINARSSON, H.
1942. On the systematic value of the spermatheca, on sexual dimorphism in *Nematoscelis*, and on the male in *Bentheuphausia*. *Videnskabelige Meddelelser Fra Dansk Naturhistorisk Forening I Kbenhavn*, 106: 263-286.
- ESCHALIER, G.
1954. Recherches expérimentales sur le rôle de "l'organ Y" dans la mue de *Carcinus moenas* (L.), Crustacé Décapode. *Comptes Rendus Hebdomadaires des Séances de L'Académie des Sciences, Paris*, 238: 523-524.
- ESTEVE, MICHELINE.
1954. Présence simultanée d'un thelycum et d'un petasma chez *Parapenaeus longirostris* (Lucas) (Crustacé, Penaeidae). *Bulletin de la Société des Sciences Naturelles de Tunisie, Tome 8*, pp. 89-92.
- FAGE, L.
1944. Sur quelques caractères sexuels secondaires des Cumacés du genre *Iphinoë*. *Archives de Zoologie Expérimentale et Générale, Paris, Notes Rev.*, 83: 112-121.
- FAXON, W.
1895. Reports . . . *Albatross*, 15: The stalk-eyed Crustacea. *Memoirs of the Museum of Comparative Zoology, Harvard*, 18: 292 pp.
- FERRIS, G. F.
1944. On certain evolutionary tendencies in the heads of insects. *Microentomology*, 9: 78-84.
- FESQUET, A. E. J.
1933. Anotaciones para una monografía sobre el Langostin (*Pleoticus mülleri* Bate). A. Kapelusz y Cia, Buenos Aires, pp. 1-35.
1941a. Estudio de la morfología externa del Camaron de rostro largo (*Artemesia*). *Anales de la Sociedad Científica Argentina, Buenos Aires*, 131: 60-95.
1941b. Descripción del mecanismo articular de los pedunculos oculares de *Artemesia longinaris* y de *Himpenopenaeus mülleri* (Bate). *Holmbergia: Revista del Centro de Estudiantes del Doctorado en Ciencias Naturales, Buenos Aires*, 3 (6): 64-67.
- FISCHBACH, E.
1951. Licht-, Schwere- und Geruchssinn bei Isopoden. *Die Naturwissenschaften*, 38 (17): 408.
- GABE, M.
1953. Sur l'existence, chez quelques Crustacés Malacostracés d'un organ comparable à la glande de la mue des Insectes. *Comptes Rendus Hebdomadaires des Séances de L'Académie des Sciences, Paris*, 237 (18): 1111-1113.

- GATES, W. H.
1910. Shrimp. Fifth Biennial Report, Director, Gulf Biologic Station, Cameron, Louisiana, pp. 8-12.
- GEORGE, W. C. and J. NICHOLS.
1948. A study of the blood of some Crustacea. *Journal of Morphology*, 83: 425-440.
- GERARD, P.
1937. Sur l'élimination rénale des colorants acides chez *Astacus fluviatilis*. *Bulletin de la Classe des Sciences, Académie Royale de Belgique*, 5^e Serie, 23: 456-463.
- GOWANLOCH, J. N.
1931. The probable cause of "iodine shrimp." *Louisiana Conservation Review*, 2: 31-32.
- GRABER, V.
1888. Ueber die Empfindlichkeit einiger Meertiere gegen Riechstoffe. *Biologisches Zentralblatt*, Leipzig, 8: 743-754.
- GROBEN, K.
1881. Die Antennendrüse der Crustaceen. *Arbeiten aus dem Zoologischen Institute der Universität Wien und der Zoologischen Station in Triest*, 3: 93-110.
1892. Zur Kenntnis des Stammbaumes und des Systems der Crustaceen. *Sitzungsberichte der Kaiserliche Akademie der Wissenschaften*, Wien, Abt. 1, 101: 237-274.
- GUILBEAU, B. H.
1908. Shrimp. Fourth Biennial Report, Director, Gulf Biologic Station, Cameron, La., pp. 11-13.
- GURNEY, R.
1924. Crustacea, Part 9. Decapod larvae. *British Antarctic (Terra Nova) Expedition, 1910. Natural History Report, Zool., Volume 8, No. 2*.
1926. The protozoal stage in decapod development. *Annals and Magazine of Natural History*, 18 (Series 9, No. 103): 19-27.
1927. Report on the larvae of the Crustacea Decapoda. *Cambridge Expedition, Suez Canal. Transactions of the Zoological Society of London*, 22 (2) No. 15.
1942. Larvae of decapod Crustacea. *The Ray Society, London*. 306 pp.
1943. The larval development of two penaeid prawns from Bermuda of the genera *Sicyonia* and *Penaeopsis*. *Proceeding of the Zoological Society of London*, 113B: 1-16.
1950. A remarkable penaeid larva. *Proceedings of the Zoological Society of London*, 119: 803-806.
- HANSON, H. J.
1893. Zur Morphologie der Gliedmassen und Mundtheile bei Crustaceen und Insecten. *Zoologischer Anzeiger*, 16: 193-198 and 201-212.
- HANSTRÖM, B.
1929. Das Deutocerebrum der Crustaceen. *Zoologische Jahrbücher*, Jena, 51: 535-548.
- HARMON, M.
1937. Les mecanismes produisant la dehiscence des spermatophores d'*Eupagurus prideauxi* Leach. *Comptes Rendus Hebdomadaires des Séances de L'Académie des Sciences*, Paris, 204: 1504-1506.
- HART, C. W., JR.
1953. Serial homologies among three pairs of abdominal appendages of certain male crayfishes (Decapoda, Astacidae). *Journal of Morphology*, 93 (2): 285-299.
- VAN HARREVELD, A.
1939a. The motor innervation of a triply innervated crustacean muscle. *Journal of Experimental Biology*, London, 16: 398-402.
1939b. The nerve supply to doubly and triply innervated crayfish muscles related to their function. *Journal of Comparative Neurology*, Philadelphia, 70: 267-284; 285-296.
- VAN HARREVELD, A., and C. A. G. WIERSMA.
1939. The function of the quintuple innervation of a crustacean muscle. *Journal of Experimental Biology*, London, 16: 121-133.
- HAY, W. P., and C. A. SHORE.
1918. The decapod crustaceans of Beaufort, N. C., and the surrounding region. *Bulletin of the Bureau of Fisheries*, 35: 371-475.
- HEATH, J. P.
1941. The nervous system of the kelp crab, *Pugettia producta*. *Journal of Morphology*, 69: 481-500.
- HEDGPETH, J. W.
1947. Interesting crustaceans about which little has been written. *Progressive Fish Culturist*, October 1947, pp. 181-184.
- HEERGAARD, P.
1953. Observations of spawning and larval history of the shrimp *Penaeus setiferus* (L.). *Publications of the Institute of Marine Science, University of Texas*, 3: 73-105.
- HELDT, J. H.
1931. Observations sur la ponte, la fécondation et les premiers stades du développement de l'oeuf chez *Penaeus caramote* Risso. *Comptes Rendus Hebdomadaires des Séances de L'Académie des Sciences*, Paris, 193: 1039-1041.
1932a. Sur quelques différences sexuelles (coloration, taille, rostre) chez deux crevettes tunisiennes: *Penaeus caramote* Risso et *Parapenaeus longirostris* Lucas. *Bulletin, Station Océanographique de Salammbô, Tunisia*, 27: 1-20.
1932b. L'appareil génital mâle des crevettes nordafricaines de la famille des Penaeidae. *Comptes Rendus Hebdomadaires des Séances de L'Académie des Sciences*, Paris, 195: 1325-1327.
1938. La reproduction chez les Crustacés Décapodes de la famille des Pénéides. *Annales de l'Institut Océanographique*, Paris, 18: 31-206.
1948. Note sur des caracteres sexuels secondaire périodiques chez deux crevettes: *Processa canaliculata* Leach et *Periclemenes amethysteus* Risso. *Bulletin de la Société des Sciences Naturelles du Tunisie*, Fasc. 1. pp. 27-29.
- HEMMING, F.
1952. Proposed correction of an erroneous entry on the "Official List of Generic Names in Zoology" relating to the name *Penaeus* Weber, 1795 (Class Crustacea, Order Decapoda) (corrections of an entry in Opinion 104). *Bulletin of Zoological Nomenclature*, 6: 318-319.

HENRY L. M.

- 1947a. The nervous system and the segmentation of the head in the Annulata. *Microentomology*, 12 (3) : 65-82.
- 1947b. The nervous system and the segmentation of the head in the Annulata. Section III, Polychaeta. *Microentomology*, 12 (4) : 83-110.

HERDMAN, W. A.

1904. An outline of the shrimp question. *Proceedings and Transactions of the Liverpool Biological Society*, 18 : 157-167.

HERRICK, F. H.

1895. The American lobster: A study of its habits and development. *Bulletin of the U. S. Fish Commission*, 5 : 1-252.
1909. Natural history of the American lobster. *Bulletin of the U. S. Bureau of Fisheries*, 29 (747), 408 pp.

HICKMAN, V. V.

1937. The embryology of the Syncarid crustacean, *Anaspides tasmaniae*. *Papers and Proceedings of the Royal Society of Tasmania*, for 1936, pp. 1-36.

HOLMES, S. J.

1901. Habits of *Amphithoe longimana* Smith. *Biological Bulletin*, 2 : 165-193.

HOLMES, S. J., and E. S. HOMUTH.

1910. The seat of smell in the crayfish. *Biological Bulletin*, 18 : 155-160.

HOLMES, W.

1942. The giant myelinated nerve fibers of the prawn. *Philosophical Transactions of the Royal Society of London, Series B, Biological Sciences*, 213B : 293-311.
1943. Innervation of crustacean muscles. *Nature*, London, 151 : 531-532.

IHLE, J. E. W., and M. E. IHLE-LANDENBERG.

1939. Die Segmentierung des Crustaceen-Kopfes und der Cephalothorax der Dekapoden. *Bijdragen Tot De Dierkunde*, Amsterdam, 27 : 27-30.

IKEMATSU, W.

1953. On the life history of *Acetes japonicus* Kishinouye, in Ariake Sea. *Bulletin of the Japanese Society of Scientific Fisheries*, 19 (6) : 771-780.
1955. On the life history of *Metapenaeus joyneri* (Miers) in Ariake Sea. *Bulletin of the Japanese Society of Scientific Fisheries*, 20 (11) : 969.

INGRAND, M.

1937. Morphologie des pinces et caracteres sexuels secondaires de *Munida bamffica*. *Travaux de la Station Biologique de Roscoff, Nouvelle Serie*, Roscoff, France, 15 : 57-86.

JANCARIK, A.

1951. Antenní chemoreceptory lasturnatky *Heterocypris incongruens* (Ostracoda). *Sbornik Klubu Prirodovedeckeho V Brné, Brno, Czechoslovakia*, 29 (1) : 21-28.

KISHINOUE, K.

1900. Japanese species of the Genus *Penaeus*. *Journal of the Fishery Bureau, Tokyo*, 8 (1) : 1-29.
1928. On the branchial organs of the Sergestidae. *Proceedings of the Imperial Academy of Tokyo*, 4 : 489-490.

KISHINOUE, K.—Continued

1929. Penaeid crustaceans with the asymmetrical petasma. *Proceedings of the Imperial Academy of Tokyo*, 5 : 280-283.

KORSCHULT, E., and K. HEIDER.

- 1890-1893. *Lehrbuch der vergleichenden Entwicklungsgeschichte der wirbellosen Thiere*. 8vo. Jena.

KREIDL, A.

1893. Weitere Beiträge zur Physiologie des Orlabyrinthes. II. Mittheilung. Versuche an Krebsen. *Sitzungsberichte der Kaiserliche Akademie der Wissenschaften, Wien, Abt. 3*, 101 : 469-489; 102 : 149-174.

KUBO, I.

1955. On mass-growth of the prawn, *Penaeus japonicus* Bate. *Bulletin of the Japanese Society of Scientific Fisheries, Tokyo*, 20 (10) : 870-872.

LANG, D., and C. M. YONGE.

1935. The function of the tegumental glands in the statocyst of *Homarus vulgaris*. *Journal of the Marine Biological Association, United Kingdom*, 20 : 333-339.

LAUBMANN, A. L.

1912. Untersuchungen über die Hautsinnesorgane bei dekapoden Krebsen auf der Gruppe der Carididen. *Zoologische Jahrbücher, Abt. für Anatomie, Ontogenie*, 35 : 105-159.

LEBOUR, M. V.

1951. Notes on some larval decapods (Crustacea) from Bermuda. II. *Proceedings of the Zoological Society of London*, 120 : 743-747.

LEONE, C. A., and C. W. PRYOR.

1952. Serological correspondence among three species of penaeid Crustacea. *Journal of the Elisha Mitchell Science Society*, 68 : 27-31.

LERMA, B.

1951. Note originali e critiche sulla morfologia comparata degli organi frontali degli Artropodi. *Annuario dell'Istituto e Museo di Zoologia dell'Universita di Napoli*, 3 (1) : 1-25.

LOWNDES, A. G.

1937. Body orientation in the Crustacea. *Nature*, London, 140 : 241-242.

McRAE, E. D.

1952. The shrimp fishery. *Bulletin of the Texas Game and Fishery Commission*, 32 : 1-21.

MENON, M. K.

1952. The life history and bionomics of an Indian penaeid prawn *Metapenaeus dobsoni* Miers. *Proceedings of the Indo-Pacific Fishery Council for 1951*, 3 : 80-93.
1937. Decapod larvae from the Madras plankton. *Bulletin of the Madras Government Museum, New Series*, 3 (5) : 1-55.

MENZEL, R. W.

1955. Marking of shrimp. *Science*, 121 : 446.

MEREDITH, S. S.

1952. A study of *Crangon vulgaris* in the Liverpool Bay area. *Proceedings and Transactions of the Liverpool Biological Society*, 58 : 75-109.

- LOCQUARD, M. F.
1883. Recherches anatomiques sur l'estomac des Crustacés podophthalmiques. Annales des Sciences Naturelles, Zoologie et Biologie Animale, 11^e Série, 16 (6) : 1-311.
- OLLITOR, A.
1937. Beiträge zur Untersuchung des Exkretstoffwechsels und der Exkretion von *Eriocheir sinensis* H. Milne-Edwards. Zoologische Jahrbücher, Jena, Allgemeine Zoologie, 57 : 323-354.
- ONTICELLI, F. S., and S. LOBIANCO.
1900. Sullo sviluppo dei Peneidi del Golfo di Napoli. Monitore Zoologico Italiano, Florence, Suppl., 11 : 23.
1901. Uova e larva di *Solenocera siphonocera* Phil. Monitore Zoologico Italiano, Volume 12.
1901. Sui Peneidi del Golfo di Napoli. Monitore Zoologico Italiano, 12 : 798.
- ORRIS, M. C., and I. BENNET.
1952. The life history of a penaeid prawn (*Metapenaeus*) breeding in a coastal lake (Tuggerah, New South Wales). Proceedings of the Linnean Society of New South Wales, Sydney, 76 (5-6) : 164-182.
- OUCHET, S.
1932. Sur l'appareil génétal mâle de *Penaeus trisulcatus* Leach. Bulletin de la Société Zoologique de France, 57 : 316-340.
- ÜLLER, F.
1878. Ueber die Naupliusbrut der Garneelen. Zeitschrift für Wissenschaftliche Zoologie, Leipzig, 30 : 163-166.
- HSIMA, Y., and D. YASUDA.
1942. An ecological note on the prawn, *Penaeopsis affinis* (Milne-Edwards). Bulletin of the Japanese Society of Scientific Fisheries, 11 : 135-145.
- KADA, Y. K.
1928. Note on the tail organs of *Acetes*. Annals and Magazine of Natural History, London, 1 (10) : 308-310.
- ANNING, A.
1926. Über die Statocyste des *Astacus fluviatilis* L. (*Potamobius astacus* Leach). Mitteilungen auf dem Zoologischen Staatsinstitut und Zoologischen Museum in Hamburg, 42 : 118-125.
- ATWARDHAN, S. S.
1935. On the structure and mechanism of the gastric mill in Decapoda. VI. The structure of the gastric mill in Natantous Macrura—Penaeidae and Stenopidea, conclusion. Proceedings of the Indian Academy of Sciences, Section B, Bangalore, 2 : 155-174.
- ARSON, J. C.
1935. Eggs of a penaeid shrimp. Science, Vol. 82, No. 2121.
- REZ, C.
1929. Sur quelques caractères sexuels des Crustacés Décapodes. International Congress of Zoology, Budapest, 10 : 1142-1146.
- ENTISS, C. W.
1901. The otocyst of decapod Crustacea; its structure, development, and functions. Bulletin of the Museum of Comparative Zoology, 38 (7) : 167-251.
- PRESSWALLA, M. J.
1933. The nervous system of *Panulirus*. Current Science, Bangalore, Vol. 2.
- RAMADAN, M. M.
1938. Crustacea: Penaeidae. The John Murray Expedition, 1933-34, Scientific Reports, British Museum (Natural History), 5 (5) : 123-245.
- REDDY, A. R.
1937. The physiology of digestion and absorption in the crab *Paratelphusa* (*Oziotelphusa*) *hydromus* (Herbst). Proceedings of the Indian Academy of Sciences, 6B : 170-193.
- REYNOLDS, J. M.
1937. A further contribution to the problem of segmentation in the Arthropods. American Journal of Science, New Haven, 34 : 30-63.
- SAMUEL, N.
1945. Some aspects of the morphology and physiology of the antennary glands of three decapod crustaceans. Journal of the University of Bombay, Science Number, Section B: Biological Sciences Including Medicine, Bombay, N. S., 14B (3) : 29-41.
- SCHÄFER, W.
1954. Form und Funktion der Brachyuren-Schere. Abhandlungen der Senckenbergischen Naturforschenden Gesellschaft, Frankfurt am Main, 489 : 1-65.
- SCHLIEPER, C.
1933. Weitere Untersuchungen über die Beziehungen zwischen Bau und Funktion bei den Exkretionsorganen Dekapoden Crustaceen. Zeitschrift für Vergleichende Physiologie, 20 : 255-257.
- SCHMITT, WALDO L.
1934. The Macruran, Anomuran and Stomatopod Crustacea. Bijdragen tot de Kennis der Fauna von Curacao. Bijdragen Tot de Dierkunde, Amsterdam, Vol. 23.
1935. The West American species of the shrimps of the Genus *Penaeus*. Proceedings of the Biological Society of Washington, Vol. 48.
- SCHÖNE, H.
1951. Die Arbeitsweise der Statocyste von *Astacus fluviatilis*. Die Naturwissenschaften, 38 (7) : 157-158.
- SCHORSTEIN, M. V.
1941. The sphincter valve of the antennal gland of *Marinogammarus marinus* (Leach) sens. str. Proceedings of the Royal Society of Edinburgh, Section B, Biology, 61B : 130-137.
1943. The larval maxillary glands of *Euphausia superba* Dana. Proceedings of the Royal Society of Edinburgh, Section B, Biology, 61B : 375-384.
- SEIFERT, R.
1930. Sinnesphysiologische Untersuchungen am Keimfuss (*Triops cancriformis* Bosc.). Zeitschrift für Wissenschaftliche Biologie, Abt. C. Zeitschrift für Vergleichende Physiologie, 11 : 386-436.
- SENOO, HIDEKI.
1910. Growth rate of *Penaeus*. Magazine of Zoology, Tokyo, 22 : 91-93.

SHEN, C. J.

1934. A comparative study of the statocysts of Eumalacostraca, with special reference to the Macrura. Proceedings of the Zoological Society of London, 104: 533-557.

SJOGREN, S.

1934. Die Blütdrüse und ihre Ausbildung bei den Dekapoden. Zoologische Jahrbücher, Jena, 58: 145-170.

SORDI, M.

1941. L'apparato digerente degli eufausiacei. Atti della Società Italiana di Scienze Naturali e del Museo Civico di Storia Naturale in Milano, 80: 5-12.

SPAULDING, M. H.

1908. Preliminary report on the life history and habits of the "lake shrimp" (*Penaeus setiferus*). Bulletin No. 11, Gulf Biologic Station, Cameron, La., pp. 1-29.

TAYLOR, G. L.

1941. The optical properties of the shrimp nerve fiber sheath. Journal of Cellular and Comparative Physiology, Philadelphia, 18: 233-242.

TEISSIER, G.

1938. Un procédé pratique pour la conservation à sec des gros Crustacés. Bulletin de la Société Zoologique de France, 63: 152-154.

TISCHLER, W.

1935. Untersuchungen am Statozystemapparat bei *Sergestes edwardsii*. Zoologischer Anzeiger, Leipzig, 110: 14-16.

TONNER, F.

- 1933a. Das Problem der Krebschere. Zeitschrift für Vergleichende Physiologie, 19: 762-784.
1933b. Ein Beitrag zur Anatomie und Physiologie des peripheren Nervensystems von *Astacus fluviatilis*. Zoologische Jahrbücher, Allgemeine Zoologie, Jena, 53: 101-152.
1934. Ueber chemorezeptoren in der Kiemenhöhle des Flusskrebses. Zeitschrift für Zellforschung und Mikroskopische Anatomie, Berlin, 20: 423-426.

TULIAN, E. A.

1923. The present status of the Louisiana shrimp industry. Transactions of the American Fisheries Society, 53d Annual Meeting.

UCHIDA, T.

1930. Observations sur le sens de la reconnaissance du sexe chez *Asellus*. Bulletin de la Société Zoologique de France, 55: 516-520.

VIEIRA, B. B.

1947. Observações sobre a maturação de *Xiphopenaeus kroyeri* no litoral de São Paulo. Boletim do Museu Nacional, Zoologia, Rio de Janeiro, 74: 1-22.

VIOSCA, P.

1920. Report of the biologist. Fourth Biennial Report, Department of Conservation of Louisiana, pp. 120-130.
1953. Observations on the life history and habits of the lake shrimp. Louisiana Conservationist, 5 (4): 2-5, 18.

WELSH, J. H., and F. A. CHACE.

1938. Eyes of deep-sea crustaceans. II. Sergestidae. Biological Bulletin, 74: 364-375.

WEYMOUTH, F. W., M. J. LINDNER, and W. W. ANDERSON.

1932. Summary of the life history of the common shrimp (*Penaeus setiferus*) of the South Atlantic and Gulf Coasts of the United States. Transactions of the American Fisheries Society, Vol. 62.
1933. Preliminary report on the life history of the common shrimp *Penaeus setiferus* (Linn.). Bulletin of the Bureau of Fisheries, 48 (14): 1-26.

WIERSMA, C. A. G.

1949. The innervation of the legs of the coconut crab, *Birgus latro* L. Physiologia Comparata et Oecologia, 1: 68-75.
1951. On the innervation of the muscles in the leg of the lobster, *Homarus vulgaris* Milne-Edwards. Archives Néerlandaises de Zoologie, 8: 384-392.

WILKS, R. A. C.

1938. A new decalcification fluid. Nature, London, 142: 958-959.

WILLIAMSON, D. I.

1951. Studies in the biology of Talitridae (Crustacea, Amphipoda): visual orientation in *Talitrus saltator*. Journal of the Marine Biological Association, United Kingdom, 30: 91-99.

WILLIAMS, AUSTIN B.

1953. Identification of juvenile shrimp (Penaeidae) in North Carolina. Journal of the Elisha Mitchell Scientific Society, 69 (2): 156-160.
1955. A contribution to the life histories of commercial shrimps (Penaeidae) in North Carolina. Bulletin of Marine Science of the Gulf and Caribbean, University of Miami, 5 (2): 116-146.

YOKOYA, Y.

1941. On the classification of Penaeid shrimps by the structural features of the appendix masculina. Journal of the College of Agriculture, Tokyo Imperial University, 15 (1): 45-68.

YONGE, C. M.

1932. On the nature and permeability of chitin. I. The chitin lining of the foregut of decapod Crustacea and the function of the tegumental glands. Proceedings of the Royal Society of London, Series B, 111: 298-329.

YOSHIDA, H.

1949. On the life history of *Acetes japonicus* Kishinouye. Journal of the Shimonoseki College of Fisheries, 1: 51-55. (In Japanese with English summary)
1949. On the life history of the Korean shrimp, *Penaeus orientalis* Kishinouye. Bulletin of the Japanese Society of Scientific Fisheries, 15: 245-248. (In Japanese with English summary.)

YU, S. C.

1936. Contribution à l'étude anatomique et histologique d'une crevette chinoise (*Palaeomon asperulus* Mart). Chinese Journal of Zoology, 2: 101-132.

ZILCH, A.

1936. Zur Frage des Flimmerepithels bei Arthropoden. Zeitschrift für Wissenschaftliche Zoologie, Leipzig, 148: 89-132.